

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
21 November 2002 (21.11.2002)

PCT

(10) International Publication Number
WO 02/092015 A2

(51) International Patent Classification⁷: **A61K**

(21) International Application Number: **PCT/US02/15982**

(22) International Filing Date: **17 May 2002 (17.05.2002)**

(25) Filing Language: **English**

(26) Publication Language: **English**

(30) Priority Data:

60/291,311	17 May 2001 (17.05.2001)	US
60/353,058	1 February 2002 (01.02.2002)	US
60/361,293	4 March 2002 (04.03.2002)	US

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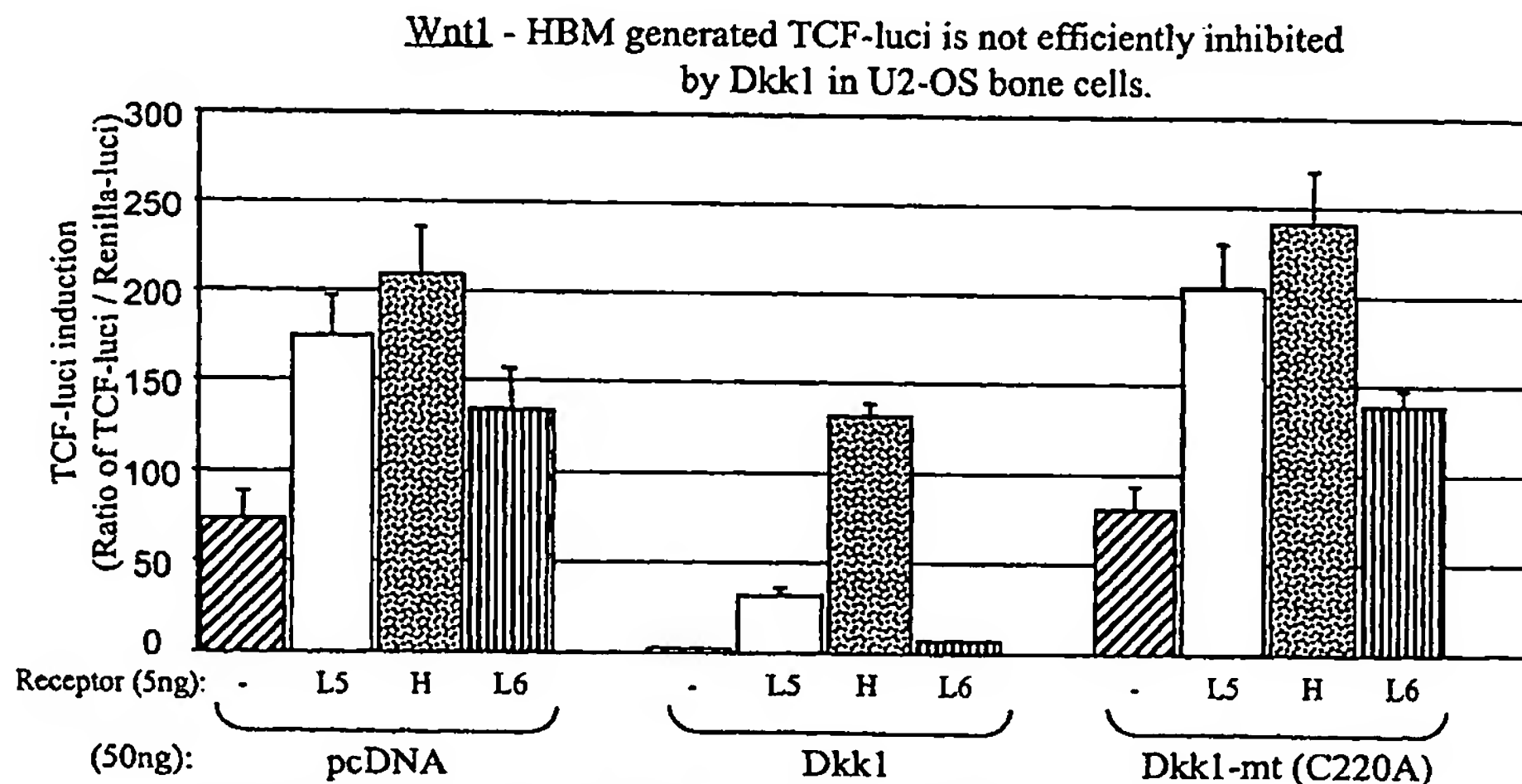
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(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW),

[Continued on next page]

(54) Title: REAGENTS AND METHODS FOR MODULATING DKK-MEDIATED INTERACTIONS



- With Wnt1 the TCF-signal generated by LRP5 is greater than that of LRP6.
- LRP5/6 -Wnt1 induced TCF- is efficiently blocked by Dkk1

(57) Abstract: The present invention provides reagents, compounds, compositions, and methods relating to novel interactions of the extracellular domain of LRP5, HBM (a variant of LRP5), and/or LRP6 with Dkk, including Dkk-1. The various nucleic acids, polypeptides, antibodies, assay methods, diagnostic methods, and methods of treatment of the present invention are related to and impact on Dkk, LRP5, LRP6, HBM, and Wnt signaling. Dkk, LRP5, LRP6, HBM, and Wnt are implicated in bone and lipid cellular signaling. Thus, the present invention provides reagents and methods for modulating lipid levels and/or bone mass and is useful in the treatment and diagnosis of abnormal lipid levels and bone mass disorders, such as osteoporosis.



Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM),
European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR,
GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent
(BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR,
NE, SN, TD, TG).

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

Published:

- *without international search report and to be republished upon receipt of that report*

REAGENTS AND METHODS FOR MODULATING DKK-MEDIATED INTERACTIONS

FIELD OF THE INVENTION

The present invention relates to signal transduction, bone development, bone
5 loss disorders, modulation of lipid-related conditions, research reagents, methods of
screening drug leads, drug development, treatments for bone and/or lipid disorders,
screening and development of therapies, molecular, cellular, and animal models of
bone and/or lipid development and maintenance, which are mediated by Dkk,
including Dkk-1, and/or LRP5, LRP6, HBM or other members of the Wnt pathway.

BACKGROUND OF THE INVENTION

Two of the most common types of osteoporosis are postmenopausal and
senile osteoporosis. Osteoporosis affects both men and women, and, taken with
other abnormalities of bone, presents an ever-increasing health risk for an aging
15 population. The most common type of osteoporosis is that associated with
menopause. Most women lose between 20-60% of the bone mass in the trabecular
compartment of the bone within 3-6 years after the cessation of menses. This rapid
bone loss is generally associated with an increase of bone resorption and formation.
However, the resorptive cycle is more dominant and the result is a net loss of bone
20 mass. Osteoporosis is a common and serious disease among postmenopausal
women. There are an estimated 25 million women in the United States alone who
are afflicted with this disease. The results of osteoporosis are personally harmful,
and also account for a large economic loss due to its chronicity and the need for
extensive and long-term support (e.g., hospitalization and nursing home care) from
25 disease sequelae. This is especially true in elderly patients. Additionally, while
osteoporosis is generally not thought of as a life-threatening condition, a 20-30%
mortality rate is related to hip fractures in elderly women. A large percentage of this
mortality rate can be directly associated with postmenopausal osteoporosis.

The most vulnerable tissue in the bone to the effects of postmenopausal osteoporosis is the trabecular bone. This tissue is often referred to as spongy bone and is particularly concentrated near the ends of the bone, near the joints, and in the vertebrae of the spine. The trabecular tissue is characterized by small structures which inter-connect with each other as well as the more solid and dense cortical tissue which makes up the outer surface and central shaft of the bone. This criss-cross network of trabeculae gives lateral support to the outer cortical structure and is critical to the biomechanical strength of the overall structure. In postmenopausal osteoporosis, it is primarily the net resorption and loss of the trabeculae which lead to the failure and fracture of the bone. In light of the loss of the trabeculae in postmenopausal women, it is not surprising that the most common fractures are those associated with bones which are highly dependent on trabecular support, e.g., the vertebrae, the neck of the femur, and the forearm. Indeed, hip fracture, Colle's fractures, and vertebral crush fractures are indicative of postmenopausal osteoporosis. Osteoporosis affects cortical as well as trabecular bone. Alterations in endosteal bone resorption and Haversian remodeling with age affect cortical thickness and structural integrity contributing the increased risk for fracture.

One of the earliest generally accepted methods for treatment of postmenopausal osteoporosis was estrogen replacement therapy. Although this therapy frequently is successful, patient compliance is low, primarily due to the undesirable side-effects of chronic estrogen treatment. Frequently cited side-effects of estrogen replacement therapy include reinitiation of menses, bloating, depression, and, potentially, increased risk of breast or uterine cancer. In order to limit the known threat of uterine cancer in women who have not had a hysterectomy, a protocol of estrogen and progestin cyclic therapy is often employed. This protocol is similar to that used in birth control regimens, and often is not tolerated by women because of the side-effects characteristic of progestin. More recently, certain antiestrogens, originally developed for the treatment of breast cancer, have been shown in experimental models of postmenopausal osteoporosis to be efficacious. Among these agents is raloxifene (See, U.S. Patent No. 5,393,763; Black *et al.*, *J.*

Clin. Invest., 93:63-69 (1994); and Ettinger *et al.*, *JAMA* 282:637-45 (1999)). In addition, tamoxifen, a widely used clinical agent for treating breast cancer, has been shown to increase bone mineral density in post menopausal women suffering from breast cancer (Love *et al.*, *N. Engl. J. Med.*, 326:852-856 (1992)).

5 Another therapy for the treatment of postmenopausal osteoporosis is the use of calcitonin. Calcitonin is a naturally occurring peptide which inhibits bone resorption and has been approved for this use in many countries (Overgaard *et al.*, *Br. Med. J.*, 305:556-561 (1992)). The use of calcitonin has been somewhat limited, however. Its effects are very modest in increasing bone mineral density, and the
10 treatment is very expensive. Another therapy for the treatment of postmenopausal osteoporosis is the use of bisphosphonates. These compounds were originally developed for treating Paget's disease and malignant hypercalcemia. They have been shown to inhibit bone resorption. Alendronate, a bisphosphonate, has been approved for the treatment of postmenopausal osteoporosis. These agents may be
15 helpful in the treatment of osteoporosis, but these agents also have potential liabilities which include osteomalacia, extremely long half-life in bone (greater than 2 years), and possible "frozen bone syndrome," e.g., the cessation of normal bone remodeling.

20 Senile osteoporosis is similar to postmenopausal osteoporosis in that it is marked by the loss of bone mineral density and resulting increase in fracture rate, morbidity, and associated mortality. Generally, it occurs in later life, *i.e.*, after 70 years of age. Historically, senile osteoporosis has been more common in females, but with the advent of a more elderly male population, this disease is becoming a major factor in the health of both sexes. It is not clear what, if any, role hormones
25 such as testosterone or estrogen have in this disease, and its etiology remains obscure. Treatment of this disease has not been very satisfactory. Hormone therapy, estrogen in women and testosterone in men, has shown equivocal results; calcitonin and bisphosphonates may be of some utility.

30 The peak mass of the skeleton at maturity is largely under genetic control. Twin studies have shown that the variance in bone mass between adult monozygotic

twins is smaller than between dizygotic twins (Slemenda *et al.*, *J. Bone Miner. Res.*, 6: 561-567 (1991); Young *et al.*, *J. Bone Miner. Res.*, 6:561-567 (1995); Pocock *et al.*, *J. Clin. Invest.*, 80:706-710 (1987); Kelly *et al.*, *J. Bone Miner. Res.*, 8:11-17 (1993)). It has been estimated that up to 60% or more of the variance in skeletal mass is inherited (Krall *et al.*, *J. Bone Miner. Res.*, 10:S367 (1993)). Peak skeletal mass is the most powerful determinant of bone mass in elderly years (Hui *et al.*, *Ann. Int. Med.*, 111:355-361 (1989)), even though the rate of age-related bone loss in adult and later life is also a strong determinant (Hui *et al.*, *Osteoporosis Int.*, 1:30-34 (1995)). Since bone mass is the principal measurable determinant of fracture risk, the inherited peak skeletal mass achieved at maturity is an important determinant of an individual's risk of fracture later in life. Thus, study of the genetic basis of bone mass is of considerable interest in the etiology of fractures due to osteoporosis.

Recently, a strong interest in the genetic control of peak bone mass has developed in the field of osteoporosis. The interest has focused mainly on candidate genes with suitable polymorphisms to test for association with variation in bone mass within the normal range, or has focused on examination of genes and gene loci associated with low bone mass in the range found in patients with osteoporosis. The vitamin D receptor locus (VDR) (Morrison *et al.*, *Nature*, 367:284-287 (1994)), PTH gene (Howard *et al.*, *J. Clin. Endocrinol. Metab.*, 80:2800-2805 (1995); Johnson *et al.*, *J. Bone Miner. Res.*, 8:11-17 (1995); Gong *et al.*, *J. Bone Miner. Res.*, 10:S462 (1995)) and the estrogen receptor gene (Hosoi *et al.*, *J. Bone Miner. Res.*, 10:S170 (1995); Morrison *et al.*, *Nature*, 367:284-287 (1994)) have figured most prominently in this work. These studies are difficult because bone mass (*i.e.*, the phenotype) is a continuous, quantitative, polygenic trait, and is confounded by environmental factors such as nutrition, co-morbid disease, age, physical activity, and other factors. Also, this type of study design requires large numbers of subjects. In particular, the results of VDR studies to date have been confusing and contradictory (Garnero *et al.*, *J. Bone Miner. Res.*, 10:1283-1288 (1995); Eisman *et al.*, *J. Bone. Miner. Res.*, 10:1289-1293 (1995); Peacock, *J. Bone Miner. Res.*, 10:1294-1297 (1995)).

Furthermore, thus far, the art has not determined the mechanism(s) whereby the genetic influences exert their effect on bone mass.

While it is well known that peak bone mass is largely determined by genetic rather than environmental factors, studies to determine the gene loci (and ultimately the genes) linked to variation in bone mass are difficult and expensive. Study designs which utilize the power of linkage analysis, e.g., sib-pair or extended family, are generally more informative than simple association studies, although the latter do have value. However, genetic linkage studies involving bone mass are hampered by two major problems. The first problem is the phenotype, as discussed briefly above. Bone mass is a continuous, quantitative trait, and establishing a discrete phenotype is difficult. Each anatomical site for measurement may be influenced by several genes, many of which may be different from site to site. The second problem is the age component of the phenotype. By the time an individual can be identified as having low bone mass, there is a high probability that their parents or other members of prior generations will be deceased and therefore unavailable for study, and younger generations may not have even reached peak bone mass, making their phenotyping uncertain for genetic analysis.

Thus, there is a need in the art for additional research tools for the elucidation of the molecular mechanism of bone modulation, for the screening and development of candidate drugs, and for treatments of bone development and bone loss disorders. The present invention is directed to these, as well as other, important ends.

In addition to bone modulation, the present invention relates to modulation of lipid levels. Cardiovascular disease is the most common cause of mortality in the United States, and atherosclerosis is the major cause of heart disease and stroke. It is widely appreciated that cholesterol plays an important role in atherogenesis. Normally, most cholesterol serves as a structural element in the walls of cells, whereas much of the rest is in transit through the blood or functions as the starting material for the synthesis of bile acids in the liver, steroid hormones in endocrine cells and vitamin D in skin. The transport of cholesterol and other lipids through the

circulatory system is facilitated by their packaging into lipoprotein carriers. These spherical particles comprise protein and phospholipid shells surrounding a core of neutral lipid, including unesterified ("free") or esterified cholesterol and triglycerides. Risk for atherosclerosis increases with increasing concentrations of low density lipoprotein (LDL) cholesterol, whereas risk is inversely proportional to levels of high-density lipoprotein (HDL) cholesterol. The receptor-mediated control of plasma LDL levels has been well-defined, and recent studies have now provided new insights into HDL metabolism.

The elucidation of LDL metabolism began in 1974 by Michael Brown and Joseph Goldstein. In brief, the liver synthesizes a precursor lipoprotein (very low density lipoprotein, VLDL) that is converted during circulation to intermediate density lipoprotein (IDL) and then to LDL. The majority of the LDL receptors expressed in the body are on the surfaces of liver cells, although virtually all other tissues ("peripheral tissues") express some LDL receptors. After binding, the receptor-lipoprotein complex is internalized by the cells via coated pits and vesicles, and the entire LDL particle is delivered to lysosomes, wherein it is disassembled by enzymatic hydrolysis, releasing cholesterol for subsequent cellular metabolism. This whole-particle uptake pathway is called "receptor-mediated endocytosis." Cholesterol-mediated feedback regulation of both the levels of LDL receptors and cellular cholesterol biosynthesis help ensure cellular cholesterol homeostasis. Genetic defects in the LDL receptor in humans results in familial hypercholesterolemia, a disease characterized by elevated plasma LDL cholesterol and premature atherosclerosis and heart attacks. One hypothesis for the deleterious effects of excess plasma LDL cholesterol is that LDL enters the artery wall, is chemically modified, and then is recognized by a special class of receptors called macrophage scavenger receptors, that mediate the cellular accumulation of the LDL cholesterol in the artery, eventually leading to the formation of an atherosclerotic lesion.

The major lipoprotein classes include intestinally derived chylomicrons that transport dietary fats and cholesterol, hepatic-derived VLDL, IDL, and LDL that can be atherogenic, and hepatic- and intestinally-derived HDL that are antiatherogenic.

Apoprotein B (ApoB) is necessary for the secretion of chylomicrons (ApoB48) and VLDL, IDL, and LDL (ApoB100). Plasma levels of VLDL triglycerides are determined mainly by the rates of secretion in LDL lipolytic activity. Plasma levels of LDL cholesterol are determined mainly by the secretion of ApoB100 into plasma, the efficacy with which VLDL are converted to LDL and by LDL receptor-mediated clearance. Regulation of HDL cholesterol levels is complex and is affected by rates of synthesis of its Apo proteins, rates of esterification of free cholesterol to cholesterol ester by LCAT, levels of triglyceride-rich lipoproteins and CETP-mediated transfer of cholesterol esters from HDL, and clearance from plasma of HDL lipids and Apo proteins.

Normal lipoprotein transport is associated with low levels of triglycerides and LDL cholesterol and high levels of HDL cholesterol. When lipoprotein transport is abnormal, lipoprotein levels can change in ways that predispose individuals to atherosclerosis and arteriosclerosis (see Ginsburg, *Endocrinol. Metab. Clin. North Am.*, 27:503-19 (1998)).

Several lipoprotein receptors may be involved in cellular lipid uptake. These receptors include: scavenger receptors; LDL receptor-related protein/ α 2-macroglobulin receptor (LRP); LDL receptor; and VLDL receptor. With the exception of the LDL receptor, all of these receptors are expressed in atherosclerotic lesions while scavenger receptors are mostly expressed in macrophages, the LRP and VLDL receptors may play an important role in mediating lipid uptake in smooth muscle cells (Hiltunen *et al.*, *Atherosclerosis*, 137 suppl.:S81-8 (1998)).

A major breakthrough in the pharmacologic treatment of hypercholesterolemia has been the development of the "statin" class of 3-hydroxy-3-methylglutaryl-CoA reductase (HMG CoA reductase) inhibitory drugs. 3-hydroxy-3-methylglutaryl-CoA reductase is the rate controlling enzyme in cholesterol biosynthesis, and its inhibition in the liver stimulates LDL receptor expression. As a consequence, both plasma LDL cholesterol levels and the risk for atherosclerosis decrease. The discovery and analysis of the LDL receptor system has had a profound impact on cell biology, physiology, and medicine.

HDL is thought to remove unesterified, or "free" cholesterol (FC) from peripheral tissues, after which most of the cholesterol is converted to cholesterol ester (CE) by enzymes in the plasma. Subsequently, HDL cholesterol is efficiently delivered directly to the liver and steroidogenic tissues via a selective uptake pathway and the HDL receptor, SR-BI (class B type I scavenger receptor) or, in some species, transferred to other lipoproteins for additional transport in metabolism (see Krieger, *Proc. Natl. Acad. Sci. USA*, 95:4077-4080 (1998)).

These issues illustrate a need in the art for additional research tools for the elucidation of the molecular mechanism of lipid modulation, for the screening and development of candidate drugs, and for treatments of lipid levels and lipid level modulation disorders. The present invention is directed to these, as well as other, important ends.

SUMMARY OF THE INVENTION

The present invention provides reagents, compounds, compositions and methods relating to novel interactions of the extracellular domain of LRP5, HBM (a variant of LRP5), and/or LRP6 with Dkk proteins. LRP5 is also referred to as Zmax1 or Zmax. Thus, when discussing methods, reagents, compounds, and compositions of the invention which relate to the interaction between Dkk and LRP5 (or Zmax1), the invention is also to be understood to encompass embodiments relating to interactions between Dkk and LRP6 and Dkk and HBM. Moreover, where Dkk is discussed herein, it is to be understood that the methods, reagents, compounds, and compositions of the present invention include the Dkk family members, including but not limited to Dkk-1, Dkk-2, Dkk-3, Dkk-4 and Soggy. Furthermore, the invention encompasses novel fragments of Dkk-1 which demonstrate a binding interaction between the ligand binding domain (LBD) of LRP5 and additional proteins and/or which can modulate an interaction between LRP5, or a variant or fragment thereof, and a Dkk protein. The invention provides assays, methods, compositions, and compounds relating to Dkk-Wnt signaling. Numerous Wnt proteins are compatible with the present invention, including Wnt1-Wnt19, and particularly, Wnt1, Wnt3,

Wnt3a, and Wnt10b. The present invention further provides reagents, compounds, compositions and methods modulating interactions between one or more other proteins and Dkk-1. The present invention also provides a series of peptide aptamers which bind to Dkk-1 or to LRP5 (or HBM and/or LRP6).

5 The polypeptides of the invention, for example in the form of peptide oligomers, aptamers, proteins, and protein fragments as well as the nucleic acids of the invention, for example in the form of nucleic acids which encode the polypeptides of the invention as well as antisense, or complimentary nucleic acids, are useful as reagents for the study of bone mass and lipid level modulation. The polypeptides
10 and nucleic acids of the invention are also useful as therapeutic and diagnostic agents.

 The present invention provides useful reagents for the modulation of Dkk proteins with LRP5, LRP6, and/or HBM, the modulation Dkk-1 and/or Dkk-1 interacting protein activity, and modulation of LRP5/Dkk-1, LRP6/Dkk1 and
15 HBM/Dkk-1 interactions and Dkk-1/Dkk-1 interacting protein interactions. The present invention provides a series of peptide aptamers which bind Dkk-1 or LRP5, LRP6, and/or HBM.

 An object of the invention is to provide for a method of regulating LRP5/LRP6/HBM/HBM-like activity in a subject comprising administering a
20 therapeutically effective amount of a composition which modulates Dkk activity. The subject can be a vertebrate or an invertebrate organism, but more preferably the organism is a canine, a feline, an ovine, a primate, an equine, a porcine, a caprine, a camelid, an avian, a bovine, or a rodent organism. A more preferred organism is a human. In a preferred embodiment, the Dkk protein is Dkk-1. In a particularly
25 preferred embodiment, Dkk-1 activity is decreased. In another embodiment, Dkk activity modulates bone mass and/or lipid levels. In a preferred embodiment, bone mass is increased and/or lipid levels are decreased. In another preferred embodiment, the modulation in bone mass is an increase in bone strength determined via one or more of a decrease in fracture rate, an increase in areal bone
30 density, an increase in volumetric mineral bone density, an increase in trabecular

connectivity, an increase in trabecular density, an increase in cortical density or thickness, an increase in bone diameter, and an increase in inorganic bone content. The invention further provides such a method wherein the composition comprises a Dkk, Dkk-1 or a LRP5/LRP6/HBM binding fragment thereof, such as those depicted
5 in Figure 6 or a mimetic of those fragments depicted in Figure 6. The invention further provides such a method wherein the composition comprises one or more of the proteins which interact with Dkk, including Dkk-1, such as those depicted in Figure 5, or a Dkk-binding fragment thereof, or an antisense, siRNA, or shRNA molecule which recognizes and binds to a nucleic acid encoding one or more Dkk
10 interacting or Dkk-1 interacting proteins. The invention further provides such a method wherein the composition comprises an LRP5/LRP6/Zmax1 antibody, Dkk antibody, a Dkk-1 antibody or an antibody to a Dkk-1 interacting protein. The invention further provides such a method wherein the compositions comprise an aptamer of Dkk or Dkk-1, such as those depicted in Figure 3 (SEQ ID NOs:171-188),
15 or a mimetic of such an aptamer. The method further provides that invention further provides such a method wherein the compositions comprise an aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer.

A composition of the present invention may modulate activity either by enhancing or inhibiting the binding of Dkk to LRP5/LRP6/Zmax1, particularly Dkk-1,
20 or the binding of Dkk-1 to a Dkk-1 interacting protein, such as those shown in Figure 5. A composition of the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figures 4 (SEQ ID NOs:189-192) (particularly, peptide (SEQ ID NO:191) and 13 (including SEQ IDNOs:204-214), or a mimetic of such an aptamer. Preferred compositions of the present invention also comprise
25 LRP5 antibodies.

Another aspect of the invention is to provide for a method of regulating Dkk-Wnt pathway activity in a subject comprising administering a therapeutically effective amount of a composition which modulates Dkk-Wnt pathway activity. In a preferred embodiment, the Dkk protein is Dkk-1. In a particularly preferred embodiment, Dkk-
30 1 activity is decreased. In another embodiment, Dkk activity modulates bone mass

and/or lipid levels. In a preferred embodiment, bone mass is increased and/or lipid levels are decreased. In another preferred embodiment, the modulation in bone mass is an increase in bone strength determined via one or more of a decrease in fracture rate, an increase in areal bone density, an increase in volumetric mineral bone density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density or thickness, an increase in bone diameter, and an increase in inorganic bone content. In another preferred embodiment, the Wnt is Wnt1-Wnt19. In a particularly preferred embodiment, the Wnt is Wnt1, Wnt3, Wnt3a, or Wnt10b. Preferred compositions comprise Dkk-modulating or Dkk-1-modulating compounds or one or more Dkk interacting or Dkk-1 interacting proteins, or a Dkk-binding fragment thereof. Other preferred Dkk modulating compositions comprise a Dkk or Dkk-1 antibody or an antibody to a Dkk interacting or Dkk-1 interacting protein. Also contemplated are antisense, siRNA, and shRNA molecules which recognize and bind to a nucleic acid encoding one or more Dkk-1 interacting proteins. The invention further provides such a method wherein the composition comprises a biologically active or LRP5/LRP6/HBM binding fragment of Dkk, including Dkk-1, such as those depicted in Figure 6 or a mimetic of those fragments depicted in Figure 6. The Dkk modulating composition may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. A composition of the present invention may modulate activity either by enhancing or inhibiting the binding of Dkk, including Dkk-1, to LRP5, LRP6, or HBM or the binding of Dkk, including Dkk-1, to a Dkk interacting protein, such as those shown in Figure 5. The invention further provides such a method wherein the composition comprises an aptamer of Dkk or Dkk-1, such as those depicted. A composition of the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208). Preferred compositions of the present invention also comprise LRP5 antibodies.

A further aspect of the invention is to provide for a method of modulating Wnt signaling in a subject comprising administering a therapeutically effective amount of a composition which modulates Dkk activity or modulates Dkk interaction with LRP5

(or LRP6 or HBM). In a preferred embodiment, the Dkk protein is Dkk-1. In a particularly preferred embodiment, Dkk-1 activity is decreased. In another embodiment, Dkk activity modulates bone mass and/or lipid levels. In a preferred embodiment, bone mass is increased and/or lipid levels are decreased. In another preferred embodiment, the modulation in bone mass is an increase in bone strength determined via one or more of a decrease in fracture rate, an increase in areal bone density, an increase in volumetric mineral bone density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density or thickness, an increase in bone diameter, and an increase in inorganic bone content.

In another preferred embodiment, the Wnt is Wnt1-Wnt19. In a particularly preferred embodiment, the Wnt is Wnt1, Wnt3, Wnt3a, or Wnt10b. Preferred Wnt modulating compositions comprise one or more Dkk interacting or Dkk-1 interacting proteins, or a biologically active or LRP5/LRP6/HBM binding fragment thereof. Also

contemplated are antisense, siRNA, and shRNA molecules which recognize and bind to a nucleic acid encoding one or more Dkk interacting or Dkk-1 interacting proteins. The invention further provides such a method wherein the composition comprises a biologically active or LRP5/LRP6/HBM binding fragment of Dkk or Dkk-1, such as those depicted in Figure 6 or a mimetic of those fragments depicted in Figure 6. The Dkk modulating composition may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. A composition of the present invention may modulate activity either by enhancing or blocking the binding of Dkk, including Dkk-1, to LRP5, LRP6, or HBM or the binding of Dkk or Dkk-1 to a Dkk interacting or Dkk-1 interacting protein, such as those shown in Figure 5. The invention further provides such a method wherein compositions comprising an aptamer of Dkk or Dkk-1, such as those depicted in Figure 3 (SEQ ID NOs:171-188), or a mimetic of such an aptamer. The invention further provides such a method wherein the composition comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk interacting or Dkk-1 interacting protein. The invention further provides such a method wherein compositions of an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figures 4 (SEQ ID NO:189-192

(particularly peptide (SEQ ID NO:191) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Additional preferred compositions of the present invention also comprise LRP5 antibodies.

Additionally, the invention provides for a method of modulating bone mass and/or lipid levels in a subject comprising administering to the subject a composition which modulates Dkk activity or Dkk interaction with LRP5 in an amount effective to modulate bone mass and/or lipid levels, wherein bone mass and/or lipid levels are in need of modulation. In a preferred embodiment, the Dkk protein is Dkk-1. In a particularly preferred embodiment, Dkk-1 activity is decreased. In another embodiment, Dkk activity modulates bone mass and/or lipid levels. In a preferred embodiment, bone mass is increased and/or lipid levels are decreased. In another preferred embodiment, the modulation in bone mass is an increase in bone strength determined via one or more of a decrease in fracture rate, an increase in areal bone density, an increase in volumetric mineral bone density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density or thickness, an increase in bone diameter, and an increase in inorganic bone content. Preferred bone mass and/or lipid modulating compositions comprise one or more Dkk interacting or Dkk-1 interacting proteins, or a biologically active or LRP5/LRP6/HBM binding fragment thereof. Also contemplated are antisense, siRNA, and shRNA molecules which recognize and bind to a nucleic acid encoding one or more Dkk interacting or Dkk-1 interacting proteins. The invention further provides such a method wherein the composition comprises a biologically active or LRP5/LRP6/HBM binding fragment of Dkk, including Dkk-1, such as those depicted in Figure 6 or a mimetic of those fragments depicted in Figure 6. The Dkk modulating composition may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. The invention further provides such a method wherein the composition comprises an aptamer of Dkk or Dkk-1, such as those depicted in Figure 3 (SEQ ID NOs:171-188), or a mimetic of such an aptamer. A composition of the present invention may modulate activity either by enhancing or inhibiting the binding of Dkk, including Dkk-1, to LRP5, LRP6,

or HBM or the binding of Dkk, including Dkk-1, to a Dkk interacting protein, such as those shown in Figure 5. The invention further provides such a method wherein the composition comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk interacting or Dkk-1 interacting protein. A composition of the present invention may comprise
5 an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figures 4 (SEQ ID NOs:189-192 (particularly peptide 13 (SEQ ID NO:191)) and 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Preferred compositions of the present invention also comprise LRP5 antibodies. It is a further aspect of the invention that such lipid-modulated diseases include a cardiac condition,
10 atherosclerosis, familial lipoprotein lipase deficiency, familial apoprotein CII deficiency, familial type 3 hyperlipoproteinemia, familial hypercholesterolemia, familial hypertriglyceridemia, multiple lipoprotein-type hyperlipidemia, elevated lipid levels due to dialysis and/or diabetes, and an elevated lipid level of unknown etiology.

15 Bone disorders contemplated for treatment and/or diagnosis by the methods and compositions disclosed herein include a bone development disorder, a bone fracture, age related loss of bone, a chondrodystrophy, a drug-induced bone disorder, high bone turnover, hypercalcemia, hyperostosis, osteogenesis imperfecta, osteomalacia, osteomyelitis, osteoporosis, Paget's disease, osteoarthritis, and
20 rickets.

It is a further object of the invention to provide a method of screening for compounds or compositions which modulates the interaction of Dkk with LRP5, LRP6, HBM, or a Dkk-binding fragment of LRP5, LRP6, or HBM comprising:

- 25 (a) exposing Dkk or a LRP5/LRP6/HBM binding fragment thereof to a compound; and
(b) determining whether said compound binds to Dkk or the LRP5/LRP6/HBM binding fragment thereof.

In a preferred embodiment, the Dkk is Dkk-1. In a particularly preferred embodiment, the binding of Dkk-1 to LRP5/LRP6/HBM is decreased.

It is a further object of the invention to provide a method of screening compounds or compositions which modulate the interaction of DKK with LRP5, LRP6, HBM, or a DKK-binding fragment thereof comprising:

- (a) exposing DKK or a LRP5/LRP6/HBM binding fragment thereof to a compound; and,
- (b) determining whether said compound modulates the interaction of Dkk with LRP5, LRP6, or HBM, or the Dkk-binding fragment of LRP5/LRP6/HBM.

In a preferred embodiment, the Dkk is Dkk-1. In a particularly preferred embodiment, the interaction of Dkk-1 with LRP5/LRP6/HBM is decreased.

It is a further object of the invention to provide a method of screening for compounds or compositions which modulates the interaction of Dkk with LRP5, LRP6, HBM, or a Dkk-binding fragment of LRP5, LRP6, or HBM comprising:

- (a) exposing Dkk or a LRP5/LRP6/HBM binding fragment thereof to a compound;
- (b) determining whether said compound binds to Dkk or the LRP5/LRP6/HBM binding fragment thereof; and,
- (c) further determining whether said compound modulates the interaction of Dkk with LRP5, LRP6, or HBM, or the Dkk-binding fragment of LRP5/LRP6/HBM.

In preferred embodiments of such methods, Dkk or a biologically active fragment thereof is attached to a solid substrate. In an alternative embodiment of the invention, LRP5/LRP6/HBM, or a biologically active fragment thereof (such as the ligand binding domain), is exposed to the compound. Another aspect of the invention provides for compounds and compositions identified by the disclosed methods. A preferred embodiment of the invention provides that the compound screened in an afore-mentioned method is one or more proteins which interact with Dkk, particularly Dkk-1, as depicted in Figure 5, or a LRP5/LRP6/HBM-binding fragment thereof. Another preferred embodiment provides that the compound comprises a Dkk or Dkk-1 peptide aptamer, such as those depicted in Figure 3 (SEQ

ID NOs:171-188), or a mimetic of such aptamers. The compound may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. The method further provides that the compound comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk-1 interacting protein.

5 The invention further provides that the compound may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191)) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Preferred compounds of the present invention also comprise LRP5 antibodies.

10 It is a further object of the invention to provide a method of screening for compounds or compositions which modulate the interaction of Dkk and a Dkk interacting protein comprising:

- (a) exposing a Dkk interacting proteins or a Dkk-binding fragment thereof to a compound; and,
- 15 (b) determining whether said compound binds to a Dkk interacting proteins or the Dkk-binding fragment thereof.

In a preferred embodiment, the Dkk is Dkk-1.

20 It is a further object of the invention to provide a method of screening for compounds or compositions which modulate the interaction of Dkk and a Dkk interacting protein comprising:

- (a) exposing Dkk interacting protein(s) or a Dkk-binding fragment thereof to a compounds; and,
- 25 (b) determining whether said compound modulates the interaction of Dkk and Dkk interacting proteins.

It is a further object of the invention to provide a method of screening for compounds or compositions which modulate the interaction of Dkk and a Dkk interacting protein comprising:

- 30 (a) exposing a Dkk interacting proteins or a Dkk-binding fragment thereof to a compound;

- (b) determining whether said compound binds to a Dkk interacting proteins or the Dkk-binding fragment thereof; and,
- (c) further determining whether said compound modulates the interaction of Dkk and Dkk interacting proteins.

In a preferred embodiment, Dkk is Dkk-1.

In preferred embodiments of such methods, the Dkk interacting proteins, particularly Dkk-1 interacting proteins, or a Dkk-binding fragment thereof are attached to a solid substrate. Another aspect of the invention provides for compounds and compositions identified by the disclosed methods. A preferred embodiment provides that the compound comprises a Dkk or Dkk-1 peptide aptamer, such as those depicted in Figure 3 (SEQ ID NOs:171-188), or a mimetic of such aptamers. The compound may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. The compound may also comprise an antibody to a Dkk interacting or Dkk-1 interacting protein.

It is another object of the invention to provide for a composition for treating bone mass disorders comprising a LRP5/LRP6/HBM modulating compound and a pharmaceutically acceptable excipient and/or carrier therefor. Preferred LRP5 (or LRP6 or HBM) modulating compounds include Dkk or Dkk-1 or a LRP5/LRP6/HBM binding fragment thereof. Also contemplated are compounds which comprise monoclonal or polyclonal antibodies or immunologically active fragments thereof which bind to Dkk, including Dkk-1, and a pharmaceutically acceptable excipient and/or carrier. Another preferred embodiment provides that the modulating compound comprises one or more Dkk interacting or Dkk-1 interacting proteins, or a biologically active fragment thereof. Also contemplated are compounds which comprise monoclonal or polyclonal antibodies or immunologically active fragments thereof which bind to Dkk interacting or Dkk-1 interacting proteins, or a biologically active fragment thereof, and a pharmaceutically acceptable excipient and/or carrier.

Another preferred embodiment provides that the modulating compound comprises an antisense, siRNA, and shRNA molecule which recognizes and binds to a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein. Another preferred embodiment provides that the modulating compound comprises a Dkk or Dkk-1 peptide aptamer, a mimetic of a Dkk or Dkk-1 peptide aptamer, a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. Another embodiment provides that the compound comprises an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Preferred compounds of the present invention also comprise LRP5 antibodies.

It is a further object of the invention to provide a pharmaceutical composition for treating a Dkk-mediated disease or condition comprising a compound which modulates Dkk activity and a carrier therefor, including pharmaceutically acceptable excipients. Such compositions include those wherein the compound comprises an antisense, siRNA, and shRNA molecule or an antibody which binds to Dkk, including Dkk-1, and thereby prevents it from interacting with LRP5, LRP6, or HBM. Other such compositions include one or more of Dkk interacting or Dkk-1 interacting proteins, such as those depicted in Figure 5, or a Dkk-binding fragment thereof, or a monoclonal or polyclonal antibody, or immunologically active fragment thereof, which binds to a Dkk interacting or Dkk-1 interacting protein or Dkk-binding fragment thereof. Other contemplated compositions include antisense, siRNA, and shRNA molecules which recognize and bind to a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein. Further contemplated compositions include Dkk and Dkk-1 peptide aptamers, such as those depicted in Figure 3 (SEQ ID NOs;171-188), mimetics of such aptamers, a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. Other contemplated compositions comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191))

and Figure 13 (including SEQ ID NO:204-214), or a mimetic of such an aptamer. Other preferred compositions of the present invention comprise LRP5 antibodies.

A further object of the invention to provide for a method of modulating the expression of a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein in an organism, such as those shown in Figure 5, comprising the step of administering to the organism an effective amount of composition which modulates the expression of a nucleic acid encoding a Dkk-1 interacting protein. In a preferred embodiment, said composition comprises an antisense, siRNA, or shRNA molecule which recognizes and binds to a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein.

One aspect of the invention provides for a method of modulating at least one activity of Dkk or a Dkk-1 interacting protein comprising administering an effective amount of a composition which modulates at least one activity of Dkk or a Dkk-1 interacting protein. The invention provides for a composition comprising a Dkk interacting or Dkk-1 interacting protein, such as those shown in Figure 5, or a biologically active fragment thereof. Other agents contemplated for this method are antisense, siRNA, or shRNA molecules which recognize and bind to a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein. The method further provides that the composition comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk interacting or Dkk-1 interacting protein. In another preferred embodiment, the composition comprises a Dkk or Dkk-1 peptide aptamer, a mimetic of a Dkk or Dkk-1 peptide aptamer, a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. The method provides that a composition of the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NO:189-192) (particularly peptide including (SEQ ID NO:191)) and Figure including (SEQ ID NOs:204-214), or a mimetic of such an aptamer. Preferred compositions of the present invention also comprise LRP5 antibodies. In a further preferred embodiment, the modulated Dkk activity is lipid modulation or bone mass modulation.

In all of the testing/screening embodiments of the present invention discussed below to obtain compounds or compositions which ultimately impact LRP5/LRP6/HBM signaling, one skilled in the art will recognize that HBM can be used as a control in the absence of a test sample or compound. Further, the effect of a test sample of compound on Wnt signaling through the interaction of Dkk with LRP5/LRP6/HBM does not necessarily require a direct measurement of an association or interaction of Dkk and LRP5/LRP6/HBM. Other positive phenotypes/activities established by the High Bone Mass phenotype or by using HBM as a control.

One aspect of the invention provides for a method of identifying binding partners for a Dkk protein comprising the steps of:

- (a) exposing the Dkk protein(s) or a LRP5/LRP6 binding fragment thereof to a potential binding partner; and
- (b) determining if the potential binding partner binds to a Dkk protein or the LRP5/LRP6 binding fragment thereof.

In a preferred embodiment, the Dkk is Dkk-1.

Another aspect of the invention is to provide for a method of identifying a compound that effects Dkk-mediated activity comprising

- (a) providing a group of transgenic animals having (1) a regulatable one or more Dkk interacting protein genes, (2) a knock-out of one or more Dkk interacting protein genes, or (3) a knock-in of one or more Dkk interacting protein genes;
- (b) providing a second group of control animals respectively for the group of transgenic animals in step (a); and
- (c) exposing the transgenic animal group and the control animal group to a potential Dkk-modulating compound which modulates bone mass or lipid levels; and

- (d) comparing the transgenic animal group and the control animal group and determining the effect of the compound on bone mass or lipid levels in the transgenic animals as compared to the control animals.

5

In a preferred embodiment, the Dkk is Dkk-1.

It is another aspect of the invention to provide for a method for determining whether a compound modulates a Dkk interacting protein, said method comprising the steps of:

- 10 (a) mixing the Dkk interacting protein or a Dkk-binding fragment thereof with the ligand binding domain of Dkk in the presence of said at least one compound;
- 15 (b) measuring the amount of said binding domain of Dkk bound to said Dkk interacting protein or the Dkk-binding fragment thereof as compared to a control without said at least one compound; and
- 20 (c) determining whether the compound reduces the amount of said binding domain of Dkk binding to said Dkk interacting protein or Dkk-binding fragment thereof.

In a preferred embodiment, the Dkk is Dkk-1.

In a preferred embodiment, the binding domain is attached to a solid substrate. The invention further provides for compounds identified by this method.

- 25 In a preferred embodiment, the invention provides that the Dkk interacting or Dkk-1 interacting protein is detected by antibodies. In another preferred embodiment, the solid substrate is a microarray. Another preferred embodiment provides that the ligand binding domain of Dkk and/or Dkk interacting protein is fused or conjugated to a peptide or protein. The invention also provides that the compounds include Dkk

and Dkk-1 peptide aptamers, mimetics of Dkk and Dkk-1 peptide aptamers, Dkk and Dkk-1 interacting proteins peptide aptamers, or mimetics of such aptamers.

An aspect of the invention provides a composition comprising one or more polypeptide sequences of one or more Dkk-1 interacting proteins, or a biologically active fragment thereof, one or more Dkk proteins, or a biologically active fragment thereof, or LRP5/LRP6/HBM polypeptide sequences or a biologically active fragment thereof (for example, the ligand binding domain) and a pharmaceutically acceptable excipient and/or carrier. Another aspect of the invention provides that the composition comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk interacting or Dkk-1 interacting protein and a pharmaceutically acceptable excipient. A composition of the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191)) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. A composition of the present invention may comprise a Dkk peptide aptamer, for example as shown in Figure 3 (SEQ ID NOs:171-188). Preferred compositions of the present invention also comprise LRP5 antibodies.

Another aspect of the invention is to provide an antibody or immunologically active antibody fragment which recognizes and binds to a Dkk-1 amino acid sequence selected from the group consisting of: Asn34-His266 (SEQ ID NO:110), Asn34-Cys245 (SEQ ID NO:111), Asn34-Lys182 (SEQ ID NO:112), Cys97-His266 (SEQ ID NO:113), Val139-His266 (SEQ ID NO:114), Gly183-His266 (SEQ ID NO:115), Cys97-Cys245 (SEQ ID NO:116), or Val139-Cys245 (SEQ ID NO:117) of human Dkk-1. Additional antibodies may bind to any of the sequences depicted in Figures 3 (SEQ ID NOs:171-188) and Figure 4 (SEQ ID NOs:189-192). Another aspect of the invention is to provide for polyclonal antibodies to one or more amino acid sequences: Peptide 1 -GNKYQTIDNYQPYPYPC (SEQ ID NO:118), Peptide 2 -LDGYSRRTTLSSKMYHTKGQEG (SEQ ID NO:119), Peptide 3 -RIQKDHQASNSSRLHTCQRH (SEQ ID NO:120), Peptide 4 -RGEIETITESFGND (SEQ ID NO:121), and Peptide 5 -EIFQRCYCGEGLSCRIQKD (SEQ ID NO: 122).

It is a further object of the invention to provide a nucleic acid encoding a Dkk protein, e.g. Dkk-1, a Dkk interacting or Dkk-1 interacting protein aptamer, or an LRP5 aptamer comprising a nucleic acid encoding a scaffold protein in-frame with the activation domain of Gal4 or LexA that is in-frame with a nucleic acid which
5 encodes for a Dkk or Dkk-1 or Dkk interacting or Dkk-1 interacting protein amino acid sequence. Preferably the scaffold protein is thioredoxin (trxA), S1 nuclease from *Staphylococcus* or M13. Other preferable embodiments include Dkk-1 amino acid sequences selected from Figure 6.

It is yet a further object of the invention to provide a composition comprising a
10 polypeptide sequence of Figure 3 (SEQ ID NOs:171-188), Figure 4 (SEQ ID NO:189-192), or of Dkk-1 interacting proteins identified in Figure 5 and a pharmaceutically acceptable excipient and/or carrier.

Another aspect of the invention includes a method of detecting the modulatory activity of a compound on the binding interaction of a first peptide and a second
15 peptide of a peptide binding pair that bind through extracellular interaction in their natural environment, comprising:

(i) culturing at least one eukaryotic cell, wherein the eukaryotic cell comprises;

a) a nucleotide sequence encoding a first heterologous
20 fusion protein comprising the first peptide or a segment thereof joined to a DNA binding domain of a transcriptional activation protein;

b) a nucleotide sequence encoding a second heterologous
25 fusion protein comprising the second peptide or a segment thereof joined to a transcriptional activation domain of a transcriptional activation protein;

wherein binding of the first peptide or segment thereof and the second peptide or segment thereof reconstitutes a transcriptional activation protein; and

c) a reporter element activated under positive transcriptional control of the reconstituted transcriptional activation protein, wherein expression of the reporter element produces a selected phenotype;

- 5 (ii) incubating a compound with the eukaryotic cell under conditions suitable to detect the selected phenotype; and
- (iii) detecting the ability of the compound to affect the binding interaction of the peptide binding pair by determining whether the compound affects the expression of the reporter element which produces the
- 10 selected phenotype;

wherein (1) said first peptide is a Dkk peptide and said second peptide is a peptide selected from LRP5, HBM, LRP6, and the Dkk-binding portion of LRP5/LRP6/HBM or (2) said first peptide is a Dkk-interacting protein or the Dkk-binding fragment thereof, and said second peptide is a Dkk peptide.

15 In one embodiment, the eukaryotic cell is a yeast cell. In a preferred embodiment, the yeast cell is *Saccharomyces*. In a particularly preferred embodiment, the *Saccharomyces* cell is *Saccharomyces cerevisiae*. The invention further provides that the compound may comprise a Dkk interacting or Dkk-1 interacting protein, or a biologically active fragment thereof. In one embodiment, the

20 Dkk interacting or Dkk-1 interacting protein, or a Dkk-binding fragment thereof, is added directly to the assay. In another embodiment, the Dkk interacting or Dkk-1 interacting protein, or a Dkk-binding fragment thereof, is recombinantly expressed by the eukaryotic cell in addition to the first and second peptides. In a preferred

25 embodiment the compound comprises a Dkk or Dkk-1 aptamer, a mimetic of a Dkk or Dkk-1 peptide aptamer, a Dkk interacting or Dkk-1 interacting protein aptamer, or a mimetic of a Dkk-1 interacting protein aptamer. Other preferred embodiments provide that the compound comprises an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an

30 aptamer. Alternatively, the present invention also provides that the compound may

comprise LRP5 antibodies or Dkk antibodies. In another embodiment, the yeast cell further comprises at least one endogenous nucleotide sequence selected from the group consisting of a nucleotide sequence encoding the DNA binding domain of a transcriptional activation protein, a nucleotide sequence encoding the transcriptional activation domain of a transcriptional activation protein, and a nucleotide sequence encoding the reporter element, wherein at least one of the endogenous nucleotide sequences is inactivated by mutation or deletion. In another embodiment, the peptide binding pair comprises a ligand and a receptor to which the ligand binds. In one embodiment, the transcriptional activation protein is Gal4, Gcn4, Hap1, Adr1, Swi5, Ste12, Mcm1, Yap1, Ace1, Ppr1, Arg81, Lac9, Qa1F, VP16, or a mammalian nuclear receptor. In another embodiment, at least one of the heterologous fusion proteins is expressed from an autonomously-replicating plasmid. In one embodiment, the DNA binding domain comprises a heterologous DNA-binding domain of a transcriptional activation protein. In a preferred embodiment, the DNA binding protein is selected from the group consisting of a mammalian steroid receptor and bacterial LexA protein. In another embodiment, the reporter element is selected from the group consisting of *lacZ*, a polynucleotide encoding luciferase, a polynucleotide encoding green fluorescent protein (GFP), and a polynucleotide encoding chloramphenicol acetyltransferase. In a particularly preferred embodiment, the reporter element is *lacZ*.

The invention further provides for a rescue screen for detecting the activity of a compound for modulating the binding interaction of a first peptide and a second peptide of a peptide binding pair, comprising:

- (i) culturing at least one yeast cell, wherein the yeast cell comprises;
 - a) a nucleotide sequence encoding a first heterologous fusion protein comprising the first peptide or a segment thereof joined to a DNA binding domain of a transcriptional activation protein;
 - b) a nucleotide sequence encoding a second heterologous

fusion protein comprising the second peptide or a segment thereof joined to a transcriptional activation domain of a transcriptional activation protein;

wherein binding of the first peptide or segment thereof and the second peptide or segment thereof reconstitutes a transcriptional activation protein; and

c) a reporter element activated under positive transcriptional control of the reconstituted transcriptional activation protein, wherein expression of the reporter gene prevents exhibition of a selected phenotype;

(ii) incubating a compound with the yeast cell under conditions suitable to detect the selected phenotype; and

(iii) detecting the ability of the compound to affect the binding interaction of the peptide binding pair by determining whether the compound affects the expression of the reporter element which prevents exhibition of the selected phenotype,

wherein said first peptide is a Dkk peptide and said second peptide is a peptide selected from LRP5, HBM, LRP6 and a Dkk-binding fragment of LRP5/LRP6/HBM.

In a preferred embodiment, the invention provides that the yeast cell is *Saccharomyces*. In a particularly preferred embodiment, the *Saccharomyces* cell is *Saccharomyces cerevisiae*. In one embodiment, the compound comprises one or more Dkk interacting or Dkk-1 interacting proteins, or a Dkk-binding fragment thereof. Compounds used in the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191)) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Alternatively, the compound may comprise LRP5 antibodies or Dkk antibodies. In another embodiment, the yeast cell further comprises at least one endogenous nucleotide sequence selected from the group consisting of a nucleotide sequence encoding the DNA binding domain of a

transcriptional activation protein, a nucleotide sequence encoding the transcriptional activation domain of a transcriptional activation protein, and a nucleotide sequence encoding the reporter gene, wherein at least one of the endogenous nucleotide sequences is inactivated by mutation or deletion. In another embodiment, the transcriptional activation protein is Gal4, Gcn4, Hap1, Adr1, Swi5, Ste12, Mcm1, Yap1, Ace1, Ppr1, Arg81, Lac9, Qa1F, VP16, or a mammalian nuclear receptor. In one embodiment, at least one of the heterologous fusion proteins is expressed from an autonomously-replicating plasmid. In another embodiment, the DNA binding domain is a heterologous DNA-binding domain of a transcriptional activation protein.

The invention also provides for a rescue screen for detecting the modulatory activity of a compound on the binding interaction of a first peptide and a second peptide of a peptide binding pair, comprising:

- (i) culturing at least one yeast cell, wherein the yeast cell comprises;
 - a) a nucleotide sequence encoding a first heterologous fusion protein comprising the first peptide or a segment thereof joined to a DNA binding domain of a transcriptional activation protein;
 - b) a nucleotide sequence encoding a second heterologous fusion protein comprising the second peptide or a segment thereof joined to a transcriptional activation domain of a transcriptional activation protein;wherein binding of the first peptide or segment thereof and the second peptide or segment thereof reconstitutes a transcriptional activation protein; and
 - c) a reporter element activated under positive transcriptional control of the reconstituted transcriptional activation protein, wherein expression of the reporter element prevents exhibition of a selected phenotype;
- (ii) incubating a compound with the yeast cell under conditions suitable to detect the selected phenotype; and

- (iii) detecting the ability of the compound to affect the binding interaction of the peptide binding pair by determining whether the compound affects the expression of the reporter element which prevents exhibition of the selected phenotype,

5 wherein said first peptide is a Dkk interacting or Dkk-1 interacting protein peptide and said second peptide is a Dkk or Dkk-1 peptide.

In a preferred embodiment of the rescue screen, the yeast cell is *Saccharomyces*. In a particularly preferred embodiment, the *Saccharomyces* cell is *Saccharomyces cerevisiae*. In another embodiment, the yeast cell further comprises
10 at least one endogenous nucleotide sequence selected from the group consisting of a nucleotide sequence encoding the DNA binding domain of a transcriptional activation protein, a nucleotide sequence encoding the transcriptional activation domain of a transcriptional activation protein, and a nucleotide sequence encoding the reporter gene, wherein at least one of the endogenous nucleotide sequences is
15 inactivated by mutation or deletion. In one embodiment, the transcriptional activation protein is Gal4, Gcn4, Hap1, Adr1, Swi5, Ste12, Mcm1, Yap1, Ace1, Ppr1, Arg81, Lac9, Qa1F, VP16, or a mammalian nuclear receptor. In another embodiment of the rescue screen, at least one of the heterologous fusion proteins is expressed from an autonomously-replicating plasmid. In another embodiment, the DNA binding domain
20 is a heterologous DNA-binding domain of a transcriptional activation protein.

The invention also provides for a method for identifying potential compounds which modulate Dkk activity comprising:

- a) measuring the effect on binding of one or more Dkk interacting protein, or a Dkk-binding fragment thereof, with Dkk or a
25 LRP5/LRP6/HBM binding fragment thereof in the presence and absence of a compound; and
- b) identifying as a potential Dkk modulatory compound a compound which modulates the binding between one or more Dkk interacting proteins or Dkk-binding fragment thereof and
30 Dkk or LRP5/LRP6/HBM fragment thereof.

In a preferred embodiment, the Dkk is Dkk-1.

The invention further provides for any of the Dkk peptide aptamers of Figure 3 (SEQ ID NOs:171-188). The invention also provides for any of the LRP peptide aptamers of Figure 4 (SEQ ID NOs:189-192).

5 Another aspect of the invention provides for a method of identifying agents which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

(a) injecting mRNA encoding Dkk and an agent into a *Xenopus* blastomere;

(b) assessing axis duplication or analyzing marker gene expression; and

10 (c) identifying agents which elicit changes in axis duplication or marker gene expression as agents which modulate the interaction of Dkk with the Wnt signaling pathway. Wherein the agent may be chosen from among mRNA encoding Dkk interacting proteins, fragments thereof, siRNA, shRNA, antisense nucleotides, and antibodies. In a preferred embodiment, Dkk is Dkk-1. In a further embodiment, mRNA of HBM, LRP5/6, any Wnt (including Wnt1-Wnt19, particularly Wnt1, Wnt3, Wnt3a, and Wnt10b), Wnt antagonist, or combination of these is co-injected into the *Xenopus* blastomere. In another embodiment, the marker gene analyzed could include Siamois, Xnr3, slug, Xbra, HNK-1, endodermin, Xlhx8, BMP2, BMP4, XLRP6, EF-1, or ODC.

20 The present invention provides for a method for identifying agents which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

(a) transfecting cells with constructs encoding Dkk and potential Dkk interacting proteins, mRNA fragments thereof, siRNA, shRNA, or antisense, antibodies to LRP5/HBM/LRP6/Dkk/Dkk-interacting protein;

25 (b) assessing changes in expression of a reporter gene linked to a Wnt-responsive promoter; and,

(c) identifying as a Dkk interacting protein any protein which alters reporter gene expression compared with cells transfected with a Dkk construct alone. In a further preferred embodiment, the cells may be HOB-03-CE6, HEK293, or U2OS cells.

In alternative embodiments, the Wnt-responsive promoter is TCF or LEF. In other preferred embodiments, the cells are co-transfected with CMV beta-galactosidase or tk-Renilla.

The present invention further provides for a LRP5/HBM monoclonal or polyclonal antibody to one or more peptides of amino acid sequences MYWTDWVETPRIE (SEQ ID NO:123), MYWTDWGETPRIE (SEQ ID NO:124), KRTGGKRKEILSA (SEQ ID NO:125), ERVEKTTGDKRTRIQGR (SEQ ID NO:126), or KQQCDSFPDCIDGSDE (SEQ ID NO:127).

Additionally, the present invention provides a method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:

- (a) immobilizing LRP5/LRP6/HBM to a solid surface; and
- (b) treating the solid surface with a secreted Dkk protein or a secreted epitope-tagged Dkk and a test compound; and
- (c) determining whether the compound regulates binding between Dkk and LRP5/LRP6/HBM using antibodies to Dkk or the epitope tag or by directly measuring activity of an epitope tag.

In one embodiment, the Dkk is Dkk-1. In a preferred embodiment, the epitope tag is alkaline phosphatase, histidine, myc, or a V5 tag.

Another embodiment of the present invention provides for a method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:

- (a) creating an LRP5, LRP6, or HBM fluorescent fusion protein using a first fluorescent tag;
- (b) creating a Dkk fusion protein comprising a second fluorescent tag;
- (c) adding a test compound; and,
- (d) assessing changes in the ratio of fluorescent tag emissions using Fluorescence Resonance Energy Transfer (FRET) or Bioluminescent Resonance Energy Transfer (BRET) to determine whether the compound modulates Dkk and LRP5/LRP6/HBM interactions.

In a preferred embodiment, the Dkk is Dkk-1.

The present invention also provides for a method of diagnosing low or high bone mass and/or low or high lipid levels in a subject comprising examining expression of Dkk, LRP5, LRP6, HBM or HBM-like variant in the subject and determining whether Dkk, LRP5, LRP6, or HBM or a HBM-like variant is over- or under-expressed to determine whether subject has (a) high or low bone mass and/or (b) high or low lipid levels.

The invention further provides for a transgenic animal wherein Dkk is knocked out in a tissue-specific fashion. In a preferred embodiment, the Dkk is Dkk-1. In one preferred embodiment, the tissue specificity is bone tissue. In another preferred embodiment, the tissue specificity is liver or other tissues or cells involved in regulating lipid metabolism or cancer tissue.

The present invention further provides a method of screening for compounds which modulate the interaction of Dkk with LRP5, LRP6, or HBM comprising:

- (a) exposing LRP5, LRP6, or HBM, or a Dkk-binding fragment of LRP5, LRP6, or HBM to a compound; and
- (b) determining whether said compound bound to LRP5, LRP6, or HBM or the Dkk-binding fragment of LRP5, LRP6, or HBM and further determining whether said compound modulates the interaction of Dkk and LRP5, LRP6, or HBM.

In one embodiment, the Dkk is Dkk-1. In a preferred embodiment, the compound comprises an LRP5 peptide aptamer. Other preferred compositions include the peptide aptamer, OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer, and an LRP5 antibody.

The present invention also provides a method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:

- (a) immobilizing LRP5/LRP6/HBM to a solid surface; and
- (b) treating the solid surface with a secreted Dkk protein or a secreted epitope-tagged Dkk and a test compound; and

- (c) determining whether the compound regulates binding between Dkk and LRP5/LRP6/HBM using antibodies to Dkk or the epitope tag or by directly measuring activity of an epitope tag. In a preferred embodiment, the epitope tag is alkaline phosphatase, histidine, myc or a V5 tag.

In a preferred embodiment, the Dkk is Dkk-1.

The invention also provides for a method for identifying compounds which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

- (a) transfecting cells with constructs containing Dkk and Wnt proteins;
(b) assessing changes in expression of a reporter element linked to a Wnt-responsive promoter; and
(c) identifying as a Dkk/Wnt interaction modulating compound any compound which alters reporter gene expression compared with cells transfected with a Dkk construct alone.

In one embodiment, the Dkk is Dkk-1. In another embodiment, the Wnt is any of Wnt1-Wnt19. In a preferred embodiment, the Wnt is Wnt1, Wnt3, Wnt3a, or Wnt10b. In a particularly preferred embodiment, the Wnt construct contains Wnt3a. In another particularly preferred embodiment, the Wnt construct contains Wnt1. In another preferred embodiment, the Wnt construct encodes for a Wnt that signals through the canonical Wnt pathway. In a particularly preferred embodiment, both Wnt3a and Wnt1 constructs are co-transfected into the cells. In another embodiment, the cells may be U2-OS, HOB-03-CE6, or HEK293 cells. In another embodiment, the reporter element used is TCF-luciferase, tk-Renilla, or a combination thereof.

The invention also provides for a method of testing compounds that modulate Dkk-mediated activity in a mammal comprising:

- (a) providing a group of transgenic animals having (1) a regulatable one or more Dkk genes, (2) a knock-out of Dkk genes, or (3) a knock-in of one or more Dkk genes;

- (b) providing a second group of control animals respectively for the group of transgenic animals in step (a); and
- (c) exposing the transgenic animal group and control animal group to a potential Dkk-modulating compound which modulates bone mass or lipid levels; and
- (d) comparing the transgenic animals and the control group of animals and determining the effect of the compound on bone mass or lipid levels in the transgenic animals as compared to the control animals.

In a preferred embodiment, the Dkk is Dkk-1.

The invention further provides variants of LRP5 which demonstrate HBM biological activity, i.e., that are "HBM-like." In preferred embodiments, variants G171F, M282V, G171K, G171Q, A65V, G171V, G171I, and A214V of LRP5 are provided. The invention further provides for the use any of these variants in the forgoing methods.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows a schematic of the components of the Wnt signal transduction pathway. Schematic obtained from:

<http://www.stanford.edu/~rnusse/pathways/cell2.html>

Figure 2 (A-C) show bait sequences (SEQ ID NOs:168-170) utilized in yeast two hybrid (Y2H) screens for protein-protein interactions.

Figure 3 shows a table of peptide aptamer insert sequences (SEQ ID NOs: 171-192) identified in Y2H screen with a Dkk-1 bait sequence.

Figure 4 shows a table of peptide aptamer insert sequences identified in a Y2H screen using a LRP5 ligand binding domain bait sequence.

Figure 5 shows a table of proteins identified in a Y2H screen using a Dkk-1 bait sequence. These proteins are identified by both their nucleic acid and amino acid accession numbers.

Figure 6 shows the results of a minimum interaction domain mapping screen of Dkk-1 with LRP5. At the top, a map of Dkk-1 showing the location of the signal

sequence, and cysteine rich domains 1 and 2. Below, the extent of domains examined using LRP5 LBD baits, LBD1 and LBD4, of Figure 2. To the right, scoring of the binding results observed in the experiment.

Figure 7 shows a diagram of the *Xenopus* Embryo Assay for Wnt activity.

5 Figure 8 shows the effects of Zmax/LRP5 and HBM on Wnt signaling in the *Xenopus* embryo assay.

Figure 9 shows the effects of Zmax/LRP5 and HBM on induction of secondary axis formation in the *Xenopus* embryo assay.

10 Figure 10 shows the effects of human Dkk-1 on the repression of the canonical Wnt pathway.

Figure 11 shows the effects of human Dkk-1 on Zmax/LRP5 and HBM-mediated Wnt signaling.

15 Figure 12 shows pcDNA3.1 construct names with nucleotide sequences (including SEQ ID NOs:193-203) for LRP5-binding peptide aptamers, Dkk-1 peptides and control constructs.

Figure 13 shows the amino acid sequences (including SEQ ID NOs:204-214) for the corresponding LRP5-binding peptides, Dkk-1 peptide aptamers and control constructs in Figure 12.

20 Figure 14 shows the effects of Dkk-1 and Dkk-2 on Wnt1 signaling with coreceptors LRP5, HBM, and LRP6 in HOB03CE6 cells.

Figure 15 shows the effects of Dkk-1 and Dkk-2 on Wnt3a signaling with coreceptors LRP5, HBM, and LRP6 in HOB03CE6 cells.

Figure 16 demonstrates that the LRP5-LBD peptide aptamer 262 activates Wnt signaling in the presence of Wnt3a in U2OS cells.

25 Figure 17 shows the differential binding of an antibody generated to a sequence (a.a. 165-177) containing the HBM mutation in LRP5 in LRP5 and HBM virus-infected cells.

30 Figure 18 shows data generated from a Y2H interaction trap where a mutant Dkk-1 (C220A) is unable to bind to LRP5 and demonstrating the window of capability of detecting small molecule effects on LRP and Dkk interactions.

Figure 19 shows that Dkk-1 represses Wnt3a-mediated Wnt signaling in U2OS bone cells using the cell-based reporter gene assay for high throughput screening.

5 Figure 20 demonstrates that Wnt1-HBM generated signaling is not efficiently inhibited by Dkk-1 in U2OS bone cells while LRP5 and LRP6-mediated signaling are using the cell-based reporter gene assay for high throughput screening.

Figure 21 shows that the TCF signal in the cell-based reporter gene assay for high throughput screening can be modulated by Dkk-1 and Dkk-1-AP without Wnt DNA transfection.

10 Figure 22 shows the morphological results in the Xenopus assay using aptamers 261 and 262 from the LRP5-LBD to activate Wnt signaling.

Figure 23 demonstrates that LRP5-LBD aptamers 261 and 262 induce Wnt signaling over other LRP5 aptamers.

15 Figure 24 shows that the mutation G171F in LRP5 produces a greater activation of the Wnt pathway than LRP5 which is consistent with HBM activity.

Figure 25 shows that the mutation M282V in LRP5 produces an activation of the Wnt pathway which is consistent with HBM activity in U2OS cells.

20 Figure 26 shows the amino acid sequence of the various peptides of dkk-1 selected to generate polyclonal antibodies, their relationship to the Dkk-1 amino acid sequence and identities of polyclonal antibodies generated.

Figure 27 shows a Western blot demonstrating that polyclonal antibody #5521 to amino acids 165-186 of Dkk-1 was able to detect Dkk1-V5 and Dkk1-AP from conditioned medium.

25 Figure 28 shows a Western blot demonstrating that polyclonal antibody #74397 to amino acids 147-161 was able to detect Dkk1-V5 in both conditioned medium and immunoprecipitated conditioned medium.

DETAILED DESCRIPTION OF THE INVENTION

1. Definitions

In general, terms in the present application are used consistent with the manner in which those terms are understood in the art. To aid in the understanding of the specification and claims, the following definitions are provided.

"Gene" refers to a DNA sequence that encodes through its template or messenger RNA a sequence of amino acids characteristic of a specific peptide. The term "gene" includes intervening, non-coding regions, as well as regulatory regions, and can include 5' and 3' ends.

By "nucleic acid" is meant to include single stranded and double stranded nucleic acids including, but not limited to DNAs, RNAs (e.g., mRNA, tRNAs, siRNAs), cDNAs, recombinant DNA (rDNA), rRNAs, antisense nucleic acids, oligonucleotides, and oligomers, and polynucleotides. The term may also include hybrids such as triple stranded regions of RNA and/or DNA or double stranded RNA:DNA hybrids. The term also is contemplated to include modified nucleic acids such as, but not limited to biotinylated nucleic acids, tritylated nucleic acids, fluorophor labeled nucleic acids, inosine, and the like.

"Gene sequence" refers to a nucleic acid molecule, including DNA which contains a non-transcribed or non-translated sequence, which comprises a gene. The term is also intended to include any combination of gene(s), gene fragment(s), non-transcribed sequence(s) or non-translated sequence(s) which are present on the same DNA molecule.

The nucleic acid sequences of the present invention may be derived from a variety of sources including DNA, cDNA, synthetic DNA, synthetic RNA or combinations thereof. Such sequences may comprise genomic DNA which may or may not include naturally occurring introns. Moreover, such genomic DNA may be obtained in association with promoter regions and/or poly (A) sequences. The sequences, genomic DNA or cDNA may be obtained in any of several ways. Genomic DNA can be extracted and purified from suitable cells by means well

known in the art. Alternatively, mRNA can be isolated from a cell and used to produce cDNA by reverse transcription or other means.

"cDNA" refers to complementary or copy DNA produced from an RNA template by the action of RNA-dependent DNA polymerase (reverse transcriptase).

5 Thus, a "cDNA clone" means a duplex DNA sequence for which one strand is complementary to an RNA molecule of interest, carried in a cloning vector or PCR amplified. cDNA can also be single stranded after first strand synthesis by reverse transcriptase. In this form, it is a useful PCR template and does not need to be carried in a cloning vector. This term includes genes from which the intervening
10 sequences have been removed. Thus, the term "gene", as sometimes used generically, can also include nucleic acid molecules comprising cDNA and cDNA clones.

"Recombinant DNA" means a molecule that has been engineered by splicing
15 *in vitro* a cDNA or genomic DNA sequence or altering a sequence by methods such as PCR mutagenesis.

"Cloning" refers to the use of *in vitro* recombination techniques to insert a particular gene or other DNA sequence into a vector molecule. In order to successfully clone a desired gene, it is necessary to use methods for generating DNA fragments, for joining the fragments to vector molecules, for introducing the
20 composite DNA molecule into a host cell in which it can replicate, and for selecting the clone having the target gene from amongst the recipient host cells.

"cDNA library" refers to a collection of recombinant DNA molecules containing cDNA inserts which together comprise the entire or a partial repertoire of genes expressed in a particular tissue or cell source. Such a cDNA library can be prepared
25 by methods known to one skilled in the art and described by, for example, Cowell and Austin, "cDNA Library Protocols," *Methods in Molecular Biology* (1997).

"Cloning vehicle" refers to a plasmid or phage DNA or other DNA sequence which is able to replicate in a host cell. This term can also include artificial chromosomes such as BACs and YACs. The cloning vehicle is characterized by one
30 or more endonuclease recognition sites at which such DNA sequences may be cut in

a determinable fashion without loss of an essential biological function of the DNA, which may contain a marker suitable for use in the identification of transformed cells.

"Expression" refers to the process comprising transcription of a gene sequence and subsequent processing steps, such as translation of a resultant mRNA to produce the final end product of a gene. The end product may be a protein (such as an enzyme or receptor) or a nucleic acid (such as a tRNA, antisense RNA, or other regulatory factor). The term "expression control sequence" refers to a sequence of nucleotides that control or regulate expression of structural genes when operably linked to those genes. These include, for example, the lac systems, the trp system, major operator and promoter regions of the phage lambda, the control region of fd coat protein and other sequences known to control the expression of genes in prokaryotic or eukaryotic cells. Expression control sequences will vary depending on whether the vector is designed to express the operably linked gene in a prokaryotic or eukaryotic host, and may contain transcriptional elements such as enhancer elements, termination sequences, tissue-specificity elements and/or translational initiation and termination sites.

"Expression vehicle" refers to a vehicle or vector similar to a cloning vehicle but which is capable of expressing a gene which has been cloned into it, after transformation into a host. The cloned gene is usually placed under the control of (i.e., operably linked to) an expression control sequence.

"Operator" refers to a DNA sequence capable of interacting with the specific repressor, thereby controlling the transcription of adjacent gene(s).

"Promoter" refers to a DNA sequence that can be recognized by an RNA polymerase. The presence of such a sequence permits the RNA polymerase to bind and initiate transcription of operably linked gene sequences.

"Promoter region" is intended to include the promoter as well as other gene sequences which may be necessary for the initiation of transcription. The presence of a promoter region is sufficient to cause the expression of an operably linked gene sequence. The term "promoter" is sometimes used in the art to generically indicate a promoter region. Many different promoters are known in the art which direct

expression of a gene in a certain cell types. Tissue-specific promoters can comprise nucleic acid sequences which cause a greater (or decreased) level of expression in cells of a certain tissue type.

"Operably linked" means that the promoter controls the initiation of expression of the gene. A promoter is operably linked to a sequence of proximal DNA if upon introduction into a host cell the promoter determines the transcription of the proximal DNA sequence(s) into one or more species of RNA. A promoter is operably linked to a DNA sequence if the promoter is capable of initiating transcription of that DNA sequence.

"Prokaryote" refers to all organisms without a true nucleus, including bacteria.

"Eukaryote" refers to organisms and cells that have a true nucleus, including mammalian cells.

"Host" includes prokaryotes and eukaryotes, such as yeast and filamentous fungi, as well as plant and animal cells. The term includes an organism or cell that is the recipient of a replicable expression vehicle.

The term "animal" is used herein to include all vertebrate animals, except humans. It also includes an individual animal in all stages of development, including embryonic and fetal stages. Preferred animals include higher eukaryotes such as avians, rodents (e.g., mice, rabbits, rats, chinchillas, guinea pigs, hamsters and the like), and mammals. Preferred mammals include bovine, equine, feline, canine, ovine, caprine, porcine, buffalo, humans, and primates.

A "transgenic animal" is an animal containing one or more cells bearing genetic information received, directly or indirectly, by deliberate genetic manipulation or by inheritance from a manipulated progenitor at a subcellular level, such as by microinjection or infection with a recombinant viral vector (e.g., adenovirus, retrovirus, herpes virus, adeno-associated virus, lentivirus). This introduced DNA molecule may be integrated within a chromosome, or it may be extra-chromosomally replicating DNA.

"Embryonic stem cells" or "ES cells" as used herein are cells or cell lines usually derived from embryos which are pluripotent meaning that they are un-

differentiated cells. These cells are also capable of incorporating exogenous DNA by homologous recombination and subsequently developing into any tissue in the body when incorporated into a host embryo. It is possible to isolate pluripotent cells from sources other than embryonic tissue by methods which are well understood in the art.

Embryonic stem cells in mice have enabled researchers to select for transgenic cells and perform gene targeting. This allows more genetic engineering than is possible with other transgenic techniques. For example, mouse ES cells are relatively easy to grow as colonies *in vitro*. The cells can be transfected by standard procedures and transgenic cells clonally selected by antibiotic resistance. See, for example, Doetschman *et al.*, 1994, *Gene transfer in embryonic stem cells*. In Pinkert (Ed.) Transgenic Animal Technology: A Laboratory Handbook. Academic Press Inc., New York, pp.115-146. Furthermore, the efficiency of this process is such that sufficient transgenic colonies (hundreds to thousands) can be produced to allow a second selection for homologous recombinants. Mouse ES cells can then be combined with a normal host embryo and, because they retain their potency, can develop into all the tissues in the resulting chimeric animal, including the germ cells. The transgenic modification can then be transmitted to subsequent generations.

Methods for deriving embryonic stem (ES) cell lines *in vitro* from early preimplantation mouse embryos are well known. See for example, Evans *et al.*, 1981 *Nature* 29: 154-6 and Martin, 1981, *Proc. Nat. Acad. Sci. USA*, 78: 7634-8. ES cells can be passaged in an undifferentiated state, provided that a feeder layer of fibroblast cells or a differentiation inhibiting source is present.

The term "somatic cell" indicates any animal or human cell which is not a sperm or egg cell or is capable of becoming a sperm or egg cell. The term "germ cell" or "germ-line cell" refers to any cell which is either a sperm or egg cell or is capable of developing into a sperm or egg cell and can therefore pass its genetic information to offspring. The term "germ cell-line transgenic animal" refers to a transgenic animal in which the genetic information was incorporated in a germ line cell, thereby conferring the ability to transfer the information to offspring. If such

offspring in fact possess some or all of that information, then they, too, are transgenic animals.

The genetic alteration of genetic information may be foreign to the species of animal to which the recipient belongs, or foreign only to the particular individual recipient. In the last case, the altered or introduced gene may be expressed differently than the native gene.

"Fragment" of a gene refers to any portion of a gene sequence. A "biologically active fragment" refers to any portion of the gene that retains at least one biological activity of that gene. For example, the fragment can perhaps hybridize to its cognate sequence or is capable of being translated into a polypeptide fragment encoded by the gene from which it is derived.

"Variant" refers to a gene that is substantially similar in structure and biological activity or immunological characteristics to either the entire gene or to a fragment of the gene. Provided that the two genes possess a similar activity, they are considered variant as that term is used herein even if the sequence of encoded amino acid residues is not identical. Preferentially, as used herein (unless otherwise defined) the variant is one of LRP5, HBM or LRP6. The variant preferably is one that yields an HBM-like phenotype (i.e., enhances bones mass and/or modulates lipid levels). These variants include missense mutations, single nucleotide polymorphisms (SNPs), mutations which result in changes in the amino acid sequence of the protein encoded by the gene or nucleic acid, and combinations thereof, as well as com in the exon domains of the *HBM* gene and mutations in LRP5 or LRP6 which result in an HBM like phenotype.

"Amplification of nucleic acids" refers to methods such as polymerase chain reaction (PCR), ligation amplification (or ligase chain reaction, LCR) and amplification methods based on the use of Q-beta replicase. These methods are well known in the art and described, for example, in U.S. Patent Nos. 4,683,195 and 4,683,202. Reagents and hardware for conducting PCR are commercially available. Primers useful for amplifying sequences from the HBM region are preferably complementary to, and hybridize specifically to sequences in the HBM region or in

regions that flank a target region therein. HBM sequences generated by amplification may be sequenced directly. Alternatively, the amplified sequence(s) may be cloned prior to sequence analysis.

"Antibodies" may refer to polyclonal and/or monoclonal antibodies and fragments thereof, and immunologic binding equivalents thereof, that can bind to the HBM proteins and fragments thereof or to nucleic acid sequences from the HBM region, particularly from the HBM locus or a portion thereof. Preferred antibodies also include those capable of binding to LRP5, LRP6 and HBM variants. The term antibody is used both to refer to a homogeneous molecular entity, or a mixture such as a serum product made up of a plurality of different molecular entities. Proteins may be prepared synthetically in a protein synthesizer and coupled to a carrier molecule and injected over several months into rabbits. Rabbit sera is tested for immunoreactivity to the HBM protein or fragment. Monoclonal antibodies may be made by injecting mice with the proteins, or fragments thereof. Monoclonal antibodies will be screened by ELISA and tested for specific immunoreactivity with HBM protein or fragments thereof. Harlow *et al.*, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1988) and *Using Antibodies: A Laboratory Manual*, Harlow, Ed and Lane, David (Cold Spring Harbor Press, 1999). These antibodies will be useful in assays as well as pharmaceuticals. By "antibody" is meant to include but not limited to polyclonal, monoclonal, chimeric, human, humanized, bispecific, multispecific, primatized™ antibodies.

"HBM protein" refers to a protein that is identical to a Zmax1 (LRP5) protein except that it contains an alteration of glycine 171 to a valine. An HBM protein is defined for any organism that encodes a Zmax1 (LRP5) true homolog. For example, a mouse HBM protein refers to the mouse Zmax1 (LRP5) protein having the glycine 170 to valine substitution.

By "HBM-like" is meant a variant of LRP5, LRP6 or HBM which when expressed in a cell is capable of modulating bone mass, lipid levels, Dkk activity, and/or Wnt activity.

In one embodiment of the present invention, "*HBM* gene" refers to the genomic DNA sequence found in individuals showing the HBM characteristic or phenotype, where the sequence encodes the protein indicated by SEQ ID NO: 4. The *HBM* gene and the *Zmax1* (*LRP5*) gene are allelic. The protein encoded by the *HBM* gene has the property of causing elevated bone mass, while the protein encoded by the *Zmax1* (*LRP5*) gene does not. The *HBM* gene and the *Zmax1* (*LRP5*) gene differ in that the *HBM* gene has a thymine at position 582, while the *Zmax1* gene has a guanine at position 582. The *HBM* gene comprises the nucleic acid sequence shown as SEQ ID NO: 2. The *HBM* gene may also be referred to as an "HBM polymorphism." Other *HBM* genes may further have silent mutations, such as those discussed in Section 3 below.

In alternative embodiments of the present invention, "HBM gene" may also refer to any allelic variant of *Zmax1* (*LRP5*) or *LRP6* which results in the HBM phenotype. Such variants may include alteration from the wild-type protein coding sequence as described herein and/or alteration in expression control sequences of *Zmax1* (*LRP5*) or contains an amino acid mutation in *LRP5* or *LRP6*, such that the resulting protein produces a phenotype which enhances bone mass and/or modulates lipid levels. A preferred example of such a variant is an alteration of the endogenous *Zmax1* (*LRP5*) promoter region resulting in increased expression of the *Zmax1* (*LRP5*) protein.

"Normal," "wild-type," "unaffected", "*Zmax1*", "*Zmax*", "*LR3*" and "*LRP5*" all refer to the genomic DNA sequence that encodes the protein indicated by SEQ ID NO: 3. *LRP5* has also been referred to *LRP7* in mouse. *Zmax1*, *LRP5* and *Zmax* may be used interchangeably throughout the specification and are meant to be the same gene, perhaps only relating to the gene in a different organism. The *Zmax1* gene has a guanine at position 582 in the human sequence. The *Zmax1* gene of human comprises the nucleic acid sequence shown as SEQ ID NO: 1. "Normal," "wild-type," "unaffected", "*Zmax1*" and "*LRP5*" also refer to allelic variants of the genomic sequence that encodes proteins that do not contribute to elevated bone

mass. The *Zmax1* (*LRP5*) gene is common in the human population, while the *HBM* gene is rare.

"Bone development" generally refers to any process involved in the change of bone over time, including, for example, normal development, changes that occur during disease states, and changes that occur during aging. This may refer to structural changes and dynamic rate changes such as growth rates, resorption rates, bone repair rates, and etc. "Bone development disorder" particularly refers to any disorders in bone development including, for example, changes that occur during disease states and changes that occur during aging. Bone development may be progressive or cyclical in nature. Aspects of bone that may change during development include, for example, mineralization, formation of specific anatomical features, and relative or absolute numbers of various cell types.

"Bone modulation" or "modulation of bone formation" refers to the ability to affect any of the physiological processes involved in bone remodeling, as will be appreciated by one skilled in the art, including, for example, bone resorption and appositional bone growth, by, *inter alia*, osteoclastic and osteoblastic activity, and may comprise some or all of bone formation and development as used herein.

Bone is a dynamic tissue that is continually adapting and renewing itself through the renewal of old or unnecessary bone by osteoclasts and the rebuilding of new bone by osteoblasts. The nature of the coupling between these processes is responsible for both the modeling of bone during growth as well as the maintenance of adult skeletal integrity through remodeling and repair to meet the everyday needs of mechanical usage. There are a number of diseases that result from an uncoupling of the balance between bone resorption and formation. With aging there is a gradual "physiologic" imbalance in bone turnover, which is particularly exacerbated in women due to menopausal loss of estrogen support, that leads to a progressive loss of bone. As bone mineral density falls below population norms there is a consequent increase in bone fragility and susceptibility to spontaneous fractures. For every 10 percent of bone that is lost, the risk of fracture doubles. Individuals with bone mineral density (BMD) in the spine or proximal femur 2.5 or

more standard deviations below normal peak bone mass are classified as osteoporotic. However, osteopenic individuals with BMD between 1 and 2.5 standard deviations below the norm are clearly at risk.

Bone is measured by several different forms of X-ray absorptiometry. All of the instruments measure the inorganic or bone mineral content of the bone. Standard DXA measurements give a value that is an areal density, not a true density measurement by the classical definition of density (mass/unit volume). Nevertheless, this is the type of measurement used clinically to diagnose osteoporosis. However, while BMD is a major contributing factor to bone strength, as much as 40% of bone strength stems from other factors including: 1) bone size (*i.e.*, larger diameters increase organ-level stiffness, even in the face of lower density); 2) the connectivity of trabecular structures; 3) the level of remodeling (remodeling loci are local concentrators of strain); and 4) the intrinsic strength of the bony material itself, which in turn is a function of loading history (*i.e.*, through accumulated fatigue damage) and the extent of collagen cross-linking and level of mineralization. There is good evidence that all of these strength/fragility factors play some role in osteoporotic fractures, as do a host of extraskeletal influences as well (such as fall patterns, soft tissue padding, and central nervous system reflex responsiveness).

Additional analytical instruments can be used to address these features of bone. For example, the pQCT allows measurement of separate trabecular and cortical compartments for size and density and the μ CT provides quantitative information on architectural features such as trabecular connectivity. The μ CT also gives a true bone density measurement. With these tools, the important non-BMD parameters can be measured for diagnosing the extent of disease and the efficacy of treatments. Current treatments for osteoporosis are based on the ability of drugs to prevent or retard bone resorption. Although newer anti-resorptive agents are proving to be useful in the therapy of osteoporosis, they are viewed as short-term solutions to the more definitive challenge to develop treatments that will increase bone mass and/or the bone quality parameters mentioned above.

Thus, bone modulation may be assessed by measuring parameters such as bone mineral density (BMD) and bone mineral content (BMC) by pDXA X-ray methods, bone size, thickness or volume as measured by X-ray, bone formation rates as measured for example by calcien labeling, total, trabecular, and mid-shaft density as measured by pQCT and/or μ CT methods, connectivity and other histological parameters as measured by μ CT methods, mechanical bending and compressive strengths as preferably measured in femur and vertebrae respectively. Due to the nature of these measurements, each may be more or less appropriate for a given situation as the skilled practitioner will appreciate. Furthermore, parameters and methodologies such as a clinical history of freedom from fracture, bone shape, bone morphology, connectivity, normal histology, fracture repair rates, and other bone quality parameters are known and used in the art. Most preferably, bone quality may be assessed by the compressive strength of vertebra when such a measurement is appropriate. Bone modulation may also be assessed by rates of change in the various parameters. Most preferably, bone modulation is assessed at more than one age.

"Normal bone density" refers to a bone density within two standard deviations of a Z score of 0 in the context of the HBM linkage study. In a general context, the range of normal bone density parameters is determined by routine statistical methods. A normal parameter is within about 1 or 2 standard deviations of the age and sex normalized parameter, preferably about 2 standard deviations. A statistical measure of meaningfulness is the P value which can represent the likelihood that the associated measurement is significantly different from the mean. Significant P values are $P < 0.05$, 0.01 , 0.005 , and 0.001 , preferably at least $P < 0.01$.

"HBM" refers to "high bone mass" although this term may also be expressed in terms of bone density, mineral content, and size.

The "HBM phenotype" and "HBM-like phenotype" may be characterized by an increase of about 2 or more standard deviations, preferably 2, 2.5, 3, or more standard deviations in 1, 2, 3, 4, 5, or more quantitative parameters of bone modulation, preferably bone density and mineral content and bone strength

parameters, above the age and sex norm for that parameter. The HBM phenotype and HBM-like phenotype are characterized by statistically significant increases in at least one parameter, preferably at least 2 parameters, and more preferably at least 3 or more parameters. The HBM phenotype and the HBM-like phenotype may also be characterized by an increase in one or more bone quality parameters and most preferably increasing parameters are not accompanied by a decrease in any bone quality parameters. Most preferably, an increase in bone modulation parameters and/or bone quality measurements is observed at more than one age. The HBM phenotype and HBM-like phenotype also includes changes of lipid levels, Wnt activity and/or Dkk activity.

The terms "isolated" and "purified" refer to a substance altered by hand of man from the natural environment. An isolated peptide may be for example in a substantially pure form or otherwise displaced from its native environment such as by expression in an isolated cell line or transgenic animal. An isolated sequence may for example be a molecule in substantially pure form or displaced from its native environment such that at least one end of said isolated sequence is not contiguous with the sequence it would be contiguous with in nature.

"Biologically active" refers to those forms of proteins and polypeptides, including conservatively substituted variants, alleles of genes encoding a protein or polypeptide fragments of proteins which retain a biological and/or immunological activity of the wild-type protein or polypeptide. Preferably the activity is one which induces a change in Dkk activity, such as inhibiting the interaction of Dkk with a ligand binding partner (e.g., LRP5 or LRP6 or Dkk-1 with a Dkk-1 interacting protein such as those shown in Figure 5). By biologically active is also meant to include any form which modulates Wnt signaling.

By "modulate" and "regulate" is meant methods, conditions, or agents which increase or decrease the wild-type activity of an enzyme, inhibitor, signal transducer, receptor, transcription activator, co-factor, and the like. This change in activity can be an increase or decrease of mRNA translation, mRNA or DNA transcription, and/or

mRNA or protein degradation, which may in turn correspond to an increase or decrease in biological activity.

By "modulated activity" is meant any activity, condition, disease or phenotype which is modulated by a biologically active form of a protein. Modulation may be effected by affecting the concentration or subcellular localization of biologically active protein, *i.e.*, by regulating expression or degradation, or by direct agonistic or antagonistic effect as, for example, through inhibition, activation, binding, or release of substrate, modification either chemically or structurally, or by direct or indirect interaction which may involve additional factors.

By "effective amount" or "dose effective amount" or "therapeutically effective amount" is meant an amount of an agent which modulates a biological activity of the polypeptide of the invention.

By "immunologically active" is meant any immunoglobulin protein or fragment thereof which recognizes and binds to an antigen.

By "Dkk" is meant to refer to the nucleic acids and proteins of members of the Dkk (Dickkopf) family. This includes, but is not limited to, Dkk-1, Dkk-2, Dkk-3, Dkk-4, Soggy, and related Dkk proteins. Dkk-1 is a preferred embodiment of the present invention. However, the Dkk proteins have substantial homology and one skilled in the art will appreciate that all of the embodiments of the present invention utilizing Dkk-1 may also be utilized with the other Dkk proteins.

By "Dkk-1" is meant to refer to the Dkk-1 protein and nucleic acids which encode the Dkk-1 protein. Dkk-1 refers to Dickkopf-1, and in *Xenopus* it is related to at least Dkk-2, Dkk-3, and Dkk-4 (see Krupnik *et al.*, *Gene* 238:301-313 (1999)). Dkk-1 was first identified in *Xenopus* (Glinka *et al.*, *Nature* 391:357-62 (1998)). It was recognized as a factor capable of inducing ectopic head formation in the presence of inhibition of the BMP pathway. It was then also found to inhibit the axis-inducing activity of several *Xenopus* Wnt molecules by acting as an extracellular antagonist of Wnt signaling. Mammalian homologs have been found including Dkk-1, Dkk-2, Dkk-3, Dkk-4 and soggy (Fedi *et al.*, 1999 and Krupnick *et al.* 1999). Human Dkk-1 was also referred to as sk (Fedi *et al.* 1999). As used herein, Dkk-1 is

meant to include proteins from any species having a Wnt pathway in which Dkk-1 interacts. Particularly preferred are mammalian species (e.g., murine, caprine, canine, bovine, feline, equine, primate, ovine, porcine and the like), with particularly preferred mammals being humans. Nucleic acid sequences encoding Dkk-1 include, but are not limited to human Dkk-1 (GenBank Accession Nos. AH009834, XM_005730, AF261158, AF261157, AF177394, AF127563 and NM_012242), *Mus musculus* dickkopf homolog 1 (GenBank Accession No. NM_010051), and *Danio rerio* dickkopf-1 (GenBank Accession Nos. AF116852 and AB023488). The genomic sequences with exon annotation are GenBank Accession Nos. AF261157 and AF261158. Also contemplated are homologs of these sequences which have Dkk-1 activity in the Wnt pathway. Dkk-1 amino acid sequences include, but are not limited to human dickkopf homolog 1 (GenBank Accession Nos. AAG15544, BAA34651, NP_036374, AAF02674, AAD21087, and XP_005730), *Danio rerio* (zebrafish) dickkopf1 (GenBank Accession Nos. BAA82135 and AAD22461) and murine dickkopf-1 (GenBank Accession Nos. O54908 and NP_034181). Variants and homologs of these sequences which possess Dkk-1 activity are also included when referring to Dkk-1.

By "Dkk mediated" disorder, condition or disease is any abnormal state that involves Dkk activity. The abnormal state can be induced by environmental exposure or drug administration. Alternatively, the disease or disorder can be due to a genetic defect. Dkk mediated diseases, disorders and conditions include but are not limited to bone mass disorders or conditions and lipid disorders and conditions. For example, bone mass disorders/conditions/diseases, which may be mediated by Dkk, include but are not limited to age related loss of bone, bone fractures (e.g., hip fracture, Colle's fracture, vertebral crush fractures), chondrodystrophies, drug-induced disorders (e.g., osteoporosis due to administration of glucocorticoids or heparin and osteomalacia due to administration of aluminum hydroxide, anticonvulsants, or glutethimide), high bone turnover, hypercalcemia, hyperostosis, osteogenesis imperfecta, osteomalacia, osteomyelitis, osteoporosis, Paget's disease, osteoarthritis, and rickets.

Lipid disorders/diseases/conditions, which may be mediated by Dkk, include but are not limited to familial lipoprotein lipase deficiency, familial apoprotein CII deficiency, familial type 3 hyperlipoproteinemia, familial hypercholesterolemia, familial hypertriglyceridemia, multiple lipoprotein-type hyperlipidemia, elevated lipid
5 levels due to dialysis and/or diabetes, and elevated lipid levels of unknown etiologies

The term "recognizes and binds," when used to define interactions of antisense nucleotides, siRNAs (small inhibitory RNA), or shRNA (short hairpin RNA) with a target sequence, means that a particular antisense, siRNA, or shRNA sequence is substantially complementary to the target sequence, and thus will
10 specifically bind to a portion of an mRNA encoding polypeptide. As such, typically the sequences will be highly complementary to the mRNA target sequence, and will have no more than 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 base mismatches throughout the sequence. In many instances, it may be desirable for the sequences to be exact matches, i.e. be completely complementary to the sequence to which the
15 oligonucleotide specifically binds, and therefore have zero mismatches along the complementary stretch. As such, highly complementary sequences will typically bind quite specifically to the target sequence region of the mRNA and will therefore be highly efficient in reducing, and/or even inhibiting the translation of the target mRNA sequence into polypeptide product.

Substantially complementary oligonucleotide sequences will be greater than
20 about 80 percent complementary (or `% exact-match`) to the corresponding mRNA target sequence to which the oligonucleotide specifically binds, and will, more preferably be greater than about 85 percent complementary to the corresponding mRNA target sequence to which the oligonucleotide specifically binds. In certain
25 aspects, as described above, it will be desirable to have even more substantially complementary oligonucleotide sequences for use in the practice of the invention, and in such instances, the oligonucleotide sequences will be greater than about 90 percent complementary to the corresponding mRNA target sequence to which the oligonucleotide specifically binds, and may in certain embodiments be greater than
30 about 95 percent complementary to the corresponding mRNA target sequence to

which the oligonucleotide specifically binds, and even up to and including 96%, 97%, 98%, 99%, and even 100% exact match complementary to the target mRNA to which the designed oligonucleotide specifically binds.

Percent similarity or percent complementary of any of the disclosed sequences may be determined, for example, by comparing sequence information using the GAP computer program, version 6.0, available from the University of Wisconsin Genetics Computer Group (UWGCG). The GAP program utilizes the alignment method of Needleman and Wunsch (1970). Briefly, the GAP program defines similarity as the number of aligned symbols (i.e., nucleotides or amino acids) which are similar, divided by the total number of symbols in the shorter of the two sequences. The preferred default parameters for the GAP program include: (1) a unary comparison matrix (containing a value of 1 for identities and 0 for non-identities) for nucleotides, and the weighted comparison matrix of Gribskov and Burgess (1986), (2) a penalty of 3.0 for each gap and an additional 0.10 penalty for each symbol in each gap; and (3) no penalty for end gaps.

By "mimetic" is meant a compound or molecule that performs the same function or behaves similarly to the compound mimicked.

By "reporter element" is meant a polynucleotide that encodes a polypeptide capable of being detected in a screening assays. Examples of polypeptides encoded by reporter elements include, but are not limited to, lacZ, GFP, luciferase, and chloramphenicol acetyltransferase.

2. Introduction

A polymorphism in LRP5 (Zmax), G171V, designated as HBM, has been identified as conferring a high bone mass phenotype in a population of related subjects as described in co-pending applications International Patent Application PCT/US 00/16951, and U.S. Patent Application Nos. 09/543,771 and 09/544,398, which are hereby incorporated by reference in their entirety (Little *et al.*, *Am J Hum Genet.* 70:11-19 (2002)). LRP5 is also described in International Patent Application WO 98/46743, which is incorporated by reference in its entirety. Loss of LRP5

function has been shown to have a deleterious effect on bone (Gong *et al.*, *Cell* 107:513-523 (2001)). Additionally, the HBM polymorphism and LRP5 may also be important in cardiac health and lipid-mediated disorders. Thus, methods of regulating their activity can serve as methods of treating and/or preventing cardiac and lipid-mediated disorders.

Recent studies have indicated that LRP5 participates in the Wnt signal transduction pathway. The Wnt pathway is critical in limb early embryological development. A recently published sketch of the components of Wnt signaling is shown in Figure 1

(Nusse, 2001 <http://www.stanford.edu/~rnusse/pathways/cell2.html>) (see also, Nusse, *Nature* 411:255-6 (2001); and Mao *et al.*, *Nature* 411:321-5 (2001)). Briefly summarized, Wnt proteins are secreted proteins which interact with the transmembrane protein Frizzled (Fz). LRP proteins, such as LRP5 and LRP6, are believed to modulate the Wnt signal in a complex with Fz (Tamai *et al.*, *Nature* 407:530-5 (2000)). The Wnt pathway acts intracellularly through the Disheveled protein (Dsh) which in turn inhibits glycogen synthetase kinase-3 (GSK3) from phosphorylating β -catenin. Phosphorylated β -catenin is rapidly degraded following ubiquitination. However, the stabilized β -catenin accumulates and translocates to the nucleus where it acts as a cofactor of the T-cell factor (TCF) transcription activator complex.

The protein dickkopf-1 (Dkk-1) is reported to be an antagonist of Wnt pathway. Dkk-1 is required for head formation in early development. Dkk-1 and its function in the Wnt pathway are described in *e.g.*, Krupnik, *et al.*, *Gene* 238:301-13 (1999); Fedi *et al.*, *J. Biol. Chem.* 274:19465-72 (1999); see also for Dkk-1 and the Wnt pathway, Wu *et al.*, *Curr. Biol.* 10:1611-4 (2000), Shinya *et al.*, *Mech. Dev.* 98:3-17 (2000), Mukhopadhyay *et al.*, *Dev Cell* 1:423-434 (2001) and in PCT Patent Application No. WO 00/52047, and in references cited in each. It has been known that Dkk-1 acts upstream of Dsh, however the nature of the mechanism of inhibition by Dkk-1 is just beginning to be elucidated. Dkk-1 is expressed in the mouse embryonic limb bud and its disruption results in abnormal limb morphogenesis, among

other developmental defects (Gotewold *et al.*, *Mech. Dev.* 89:151-3 (1999); and, Mukhopadhyay *et al.*, *Dev Cell* 1:423-434 (2001)).

Related U.S. provisional application 60/291,311 disclosed a novel interaction between Dkk-1 (GenBank Accession No. XM 005730) and LRP5. The interaction
5 between Dkk-1 and LRP5 was discovered by a yeast two hybrid (Y2H) screen for proteins which interact with the ligand binding domain of LRP5, as described in Example 1. The two-hybrid screen is a common procedure in the art, which is described, for example, by Gietz *et al.*, *Mol. Cell. Biochem.* 172:67-79 (1997); Young, *Biol. Reprod.* 58:302-11 (1998); Brent and Finley, *Ann. Rev. Genet.* 31:663-
10 704 (1997); and Lu and Hannon, eds., Yeast Hybrid Technologies, Eaton Publishing, Natick MA, (2000). More recently, other studies confirm that Dkk-1 is a binding partner for LRP and modulates the Wnt pathway via direct binding with LRP (R. Nusse, *Nature* 411:255-256 (2001); A. Bafico *et al.*, *Nat. Cell Biol.* 3:683-686 (2001); M. Semenov, *Curr. Biol.* 11:951-961 (2001); B. Mao, *Nature* 411:321-325 (2001),
15 Zorn, *Curr. Biol.* 11:R592-5 (2001)); and, L. Li *et al.*, *J. Biol Chem.* 277:5977-81 (2002)).

Mao and colleagues (2001) identified Dkk-1 as a ligand for LRP6. Mao *et al.* suggest that Dkk-1 and LRP6 interact antagonistically where Dkk proteins inhibit the Wnt coreceptor functions of LRP6. Using co-immunoprecipitation, the group verified
20 that the Dkk-1/LRP6 interaction was direct. Dkk-2 was also found to directly bind LRP6. Contrary to data contained in provisional application 60/291,311, Mao *et al.* report that no interaction was detected between any Dkk protein and LRP5, as well as no interaction with LDLR, VLDLR, ApoER, or LRP). Additionally, Mao *et al.* demonstrated that LRP6 can titrate Dkk-1's effects of inhibiting Wnt signaling using
25 the commercial TCF-luciferase reporter gene assay (TOPFLASH). A similar conclusion was drawn from analogous studies in *Xenopus* embryos. Deletion analyses of LRP6 functional domains revealed that EGF repeats (beta-propellers) 3 and 4 were necessary for Dkk-1 binding and that the ligand binding domains of LRP6 had no effect on Dkk-1 binding. The findings of Mao *et al.* contrast with data
30 obtained by the present inventors indicating that the ligand binding domains of LRP5

were necessary and sufficient for Dkk-1 binding in yeast. Using classical biochemical ligand-receptor studies, Mao *et al.* determined a $K_d=0.34$ nM for Dkk-1/LRP6 and a $K_d=0.73$ nM for Dkk-2/LRP6.

Semenov *et al.* (2001) verified the Mao group's results and confirmed by coimmunoprecipitation that Dkk-1 does not directly bind to Wnt or Frizzled but rather interacts with LRP6. Their Scatchard analyses found a $K_d=0.5$ nM for Dkk-1/LRP6. Semenov *et al.* also demonstrated that Dkk-1 could abolish an LRP5/Frizzled8 complex implying that Dkk-1 can also repress Wnt signaling via interactions with LRP5. A Dkk-1 mutant where cysteine 220 was changed to alanine abolished LRP6 binding and was unable to repress Wnt signaling. Studies in *Xenopus* embryos confirmed the results and revealed a functional consequence of Dkk-1/LRP6: repression of Wnt signaling. Their *Xenopus* work also suggested that LRP6/Dkk-1 may be specific for the canonical, β -catenin-mediated, Wnt pathways as opposed to the Wnt Planar Cell Polarity pathway.

Bafico *et al.* (2001) employed a ^{125}I -labeled Dkk-1 molecule to identify LRP6 as its sole membrane receptor with a $K_d=0.39$ nM. Again, the functional consequences of the Dkk-1/LRP6 interaction was a repression of the canonical Wnt signaling even when Dkk-1 was added at extremely low concentrations (30 pM).

Not wishing to be bound by theory, it is believed that the present invention provides an explanation for the mechanism of Dkk-1 inhibition of the Wnt pathway and provides a mechanism whereby the Wnt pathway may be modulated. The present application and related provisional application 60/291,311 describe Dkk-1/LRP5 interactions and demonstrate that the interaction between LRP5/LRP6/HBM and Dkk can be used in a method as an intervention point in the Wnt pathway for an anabolic bone therapeutic or a modulator of lipid metabolism.

As detailed below, in the section "Methods to Identify Binding Partners" and Examples 6 and 7, Dkk-1 is able to repress LRP5-mediated Wnt signaling but not HBM-mediated Wnt signaling. This observation is of particular interest because the HBM mutation in LRP5 is a gain of function or activation mutation. That is, Wnt signaling, via the canonical pathway, is enhanced with HBM versus LRP5. The

present data suggest the mechanism of this functional activation: the inability of Dkk-1 to repress HBM-mediated Wnt signaling. Further investigations of other Wnt or Dkk family members show differential activities in the canonical Wnt pathway that demonstrate the complexity and variability in Wnt signaling that can be achieved depending on the LRP/Dkk/Wnt/Frizzled repertoire that is expressed in a particular cell or tissue. This may attest to the apparent bone specificity of the HBM phenotype in humans and in the HBM transgenic animals.

Furthermore, the present data reveal the importance and functional consequence for the potential structural perturbation of the first beta-propeller domain of LRP5. Our data identified the ligand binding domain of LRP5 as the interacting region with Dkk-1 while the Mao *et al.* publication demonstrated the functional role of propellers 3 and 4 in their LRP6/Dkk-1 studies. In the present invention, we implicate the first beta propeller domain, via the HBM mutation at residue 171, as having a functional consequence in the Dkk-1-mediated Wnt pathway. The involvement of position 171 of propeller 1 may be direct or indirect with Dkk-1. Direct involvement could arise from perturbations of the 3-dimensional structure of the HBM extracellular domain that render Dkk-1 unable to bind. Alternatively, residue 171 of propeller 1 may directly interact with Dkk-1; however, by itself, it is insufficient to bind and requires other LRP5 domains. Potential indirect candidate molecules may be among the proteins identified the Dkk-1 yeast-two-hybrid experiments.

It may be that the disruption of Dkk activity is not necessarily mediated by enhancing or preventing the binding of Dkk to LRP5/LRP6/HBM. More than one mechanism may be involved. Indeed, the inventors have observed that Dkk-1 binds LRP5, LRP6, and HBM. It is able to effectively inhibit LRP6, and to a slightly lesser extent, LRP5 activity. Further, has been observed that different members of the Dkk family differentially affect LRP5/LRP6/HBM activity. For example, Dkk-1 inhibits LRP5/LRP6/HBM activity while another Dkk may enhance LRP5/LRP6/HBM activity. An endpoint to consider is the modulation of the LRP5/LRP6/HBM activity, not simply binding.

The present disclosure shows that targeting the disruption of the Dkk-1/LRP5 interaction is a therapeutic intervention point for an HBM mimetic agent. A therapeutic agent of the invention may be a small molecule, peptide or nucleic acid aptamer, antibody, or other peptide/protein, etc. Methods of reducing Dkk-1 expression may also be therapeutic using methodologies such as: RNA interference, antisense oligonucleotides, morpholino oligonucleotides, PNAs, antibodies to Dkk-1 or Dkk-1 interacting proteins, decoy or scavenger LRP5 or LRP6 receptors, and knockdown of Dkk-1 or Dkk-1 interactor transcription.

In an embodiment of the present invention, the activity of Dkk-1 or the activity of a Dkk-1 interacting protein may be modulated for example by binding with a peptide aptamer of the present invention. In another embodiment, LRP5 activity may be modulated by a reagent provided by the present invention (e.g., a peptide aptamer). In another embodiment, the Dkk-1/LRP5 interaction may be modulated by a reagent of the present invention (e.g., a Dkk-1 interacting protein such as those identified in Figure 5). In another embodiment, the Wnt signal transduction pathway may be modulated by use of one or more of the above methods. In a preferred embodiment of the present invention, the Dkk-1 mediated activity of the Wnt pathway may be specifically modulated by one or more of the above methods. In another preferred embodiment of the present invention, the Wnt signal transduction pathway may be stimulated by down-regulating Dkk-1 interacting protein activity; such down-regulation could, for example, yield greater LRP5 activity. In a more preferred embodiment, by stimulating LRP5 activity, bone mass regulation may be stimulated to restore or maintain a more optimal level. In another preferred embodiment, by stimulating LRP5 activity, lipid metabolism may be stimulated to restore or maintain a more optimal level. Alternative embodiments provide methods for screening candidate drugs and therapies directed to correction of bone mass disorders or lipid metabolism disorders. And, preferred embodiments of the present invention provide drugs and therapies developed by the use of the reagents and/or methods of the present invention. One skilled in the art will understand that the present invention provides important research tools to develop an effective model of

osteoporosis, to increase understanding of bone mass and lipid modulation, and to modulate bone mass and lipid metabolism.

Previous investigation of a large family in which high bone mass is inherited as a single gene (autosomal dominant) trait (HBM-1) has provided important insight into the mechanism by which bone density might be modulated. Members of this family have significantly increased spinal and hip BMD (>3 standard deviations above the norm) which affects young adults as well as elderly family members into the ninth decade. The bones of affected members, while appearing very dense radiographically, have normal external shape and outer dimensions. Cortical bone is thickened on endosteal surfaces and "affected" individuals are asymptomatic without any other phenotypic abnormalities. Assays of biochemical markers that reflect skeletal turnover suggest that the disorder is associated with a normal rate of bone remodeling. Affected individuals have achieved a balance in bone turnover at a density that is significantly greater than necessary for normal skeletal stresses. Importantly, the bones most affected are load-bearing bones which are subjected to the greatest mechanical and gravitational stresses (spine and hip). These are the most important bones to target for therapeutic interventions in osteoporosis. The gene identified as being responsible for this phenotype, Zmax or LRP5, was not previously associated with bone physiology. The fact that modification of this gene, such as that produced by the polymorphism leading to the autosomal dominant inheritance of the HBM family phenotype, identifies Zmax/LRP5 and the pathway by which it is regulated, including Dkk/Wnt pathways discussed above, as an important target for developing modulators of bone density. Modulation of Zmax/LRP5 to mimic the gain in function provided by the HBM polymorphism would be expected to provide an important therapy for bone wasting conditions. Additionally, such modulation in young adults could enhance peak bone mass and prevent or delay fracture risk later in life. Alternatively, modulation to reduce function could be employed to treat conditions where bone is being inappropriately produced.

3. Polypeptides

Polypeptides contemplated for use in this invention include those which modulate Dkk and Dkk interacting protein activities. Preferred polypeptides and peptides include those which modulate the Wnt pathway. Examples of preferred sequences include the Y2H baits exemplified in Figure 2, peptide aptamers of Figure 3 (SEQ ID NOs:171-188) and Figure 4 (SEQ ID NOs:189-192), the polypeptides of the Dkk-1 interacting proteins identified in Figure 5, those polypeptides shown in Figure 6, the LRP binding domain of Dkk (amino acids 138-266 of hDkk1), the cysteine-rich domain 2 (a.a. 183-245 of hDkk-1), the cysteine-rich domain 1 (a.a. 97-138 of hDkk), and LRP5 binding aptamers of Figure 13 (including SEQ ID NOs:204-213). Although Dkk-1 is exemplified, the other Dkk proteins contain substantially similar regions and may also be used according to the present invention.

For example, the baits depicted in Figure 2 were used in a yeast two hybrid (Y2H) screen. The Y2H screen was performed as described in Example 2 to determine the minimum required binding domain for Dkk-1 to bind LRP5. The minimum binding domain constructs (*i.e.*, residues 139-266 in bold below and residues 97-245 which are underlined, of Dkk-1) include the second cysteine rich domain which has sequence homology to a colipase fold.

mmalgaagat rvfvamva aa lgghp1lgvs atlnsvlnsn aiknlppplg gaaghpgsav 60
saapgilypg gnkyqtidny qpypcaedee cgtdeycasp trggdagvqi clacrkrkr 120
cmrhamccpg nyckngicvs sdqnhfrgei eetitesfgn dhstldgyar rttlsskmyh 180
tkgqegsvcl rssdcasqlc carhfwsklc kplvkeggvc tkhrrkgshq leifqrcycq 240
eglscrigkd hhqasnssrl htcqrh (GenBank Accession No. XP_005730) (SEQ ID NO:128).

This homology suggests a lipid-binding function and may facilitate Dkk-1 interactions at the plasma membrane (van Tilbeurgh, H., *Biochim. Biophys. Acta.* 1441:173-84 (1999)). An interaction domain of Dkk-1 that is able to interact with the ligand binding domain (LBD) of LRP5 is a useful reagent in the modulation of LRP5 activity

and modulation of Dkk-1/LRP5 complex formation. Similar screens can be prepared for Dkk-1 and Dkk-1 interacting proteins or polypeptides.

A set of peptide aptamers was identified from a library of random peptides constrained and presented in a thioredoxin A (trxA) scaffold as described in Example 3. Peptide aptamers are powerful new tools for molecular medicine as reviewed by Hoppe-Seyler & Butz, *J. Mol. Med.*, 78:426-430 (2000); Brody and Gold, *Rev. Mol. Biotech.*, 74:5-13 (2000); and Colas, *Curr. Opin. in Chem. Biol.* 4:54-9 (2000) and the references cited therein. Briefly, peptide aptamers have been shown to be highly specific reagents capable of binding *in vivo*. As such, peptide aptamers provide a method of modulating the function of a protein and may serve as a substitute for conventional knock-out methods, knock-down or complete loss of function. Peptide aptamers are also useful reagents for the validation of targets for drug development and may be used as therapeutic compounds directly or provide the necessary foundation for drug design. Once identified, the peptide insert may be synthesized and used directly or incorporated into another carrier molecule. References reviewed and cited by Brody and Gold (2000, *supra*) describe demonstrated therapeutic and diagnostic applications of peptide aptamers and would be known to the skilled artisan.

The peptide aptamers of the present invention are useful reagents in the binding of Dkk-1 to its ligands and thereby modulation of the Wnt pathway and may be used to prevent Dkk-1 from inhibiting LRP5 modulation or Dkk-1 interacting protein modulation of the Wnt pathway. The sequence of these peptide aptamers is shown in Figure 3 (SEQ ID NOs:171-188). The peptide aptamers refers to the peptide constrained by the thioredoxin scaffold. The aptamers are also contemplated as therapeutic agents to treat Dkk-1 mediated diseases and conditions. Such aptamers are useful structural guides to chemists, for the design of mimetic compounds of the aptamers.

Peptide aptamers were likewise developed to the LRP5 ligand binding domain (LBD) bait sequences. The sequences of these peptide aptamers is shown in Figure 4 (SEQ ID NOs:189-192). These are useful reagents which may be used to disrupt

the Dkk-1/LRP5 binding interface while leaving Dkk-1 undisturbed. These can be used as comparative controls for Wnt signaling, thus, a control is provided for the specificity of any drug or therapy screened. The aptamers are also useful therapeutic agents to treat LRP mediated diseases and conditions. Such aptamers may also be used as structural guides to chemists, for the design of mimetic compounds of the aptamers.

Thirty proteins were identified which interact with Dkk-1, Dkk-1 interacting proteins, were identified in a yeast-two-hybrid screen using the Dkk-1 bait and are shown in Figure 5. It was noted that these results suggest an interaction of Dkk-1 with Notch-2. It has been suggested that cross-talk exists between the Wnt and Notch signaling pathways. For instance, Presenilin1 (Ps1) is required for Notch processing and inhibits the downstream Wnt pathway. The extracellular domain of Notch is thought to interact with Wnt. Furthermore, the Notch intracellular domain is thought to interact with disheveled and in signal induced processing, the intracellular domain is thought to interact with presenilin. (Soriano *et al.*, *J. Cell Biol.* 152:785-94 (2001)). For additional information regarding the relationships between Notch and Wnt signaling, see Wesley, *Mol. Cell. Biol.* 19:5743-58 (1999) and Axelrod *et al.*, *Science* 271:1826-32 (1996).

An interaction between Dkk-1 and chordin has also been noted; suggesting that cross-talk exists between the Wnt and TGF-beta/BMP signaling pathways (Letamendia *et al.*, *J. Bone Joint Surg. Am.* 83A:S31 (2001); Labbe *et al.*, *Proc. Natl. Acad. Sci. USA* 97:8358-63 (2000); Nishita *et al.*, *Nature* 403:781-5 (2000); DeRobertis *et al.*, *Int. J. Dev. Biol.* 45:1389-97 (2001); and Saint-Jeannet *et al.*, *Proc. Natl. Acad. Sci. USA* 94:13713-8 (1997)). The BMP signaling pathway has an established role in bone and connective tissue development, repair and homeostasis (review in Rosen and Wozney "Bone Morphogenetic Proteins" In: *Principles of Bone Biology*, 2nd Edition, Eds. J. Bilezikian, L. Raisz and G. Rodan, Academic Press, pp. 919-28 (2002)). Chordin is an important molecule during development which also modulates BMP signaling in adults by sequestering BMPs in latent complexes (Piccolo *et al.*, *Cell* 86:589-98 (1996) reviewed in Reddi, *Arthritis Res.* 3:1-5 (2001);

DeRobertis *et al.*, *Int. J. Dev. Biol.* 45:189-97 (2001)). It may be that Dkk effects bone mass modulation through both the Wnt signaling pathway via LRP and the BMP pathway via chordin.

Moreover, a number of putative growth factors, growth factor related proteins, and extracellular matrix proteins have been identified as Dkk-1 interacting proteins. Additional information regarding Dkk-1 interacting proteins identified in the Y2H assay may be obtained from publicly available databases such as PubMed via the use of the accession numbers provided in the present application. In a preferred embodiment of the invention, the amino acid sequences of these Dkk-1 interacting proteins or biologically active fragments thereof be used to modulate Dkk, Dkk-1, LRP5, LRP6, HBM, or Wnt activity. Although these proteins were identified as interacting with Dkk-1, due to the substantial homology between the various Dkk proteins, such interacting proteins are contemplated to interact with the other Dkk family members.

4. Aptamer Mimetics

The present invention further provides for mimetics of Dkk, particularly Dkk-1, and LRP5 peptide aptamers. Such aptamers may serve as structural guides to chemists for the design of mimetic compounds of the aptamers. The aptamers and their mimetics are useful as therapeutic agents to treat LRP- or Dkk-mediated diseases and conditions.

5. Nucleic Acid Molecules

The present invention further provides nucleic acid molecules that encode polypeptides and proteins which interact with Dkk and Dkk interacting proteins, and/or LRP5 (also LRP6 and HBM) to modulate biological activities of these proteins. Preferred embodiments provide nucleic acids encoding for fragments of Dkk-1 protein, including the nucleic acids of Figure 7, the Dkk-1 interacting proteins listed in Figure 5, polypeptide aptamers of Dkk-1 (Figure 3 - SEQ ID NOs:171-188), LRP5 (Figure 4 - SEQ ID NOs:189-192), Figure 13 peptide aptamers (including SEQ

ID NO:204-214) encoded by Figure 12 polynucleotides (including SEQ ID NO:193-203), LRP6 and HBM and the related fusion proteins herein described, preferably in isolated or purified form. As used herein, "nucleic acid" is defined as RNA, DNA, or cDNA that encodes a peptide as defined above, or is complementary to a nucleic acid sequence encoding such peptides, or hybridizes to either the sense or antisense strands of the nucleic acid and remains stably bound to it under appropriate stringency conditions. The nucleic acid may encode a polypeptide sharing at least about 75% sequence identity, preferably at least about 80%, and more preferably at least about 85%, with the peptide sequences; at least about 90%, 95%, 96%, 97%, 98%, and 99% or greater are also contemplated. Specifically contemplated are genomic DNA, cDNA, mRNA, antisense molecules, enzymatically active nucleic acids (e.g., ribozymes), as well as nucleic acids based on an alternative backbone or including alternative bases, whether derived from natural sources or synthesized. Such hybridizing or complementary nucleic acids, however, are defined further as being novel and nonobvious over any prior art nucleic acid including that which encodes, hybridizes under appropriate stringency conditions, or is complementary to a nucleic acid encoding a protein according to the present invention.

As used herein, the terms "hybridization" (hybridizing) and "specificity" (specific for) in the context of nucleotide sequences are used interchangeably. The ability of two nucleotide sequences to hybridize to each other is based upon the degree of complementarity of the two nucleotide sequences, which in turn is based on the fraction of matched complementary nucleotide pairs. The more nucleotides in a given sequence that are complementary to another sequence, the greater the degree of hybridization of one to the other. The degree of hybridization also depends on the conditions of stringency which include temperature, solvent ratios, salt concentrations, and the like. In particular, "selective hybridization" pertains to conditions in which the degree of hybridization of a polynucleotide of the invention to its target would require complete or nearly complete complementarity. The complementarity must be sufficiently high so as to assure that the polynucleotide of

the invention will bind specifically to the target nucleotide sequence relative to the binding of other nucleic acids present in the hybridization medium. With selective hybridization, complementarity will be about 90-100%, preferably about 95-100%, more preferably about 100%.

5 "Stringent conditions" are those that (1) employ low ionic strength and high temperature for washing, for example: 0.015 M NaCl, 0.0015 M sodium titrate, 0.1% SDS at 50°C; or (2) employ during hybridization a denaturing agent such as formamide, for example, 50% (vol/vol) formamide with 0.1% bovine serum albumin, 0.1% Ficoll, 0.1% polyvinylpyrrolidone, 50 mM sodium phosphate buffer at pH 6.5
10 with 750 mM NaCl, 75 mM sodium citrate at 42°C. Another example is use of 50% formamide, 5X SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5X Denhardt's solution, sonicated salmon sperm DNA (50 µg/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2X SSC and 0.1% SDS. A skilled artisan can readily determine
15 and vary the stringency conditions appropriately to obtain a clear and detectable hybridization signal.

 As used herein, a nucleic acid molecule is said to be "isolated" or "purified" when the nucleic acid molecule is substantially separated from contaminant nucleic acid encoding other polypeptides from the source of nucleic acid. Isolated or purified
20 is also meant to include nucleic acids which encode Dkk or fragments thereof which lack surrounding genomic sequences that flank the *Dkk* gene. Isolated or purified is further intended to include nucleic acids which encode Dkk interacting proteins or biologically active fragments thereof which lack surrounding genomic sequences that flank the Dkk interacting protein genes.

25 The present invention further provides fragments of the encoding nucleic acid molecule. As used herein, a fragment of an encoding nucleic acid molecule refers to a small portion of the entire protein encoding sequence. The size of the fragment will be determined by the intended use. For example, if the fragment is chosen so as to encode an active portion of the protein, the fragment will need to be large enough
30 to encode the functional region(s) of the protein. If the fragment is to be used as a

nucleic acid probe or PCR primer, then the fragment length is chosen so as to obtain a relatively small number of false positives during probing/priming.

Fragments of the encoding nucleic acid molecules of the present invention (*i.e.*, synthetic oligonucleotides) that are used as probes or specific primers for the polymerase chain reaction (PCR), or to synthesize gene sequences encoding proteins of the invention can easily be synthesized by chemical techniques, for example, the phosphotriester method of Matteucci *et al.* (*J. Am. Chem. Soc.* 103:3185-3191 (1981)) or using automated synthesis methods. In addition, larger DNA segments can readily be prepared by well known methods, such as synthesis of a group of oligonucleotides that define various modular segments of the gene, followed by ligation of oligonucleotides to build the complete modified gene.

The polypeptide encoding nucleic acid molecules of the present invention may further be modified to contain a detectable label for diagnostic and probe purposes. A variety of such labels are known in the art and can readily be employed with the encoding molecules herein described. Suitable labels include, but are not limited to, biotin, radiolabeled nucleotides and the like. A skilled artisan can employ any of the art known labels to obtain a labeled encoding nucleic acid molecule.

Modifications to the primary structure itself by deletion, addition, or alteration of the amino acids incorporated into the protein sequence during translation can be made without destroying the activity of the protein. Such substitutions or other alterations result in proteins having an amino acid sequence encoded by a nucleic acid falling within the contemplated scope of the present invention.

Antisense molecules corresponding to the polypeptide coding or complementary sequence may be prepared. Methods of making antisense molecules which bind to mRNA, form triple helices or are enzymatically active and cleave TSG RNA and single stranded DNA (ssDNA) are known in the art. See, *e.g.*, Antisense and Ribozyme Methodology: Laboratory Companion (Ian Gibson, ed., Chapman & Hall, 1997) and Ribozyme Protocols: Methods in Molecular Biology (Phillip C. Turner, ed., Humana Press, Clifton, NJ, 1997).

Also contemplated is the use of compounds which mediate postranscriptional gene silencing (PTGS), quelling and RNA interference (RNAi). These compounds typically are about 21 to about 25 nucleotides and are also known as short interfering RNAs or short inhibitory RNAs (siRNAs). The siRNAs are produced from an initiating double stranded RNA (dsRNA). Although the full mechanism by which the siRNAs function is not fully elucidated, it is known that these siRNAs transform the target mRNA into dsRNA, which is then degraded. Preferred forms are 5' phosphorylated siRNAs, however, hydroxylated forms may also be utilized. For additional background regarding the preparation and mechanism of siRNAs generally, see, e.g., Lipardi *et al.*, *Cell* 107(3): 297-307 (2001); Boutla *et al.*, *Curr. Biol.* 11(22): 1776-80 (2001); Djikeng *et al.*, *RNA* 7(11): 1522-30 (2001); Elbashir *et al.*, *EMBO J.* 20(23): 6877-88 (2001); Harborth *et al.*, *J. Cell. Sci.* 114(Pt. 24): 4557-65 (2001); Hutvagner *et al.*, *Science* 293(5531): 811-3 (2001); and Elbashir *et al.*, *Nature* 411:494-98 (2001).

Also contemplated are short hairpin RNAs (shRNAs). shRNAs are a modification of the siRNA method described above. Instead of transfecting exogenously synthesized dsRNA into a cell, sequence-specific silencing can be achieved by stably expressing siRNA from a DNA template as a fold-back stem-loop, or hairpin. This approach is known as shRNA. This method permits the analysis of loss of function phenotypes due to sequence-specific gene silencing in mammalian cells by avoiding many of the problems associated with siRNAs, such as RNase degradation of the reagents, expensive chemical synthesis, etc. For additional background regarding the preparation and mechanism of shRNAs generally, see, e.g., Yu *et al.*, *PNAS* 99:6047-6052 (2002); Paddison *et al.*, *Genes and Devel.* 16:948-58 (2002); and Brummelkamp *et al.*, *Science* 296:550-553 (2002). For additional background on the use of this method in mammalian gene knockdown methodologies, see Tuschl, *Nature Biotech.* 20:446-448 (2002) (and references therein).

In one preferred embodiment, the siRNA or shRNA is directed to a Dkk encoding mRNA, wherein a preferred Dkk is Dkk-1. In another embodiment, the

siRNA or shRNA is directed towards a protein which binds to and modulates the activity of or is modulated by a Dkk; these proteins include LRP5, LRP6 and HBM as well as other members of the Wnt pathway.

5 **6. Isolation of Other Related Nucleic Acid Molecules**

 The identification of the nucleic acid molecule of Dkk allows a skilled artisan to isolate nucleic acid molecules that encode other members of the Dkk family (see, Krupnik *et al.*, 1999). Further, the presently disclosed nucleic acid molecules allow a skilled artisan to isolate nucleic acid molecules that encode Dkk-1-like proteins, in
10 addition to Dkk-1. The presently disclosed Dkk-1 interacting proteins and their corresponding nucleic acid molecules allows a skilled artisan to further isolate other related protein family members which interact with Dkk-1.

 A skilled artisan can readily use the amino acid sequence of Dkk and Dkk interacting proteins to generate antibody probes to screen expression libraries prepared from appropriate cells. Typically, polyclonal antiserum from mammals such
15 as rabbits immunized with the purified protein (as described below) or monoclonal antibodies can be used to probe a mammalian cDNA or genomic expression library, such as a human macrophage library, to obtain the appropriate coding sequence for other members of the protein family. The cloned cDNA sequence can be expressed as
20 a fusion protein, expressed directly using its own control sequences, or expressed by constructions using control sequences appropriate to the particular host used for expression of the desired protein.

 Alternatively, a portion of the coding sequence herein described can be synthesized and used as a probe to retrieve DNA encoding a member of the protein
25 family from any mammalian organism. Oligomers containing approximately 18-20 nucleotides (encoding about a 6-7 amino acid stretch) are prepared and used to screen genomic DNA or cDNA libraries to obtain hybridization under stringent conditions or conditions of sufficient stringency to eliminate an undue level of false positives.

 Additionally, pairs of oligonucleotide primers can be prepared for use in a
30 polymerase chain reaction (PCR) to selectively clone an encoding nucleic acid

molecule. A PCR denature/anneal/extend cycle for using such PCR primers is well known in the art and can readily be adapted for use in isolating other encoding nucleic acid molecules. For example, degenerate primers can be utilized to obtain sequences related to Dkk-1 or Dkk-1 interacting proteins. Primers can be designed that are not perfectly complementary and can still hybridize to a portion of a target sequence or flanking sequence and thereby provide for amplification of all or a portion of a target sequence. Primers of about 20 nucleotides or less, preferably have about one to three mismatches located at the 5' and/or 3' ends. Primers of about 20 to 30 nucleotides have up to about 30% mismatches and can still hybridize to a target sequence.

Hybridization conditions for primers with mismatch can be determined by the method described in Maniatis *et al.*, Molecular Cloning: A Laboratory Manual (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1982) or by reference to known methods. The ability of the primer to hybridize to a sequence of either Dkk-1, a Dkk-1 interacting protein, or a related sequence under varying conditions can be determined using this method. Because a target sequence is known, the effect of mismatches can be determined by methods known to those of skill in the art. Degenerate primers would be based on putative conserved amino acid sequences of the Dkk-1 and Dkk-1 interacting protein genes.

7. rDNA Molecules for Polypeptide Expression

The present invention further provides recombinant DNA molecules (rDNAs) that contain a polypeptide coding sequence. As used herein, a rDNA molecule is a DNA molecule that has been subjected to molecular manipulation *in situ*. Methods for generating rDNA molecules are well known in the art, for example, see Sambrook *et al.*, Molecular Cloning: A Laboratory Manual (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989). In the preferred rDNA molecules, a coding DNA sequence is operably linked to expression control sequences and/or vector sequences.

The choice of vector and/or expression control sequences to which one of the protein family encoding sequences of the present invention is operably linked depends directly, as is well known in the art, on the functional properties desired, e.g., protein

expression, and the host cell to be transformed. A vector contemplated by the present invention is at least capable of directing the replication and/or insertion into the host chromosome, and preferably also expression, of the structural gene included in the rDNA molecule.

5 Expression control elements that are used for regulating the expression of an operably linked protein encoding sequence are known in the art and include, but are not limited to, inducible promoters, constitutive promoters, secretion signals, and other regulatory elements. Preferably, the inducible promoter is readily controlled, such as being responsive to a nutrient in the host cell's medium. Preferred promoters include
10 yeast promoters, which include promoter regions for metallothionein, 3-phosphoglycerate kinase or other glycolytic enzymes such as enolase or glyceraldehyde-3-phosphate dehydrogenase, enzymes responsible for maltose and galactose utilization, and others. Vectors and promoters suitable for use in yeast expression are further described in EP 73,675A. Appropriate non-native mammalian
15 promoters might include the early and late promoters from SV40 (Fiers et al, *Nature*, 273:113 (1978)) or promoters derived from Moloney murine leukemia virus, mouse tumor virus, avian sarcoma viruses, adenovirus II, bovine papilloma virus or polyoma. In addition, the construct may be joined to an amplifiable gene (e.g., DHFR) so that multiple copies of the gene may be made. For appropriate enhancer and other
20 expression control sequences, see also Enhancers and Eukaryotic Gene Expression (Cold Spring Harbor Press, Cold Spring Harbor, NY, 1983). Preferred bone related promoters include CMVbActin or type I collagen promoters to drive expression of the human HBM, Zmax1/LRP5 or LRP6 cDNA. Other preferred promoters for mammalian expression are from cytomegalovirus (CMV), Rous sarcoma virus (RSV), Simian virus
25 40 (SV40), and EF-1a (human elongation factor 1a-subunit).

 In one embodiment, the vector containing a coding nucleic acid molecule will include a prokaryotic replicon, *i.e.*, a DNA sequence having the ability to direct autonomous replication and maintenance of the recombinant DNA molecule extrachromosomally in a prokaryotic host cell, such as a bacterial host cell, transformed
30 therewith. Such replicons are well known in the art. In addition, vectors with a

prokaryotic replicon may also include a gene whose expression confers a detectable marker such as a drug resistance. Typical bacterial drug resistance genes are those that confer resistance to ampicillin or tetracycline.

5 Vectors that include a prokaryotic replicon can further include a prokaryotic or bacteriophage promoter capable of directing the expression (transcription and translation) of the coding gene sequences in a bacterial host cell, such as *E. coli*. A promoter is an expression control element formed by a DNA sequence that permits binding of RNA polymerase and transcription to occur. Promoter sequences compatible with bacterial hosts are typically provided in plasmid vectors containing
10 convenient restriction sites for insertion of a DNA segment of the present invention. Typical of such vector plasmids are pUC8, pUC9, pBR322 and pBR329 available from Biorad Laboratories, (Richmond, CA), and pPL and pKK223 available from Pharmacia (Piscataway, NJ).

15 Expression vectors compatible with eukaryotic cells, preferably those compatible with vertebrate cells, can also be used to form a rDNA molecule that contains a coding sequence. Eukaryotic cell expression vectors are well known in the art and are available from several commercial sources. Typically, such vectors are provided containing convenient restriction sites for insertion of a desired DNA segment. Typical of such vectors are pSVL and pKSV-10 (Pharmacia), pBPV-1/pML2d (International
20 Biotechnologies, Inc.), vector systems that include Histidine Tags and periplasmic secretion, or other vectors described in the art.

Eukaryotic cell expression vectors used to construct the rDNA molecules of the present invention may further include a selectable marker that is effective in an eukaryotic cell, preferably a drug resistance selection marker. A preferred drug
25 resistance marker is the gene whose expression results in neomycin resistance, *i.e.*, the neomycin phosphotransferase (*neo*) gene (Southern *et al.*, *J. Mol. Anal. Genet.* 1:327-341 (1982)). Alternatively, the selectable marker can be present on a separate plasmid, and the two vectors introduced by co-transfection of the host cell, and selected by culturing in the appropriate drug for the selectable marker.

8. Host Cells Containing an Exogenously Supplied rDNA Nucleic Acid Molecule

The present invention further provides host cells transformed with a nucleic acid molecule that encodes a polypeptide or protein of the present invention. The host cell can be either prokaryotic or eukaryotic. Eukaryotic cells useful for expression of a protein of the invention are not limited, so long as the cell line is compatible with cell culture methods and compatible with the propagation of the expression vector and expression of the gene product. Preferred eukaryotic host cells include, but are not limited to, yeast, insect and mammalian cells, preferably vertebrate cells such as those from a mouse, rat, monkey or human cell line but also can include invertebrates with, for example, cartilage. Preferred eukaryotic host cells include but are not limited to Chinese hamster ovary (CHO) cells (ATCC No. CCL61), NIH Swiss mouse embryo cells NIH/3T3 (ATCC No. CRL 1658), baby hamster kidney cells (BHK), HOB-03-CE6 osteoblast cells, and other like eukaryotic tissue culture cell lines.

Any prokaryotic host can be used to express a rDNA molecule encoding a protein of the invention. A preferred prokaryotic host is *E. coli*.

Transformation of appropriate cell hosts with a recombinant DNA (rDNA) molecule of the present invention is accomplished by well known methods that typically depend on the type of vector used and host system employed. With regard to transformation of prokaryotic host cells, electroporation and salt treatment methods are typically employed; see, for example, Cohen *et al.*, *Proc. Natl. Acad. Sci. USA* 69: 2110 (1972); Maniatis *et al.* (1982); and Sambrook *et al.* (1989). With regard to transformation of vertebrate cells with vectors containing rDNAs, electroporation, cationic lipid or salt treatment methods are typically employed; see, for example, Graham *et al.*, *Viol.* 52: 456 (1973); Wigler *et al.*, *Proc. Natl. Acad. Sci. USA* 76: 1373-76 (1979).

Successfully transformed cells, *i.e.*, cells that contain a rDNA molecule of the present invention, can be identified by well known techniques including the selection for a selectable marker. For example, cells resulting from the introduction of an rDNA of the present invention can be cloned to produce single colonies. Cells from those

colonies can be harvested, lysed and their DNA content examined for the presence of the rDNA using a method such as that described by Southern, *J. Mol. Biol.* 98: 503 (1975), or Berent *et al.*, *Biotech.* 3: 208 (1985). Alternatively, the cells can be cultured to produce the proteins encoded by the rDNA and the proteins harvested and assayed, using for example, any suitable immunological method. See, e.g., Harlow *et al.*, (1988).

Recombinant DNA can also be utilized to analyze the function of coding and non-coding sequences. Sequences that modulate the translation of the mRNA can be utilized in an affinity matrix system to purify proteins obtained from cell lysates that associate with the Dkk-1 or Dkk-1 interacting protein or expression control sequence. Synthetic oligonucleotides would be coupled to the beads and probed with the lysates, as is commonly known in the art. Associated proteins could then be separated using, for example, a two dimensional SDS-PAGE system. Proteins thus isolated could be further identified using mass spectroscopy or protein sequencing. Additional methods would be apparent to the skilled artisan.

9. Production of Recombinant Peptides and Proteins using a cDNA or Other Recombinant Nucleic Acids

The invention also relates to nucleic acid molecules which encode a Dkk protein and polypeptide fragments thereof, and proteins and polypeptides which bind to Dkk (e.g., LRP5, LRP6 and HBM, Dkk interacting proteins such as the proteins of Figure 5) and molecular analogues. The polypeptides of the present invention include the full length Dkk and polypeptide fragments thereof, Dkk binding proteins and polypeptides thereof. Preferably these proteins are mammalian proteins, and most preferably human proteins and biologically active fragments thereof. Alternative embodiments include nucleic acid molecules encoding polypeptide fragments having a consecutive amino acid sequence of at least about 3, 5, 7, 8, 9, 10, 15, 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, 125, 150, 175, or 200 amino acid residues from a common polypeptide sequence; amino acid sequence variants of a common polypeptide sequence wherein an amino acid residue has been inserted N- or C-terminal to, or within, the polypeptide sequence or its fragments; and amino acid sequence variants of the common

polypeptide sequence or its fragments, which have been substituted by another conserved residue. Recombinant nucleic acid molecules which encode polypeptides include those containing predetermined mutations by, e.g., homologous recombination, site-directed or PCR mutagenesis, and recombinant Dkk proteins or polypeptide fragments of other animal species, including but not limited to vertebrates (e.g., rabbit, rat, murine, porcine, camelid, reptilian, caprine, avian, fish, bovine, ovine, equine and non-human primate species) as well as invertebrates, and alleles or other naturally occurring variants and homologs of Dkk binding proteins of the foregoing species and of human sequences. Also contemplated herein are derivatives of the commonly known Dkk, Dkk interacting proteins, or fragments thereof, wherein Dkk, Dkk interacting proteins, or their fragments have been covalently modified by substitution, chemical, enzymatic, or other appropriate means with a moiety other than a naturally occurring amino acid (for example a detectable moiety such as an enzyme or radioisotope) and soluble forms of Dkk. It is further contemplated that the present invention also includes nucleic acids with silent mutations which will hybridize to the endogenous sequence and which will still encode the same polypeptide.

The nucleic acid molecules encoding Dkk binding proteins, the LRP5 binding domain fragment of Dkk, or other polypeptides of the present invention are preferably those which share a common biological activity (e.g., mediate Dkk activity such as its interaction with LRP5, HBM or LRP6). The polypeptides of the present invention include those encoded by a nucleic acid molecule with silent mutations, as well as those nucleic acids encoding a biologically active protein with conservative amino acid substitutions, allelic variants, and other variants of the disclosed polypeptides which maintain at least one Dkk activity.

The amino acid compounds of the invention are polypeptides which are partially defined in terms of amino acid residues of designated classes. Polypeptide homologs would include conservative amino acid substitutions within the amino acid classes described below. Amino acid residues can be generally sub-classified into four major subclasses as follows:

Acidic: The residue has a negative charge due to loss of H^+ ion at physiological pH, and the residue is attracted by aqueous solution so as to seek the surface positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium, at physiological pH.

5 Basic: The residue has a positive charge due to association with H^+ ion at physiological pH, and the residue is attracted by aqueous solution so as to seek the surface positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium at physiological pH.

10 Neutral/non-polar: The residues are not charged at physiological pH, but the residue is repelled by aqueous solution so as to seek the inner positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium. These residues are also designated "hydrophobic."

15 Neutral/polar: The residues are not charged at physiological pH, but the residue is attracted by aqueous solution so as to seek the outer positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium.

20 It is understood, of course, that in a statistical collection of individual residue molecules some molecules will be charged, and some not, and there will be an attraction for or repulsion from an aqueous medium to a greater or lesser extent. To fit the definition of "charged", a significant percentage (at least approximately 25%) of the individual molecules are charged at physiological pH. The degree of attraction or repulsion required for classification as polar or nonpolar is arbitrary and, therefore, amino acids specifically contemplated by the invention have been classified as one or the other. Most amino acids not specifically named can be classified on the basis of known behavior.

25 Amino acid residues can be further subclassified as cyclic or noncyclic, and aromatic or non-aromatic, self-explanatory classifications with respect to the side chain substituent groups of the residues, and as small or large. The residue is considered small if it contains a total of 4 carbon atoms or less, inclusive of the carboxyl carbon. Small residues are, of course, always nonaromatic.

The gene-encoded secondary amino acid proline, although technically within the group neutral/nonpolar/large/cyclic and nonaromatic, is a special case due to its known effects on the secondary conformation of peptide chains, and is not, therefore, included in this defined group.

5 Other amino acid substitutions of those encoded in the gene can also be included in peptide compounds within the scope of the invention and can be classified within this general scheme according to their structure.

All of the compounds of the invention may be in the form of the pharmaceutically acceptable salts or esters. Salts may be, for example, Na^+ , K^+ , Ca^{+2} , Mg^{+2} and the like;
10 the esters are generally those of alcohols of 1-6 carbons.

The present invention further provides methods for producing a protein of the invention using nucleic acid molecules herein described. In general terms, the production of a recombinant form of a protein typically involves the following steps.

First, a nucleic acid molecule is obtained that encodes Dkk, such as a nucleic acid molecule encoding human Dkk or any other Dkk sequence, or that encodes a Dkk binding protein, a Dkk aptamer or a biologically active fragment thereof. Particularly for Dkk binding peptides, the nucleotides encoding the peptide are incorporated into a nucleic acid in the form of an in-frame fusion, insertion into or appended to a thioredoxin coding sequence. The coding sequence (ORF) is directly suitable for expression in any
15 host, as it is not interrupted by introns.

These DNAs can be transfected into host cells such as eukaryotic cells or prokaryotic cells. Eukaryotic hosts include mammalian cells and vertebrate (e.g., osteoblasts, osteosarcoma cell lines, Drosophila S2 cells, hepatocytes, tumor cell lines and other bone cells of any mammal, as well as insect cells, such as Sf9 cells using
25 recombinant baculovirus). For example, a DNA expressing an open reading frame (ORF) under control of a type I collagen promoter, or such osteoblast promoters as osteocalcin histone, type I collagen, $\text{TGF}\beta 1$, MSX2 , c-fos/cJun and Cbfa1 , can be used to regulate the Dkk in animal cells. Alternatively, the nucleic acid can be placed downstream from an inducible promoter, which can then be placed into vertebrate or
30 invertebrate cells or be used in creating a transgenic animal model.

Alternatively, proteins and polypeptides of the present invention can be expressed in an heterologous system. The human cell line GM637, SV-40 transformed human fibroblasts, can be transfected, with a plasmid containing a Dkk ligand binding domain coding sequence under the control of the chicken actin promoter (Reis *et al.*,
5 EMBO J. 11: 185-193 (1992)). Such transfected cells could be used as a source of Dkk binding domain in functional assays. Alternatively, polypeptides encoding only a portion of Dkk or any of the disclosed Dkk binding peptides Dkk aptamers or a polypeptide encoding a Dkk interacting protein can be expressed alone or in the form of a fusion protein. For example, Dkk derived peptides can be expressed in bacteria (e.g.,
10 *E. coli*) as GST- or His-Tag fusion proteins. These fusion proteins are then purified and can be used to generate polyclonal antibodies or can be used to identify other Dkk ligands.

The nucleic acid coding sequence is preferably placed in operable linkage with suitable control sequences, as described above, to form an expression unit containing
15 the protein encoding open reading frame. The expression unit is used to transform a suitable host and the transformed host is cultured under conditions that allow the production of the recombinant protein. Optionally the recombinant protein is isolated from the medium or from the cells; recovery and purification of the protein may not be necessary in some instances where some impurities may be tolerated.

Each of the foregoing steps can be done in a variety of ways. For example, the
20 desired coding sequences may be obtained from genomic fragments and used directly in appropriate hosts. The construction of expression vectors that are operable in a variety of hosts is accomplished using appropriate replicons and control sequences, as set forth above. The control sequences, expression vectors, and transformation
25 methods are dependent on the type of host cell used to express the gene and were discussed in detail earlier. Suitable restriction sites can, if not normally available, be added to the ends of the coding sequence so as to provide an excisable gene to insert into these vectors. A skilled artisan can readily adapt any host/expression system
30 known in the art for use with the nucleic acid molecules of the invention to produce recombinant protein.

10. Methods to Identify Binding Partners

Another embodiment of the present invention provides methods for use in isolating and identifying binding partners of Dkk or Dkk interacting proteins. Dkk or a Dkk interacting protein or a polypeptide fragment thereof can be mixed with a potential binding partner or an extract or fraction of a cell under conditions that allow the association of potential binding partners with Dkk or with Dkk interacting proteins. After mixing, the peptides, polypeptides, proteins or other molecules that have become associated with Dkk or a Dkk interacting protein are separated from the mixture. The binding partner that bound to the polypeptide then can be purified and further analyzed. Determination of binding partners of Dkk and Dkk interacting proteins as well as agents which prevent the interaction of Dkk with one of its interacting proteins (e.g., LRP5, LRP6, HBM, or those proteins listed in Figure 5) can be performed using a variety of different competition assays as are known in the art. For example, the minimal sequence of Dkk, as described herein, can be used to identify antibodies which compete with LRP5 (or LRP6, HBM or other ligand binding partners) for binding to Dkk-1 and vice versa. The minimal Dkk sequence can be bound to the bottom of a 96-well plate (or other solid substrate), and antibodies or other potential binding agents (e.g., polypeptides, mimetics, homologs, antibody fragments and the like) can be screened in a competition assay to identify agents with binding affinities, for example, greater than the natural ligand binding partner of Dkk.

In the present invention, suitable cells are used for preparing assays, for the expression of a LRP and/or Dkk or proteins that interact therewith. The cells may be made or derived from mammals, yeast, fungi, or viruses. A suitable cell for the purposes of this invention is one that includes but is not limited to a cell that can exhibit a detectable Dkk-LRP (or HBM) interaction, and preferably, the differential interaction between Dkk-1-LRP5 and Dkk-1-HBM. For the desired assay, the cell type may vary. In several embodiments, bone cells are preferred, for example, a human osteoblast cell (e.g. hOB-03-CE6) or osteosarcoma cell (e.g. U2OS). Additional hOB cells are hOB-03-C5, hOB-02-02 and, an immortalized pre-osteocytic cell line referred to as hOB-01-C1-PS-09 cells (which are deposited with American Type Culture Collection in

Manassas, Va. with the designation PTA-785), Examples of osteosarcoma cells would include SaoS2, MG63 and HOS TE85. Immortalized refers to a substantially continuous and permanently established cell culture with substantially unlimited cell division potential. That is, the cells can be cultured substantially indefinitely, i.e., for at least about 6 months under rapid conditions of growth, preferably much longer under slower growth conditions, and can be propagated rapidly and continually using routine cell culture techniques. Alternatively stated, preferred cells can be cultured for at least about 100, 150 or 200 population doublings. These cells produce a complement of proteins characteristic of normal human osteoblastic cells and are capable of osteoblastic differentiation. They can be used in cell culture studies of osteoblastic cell sensitivity to various agents, such as hormones, cytokines, and growth factors, or in tissue therapy. Certain non bone cells such as HEK 293 cells that exhibit detectable Dkk-LRP (or HBM) interaction are also be useful for the assays of this invention.

To identify and isolate a binding partner, the entire Dkk protein (e.g., human Dkk-1, GenBank Accession No. BAA34651) or a Dkk interacting protein (Genbank Accession Nos. for some Dkk-1 interacting proteins are given in Figure 5) can be used. Alternatively, a polypeptide fragment of the protein can be used. Suitable fragments of the protein include at least about 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130, 140, 150 or more contiguous amino acid residues of any Dkk or Dkk interactor sequence. Preferable sequences of Dkk include portions or all of one or both of the cysteine rich domains (e.g., Cys-1 and Cys-2 of Dkk-1) or the conserved sequences at the amino terminus of Dkk-1 (See Krupnik *et al.*, *Gene* 238: 301-313 (1999)). Alternatively, portions of LRP5, LRP6, HBM and other Dkk interacting proteins such as those in Figure 5 that interact with Dkk-1 can be used to identify and isolate agents which modulate Dkk activity. Alternatively, peptide aptamers of LRP5, LRP6, HBM, Dkk and other Dkk interacting proteins such as those in Figure 5 that interact with Dkk-1 can be used to identify and isolate agents which modulate Dkk activity.

As used herein, a cellular extract refers to a preparation or fraction which is made from a lysed or disrupted cell. A variety of methods can be used to obtain cell

extracts. Cells can be disrupted using either physical or chemical disruption methods. Examples of physical disruption methods include, but are not limited to, sonication and mechanical shearing. Examples of chemical lysis methods include, but are not limited to, detergent lysis and enzyme lysis. A skilled artisan can readily adapt methods for preparing cellular extracts in order to obtain extracts for use in the present methods.

Once an extract of a cell is prepared, the extract is mixed with the protein of the invention under conditions in which association of the protein with the binding partner can occur. A variety of conditions can be used, the most preferred being conditions that closely resemble conditions found in the cytoplasm of a human cell. Features such as osmolarity, pH, temperature, and the concentration of cellular extract used, can be varied to optimize the association of the protein with the binding partner.

After mixing under appropriate conditions, the bound complex is separated from the mixture. A variety of techniques can be utilized to separate the mixture. For example, antibodies specific to a protein of the invention can be used to immunoprecipitate the binding partner complex. Alternatively, standard chemical separation techniques such as chromatography and density/sediment centrifugation can be used. For example, a protein of the invention is expressed with an affinity tag such as a His tag. The His labeled protein and any bound molecule may be retained and selectively eluted from a Ni-NTA column.

After removal of non-associated cellular constituents found in the extract, the binding partner can be dissociated from the complex using conventional methods. For example, dissociation can be accomplished by altering the salt concentration or pH of the mixture.

To aid in separating associated binding partner pairs from the mixed extract, the protein of the invention can be immobilized on a solid support. For example, the protein can be attached to a nitrocellulose matrix or acrylic beads. Attachment of the protein to a solid support aids in separating peptide/binding partner pairs from other constituents found in the extract. The identified binding partners can be either a single protein or a complex made up of two or more proteins.

Alternatively, the nucleic acid molecules of the invention can be used in a Y2H system. The Y2H system has been used to identify other protein partner pairs and can readily be adapted to employ the nucleic acid molecules herein described. Methods of performing and using Y2H systems are known. See, e.g., Finley *et al.*, "Two-Hybrid Analysis of Genetic Regulatory Networks," in The Yeast Two-Hybrid System (Paul L. Bartel *et al.*, eds., Oxford, 1997); Meijia Yang, "Use of a Combinatorial Peptide Library in the Two-Hybrid Assay," in The Yeast Two-Hybrid System (Paul L. Bartel *et al.*, eds., Oxford, 1997); Gietz *et al.*, "Identification of proteins that interact with a protein of interest: Applications of the yeast two-hybrid system," *Mol. & Cell. Biochem.* 172: 67-9 (1997); K. H. Young, "Yeast Two-Hybrid: So Many Interactions,(in) so Little Time," *Biol. Reprod.* 58: 302-311 (1998); R. Brent *et al.*, "Understanding Gene and Allele Function with Two-Hybrid Methods," *Annu. Rev. Genet.* 31:663-704 (1997) and U.S. Patent No. 5,989,808. The Dkk-1 interacting proteins identified in Figure 5 were identified using the Y2H interacting system using Dkk-1 as bait.

One preferred *in vitro* binding assay for Dkk modulators would comprise a mixture of a LRP binding domain of Dkk and one or more candidate binding targets or substrates. After incubating the mixture under appropriate conditions, one would determine whether Dkk or a fragment thereof bound with the candidate modulator present. For cell-free binding assays, one or more of the components usually comprises or is coupled to a label. The label may provide for direct detection, such as radioactivity, luminescence, optical or electron density, *etc.*, or indirect detection such as an epitope tag, an enzyme, *etc.* A variety of methods may be employed to detect the label depending on the nature of the label and other assay components. For example, the label may be detected bound to the solid substrate or a portion of the bound complex containing the label may be separated from the solid substrate, and the label thereafter detected. Fluorescence resonance energy transfer may be utilized to monitor the interaction of two labeled molecules. For example, a fluorescence label on Dkk and another label on LRP5 or a soluble fragment thereof such as the extracellular domain will exchange fluorescence resonance energy when in close proximity indicating that the two molecules are bound. A preferred binding partner for Dkk will

increase or decrease the affinity between Dkk and LRP5 which will be readily observable in a fluorescence spectrometer. Alternatively, an instrument, such as a surface plasmon resonance detector manufactured by BIAcore (Uppsala, Sweden), may be used to observe interactions with a fixed target. One skilled in the art knows of many other methods which may be employed for this purpose.

Thereby, the present invention provides methods for screening candidates including polypeptides of the present invention for activity which identifies these candidates as valuable drug leads. Other suitable methods are also known in the art and are suitable for use herein, including *Xenopus* oocyte injection studies and TCF luciferase assays.

Additional assays can be used to identify the activity of Dkk and Dkk interacting proteins in the Wnt pathway, as well as the impact of modulators of Dkk and Dkk interacting proteins on the Wnt pathway. These include, for example, a *Xenopus* embryo assay and a TCF-luciferase reporter gene assay to monitor Wnt signaling modulation.

Xenopus embryos are an informative *in vivo* assay system to evaluate the modulation of Wnt signaling. Ectopic expression of certain Wnts or other activators of the Wnt signaling pathway results in a bifurcation of the anterior neural plate. This bifurcation results in a duplicated body axis, which suggests a role for Wnt signaling during embryonic development (McMahon *et al.*, *Cell* 58: 1075-84 (1989); Sokol *et al.*, *Cell* 67: 741-52 (1991)). Since these original observations, the *Xenopus* embryo assay has been extensively used as an assay system for evaluating modulation of the Wnt signaling pathway. One preferred embodiment of the present invention is demonstrated in Example 6.

Constructs for *Xenopus* expression can be prepared as would be known in the art. For example, a variety of cDNAs have been engineered into the vector pCS2+ (Turner *et al.*, *Genes Devel.* 8: 1434-1447 (1994)) to facilitate the *in vitro* generation of mRNA for use in *Xenopus* embryo injection experiments. DNA inserts are subcloned in the sense orientation with respect to the vector SP6 promoter. Downstream of the insert, the vector provides an SV40 virus polyadenylation signal and a T3 promoter

sequence (*i.e.*, for the generation of antisense mRNA). Constructs can be generated for various Dkk family members, LRP5, LRP6, HBM, Dkk-1 interactors, etc. Constructs could be generated in pCS2⁺ that contain the nucleic acid sequence encoding for the peptide aptamers that were identified in yeast screens. These sequences would be fused to a 5' synthetic translation initiation sequence followed by a canonical signal sequence to ensure that the peptide aptamer would be translated and secreted from the cell.

Once these constructs are made then mRNA can be synthesized and injected into *Xenopus* oocytes. mRNA for microinjection into *Xenopus* embryos is generated by *in vitro* transcription using the cDNA constructs in the pCS2⁺ vector described above as template. Various amounts of RNA can be injected into the ventral blastomere of the 4- or 8-cell *Xenopus* embryo substantially as described in Moon *et al.*, *Technique-J. of Methods in Cell and Mol. Biol.* 1: 76-89 (1989), and Peng, *Meth. Cell. Biol.* 36: 657-62 (1991).

Previous data has shown that expression of LRP5, in the presence of Wnt5a, results in a Wnt-induced duplicated axis formation in *Xenopus* embryos (Tamai *et al.*, *Nature* 407: 530-535 (2000)). The roles of Dkk-1 and Dkk-2, and Dkk-1 interacting proteins, in modulating the LRP5-mediated Wnt response *in vivo* can be analyzed using, for example, the *Xenopus* embryo. In addition, the peptide aptamers, Dkk interacting proteins, or combinations of the above can be evaluated in a similar manner.

Experiments can also be conducted wherein RNA is injected into the dorsal blastomere to ensure the specificity of the observed phenotypes. Lineage tracing experiments can be performed where a marker gene such as green fluorescent protein (GFP) or LacZ is co-injected with the experimental RNAs. Detecting marker gene expression would identify the targeted cells of the microinjection and aid in elucidating the mechanism of action. In addition to the Wnt signaling components listed above, the point at which HBM acts upon the Wnt pathway can also be analyzed. This can be done by co-injections of various dominant-negative constructs. For example, a dominant negative TCF-3 construct would be useful to demonstrate that the observed axis duplication (and Wnt activation) is mediated via the β -catenin-TCF response. If so,

such a construct would be expected to abolish the observed duplicated axis phenotype. Another example would include a dominant negative Dsh construct. Since Dsh is far upstream in the Wnt signaling pathway, a dominant negative construct should abolish the activation of the Wnt response and the observed axis duplication. If it does not, this would suggest that axis duplication is being induced via a different signaling pathway.

The marker genes of the injected *Xenopus* embryos can be analyzed as follows. Representative embryos are collected at stage 10.5 (11 hours post fertilization) for marker gene analysis. RNA is extracted and purified from the embryos following standard protocols (Sambrook *et al.*, 1989 at 7.16). Marker genes could include the following: *Siamois* (*i.e.*, Wnt responsive gene), *Xnr3* (*i.e.*, Wnt responsive gene), *slug* (*i.e.*, neural crest marker), *Xbra* (*i.e.*, early mesoderm marker), *HNK-1* (*i.e.*, ectodermal/neural marker), *endodermin* (*i.e.*, endoderm), *Xlhbox8* (*i.e.*, pancreatic), *BMP2* and *BMP4* (*i.e.*, early mesoderm), *XLRP6* (*i.e.*, maternal and zygotic expression, it is also the LRP6 homolog in the frog), *EF-1* (*i.e.*, control) and *ODC* (*i.e.*, control). Induction of marker genes is analyzed and quantitated by RT-PCR/TaqMan®.

This type of marker analysis is excellent to monitor changes in gene expression that result very early in the embryo as a direct result of signaling perturbation. Other experiments could be designed that would monitor changes in gene expression in a more tissue or spatially-restricted fashion. Examples would include the generation of a transgenic *Xenopus* model. For example, *Zmax/LRP5* and *HBM* expression could be under the control of the *brachyury* or *cardiac-actin* promoters directing gene expression transiently in the mesoderm or in the somites, respectively. Phenotype analyses of these transgenic *Xenopus* animals would include marker gene analysis/transcriptional profiling (from a restricted tissue source) and histologic examination of the tissue.

A TCF-luciferase assay system such as that described in Example 7 can also be used to monitor Wnt signaling activity, Dkk activity and Dkk interacting protein activity. Constructs for the TCF-luciferase assays can be prepared as would be known in the art. For example, Dkk and Dkk interacting protein peptides, *LRP5/LRP6*, among others, can be expressed in *pcDNA3.1*, using Kozak and signal sequences to target peptides for secretion.

Once constructs have been prepared, cells such as osteoblasts and HEK293 cells are seeded in well plates and transfected with construct DNA, CMV beta-galactosidase plasmid DNA, and TCF-luciferase reporter DNA. The cells are then lysed and assayed for beta-galactosidase and luciferase activity to determine whether
5 Dkk, Dkk interacting proteins, or other molecules such as antibodies affect Wnt signaling.

Additional assays for monitoring Wnt signaling activity, Dkk activity, and Dkk interacting protein activity include:

Modulation of another Wnt-responsive transcription factor, LEF, as
10 visualized by a reporter gene activity. One example includes the activation of the LEF1 promoter region fused to the luciferase reporter gene (Hsu *et al.*, *Mol. Cell. Biol.* 18: 4807-18 (1999)).

Alterations in cell proliferation, cell cycle or apoptosis. There are numerous examples describing Wnt-mediated cellular transformations including
15 Shimizu *et al.*, *Cell. Growth Differ.* 8: 1349-58 (1997).

Stabilization and cellular localization of de-phosphorylated β -catenin as an indicator of Wnt activation (Shimizu *et al.*, 1997).

Additional methods of assaying Wnt signaling, through either the canonical or non-canonical pathways, would be apparent to the artisan of ordinary skill.

20
11. Methods to Identify Agents that Modulate the Expression of a Nucleic Acid Encoding the Dkk and/or LRP5 Proteins and/or Dkk interacting proteins

Another embodiment of the present invention provides methods for identifying agents that modulate the expression of a nucleic acid encoding Dkk. Such assays may
25 utilize any available means of monitoring for changes in the expression level of the nucleic acids of the invention. As used herein, an agent is said to modulate the expression of Dkk, if it is capable of up- or down-regulating expression of the nucleic acid in a cell (e.g., mRNA).

In one assay format, cell lines that contain reporter gene fusions between the
30 nucleic acid encoding Dkk (or proteins which modulate the activity of Dkk) and any

assayable fusion partner may be prepared. Numerous assayable fusion partners are known and readily available, including but not limited to the firefly luciferase gene and the gene encoding chloramphenicol acetyltransferase (Alam *et al.*, *Anal. Biochem.* 188: 245-254 (1990)). Cell lines containing the reporter gene fusions are then exposed to the agent to be tested under appropriate conditions and time. Differential expression of the reporter gene between samples exposed to the agent and control samples identifies agents which modulate the expression of a nucleic acid encoding Dkk or other protein which modulates Dkk activity. Such assays can similarly be used to determine whether LRP5 and even LRP6 activity is modulated by regulating Dkk activity.

Additional assay formats may be used to monitor the ability of the agent(s) to modulate the expression of a nucleic acid encoding Dkk, alone or Dkk and LRP5, and/or Dkk interacting proteins such as those identified in Figure 5. For instance, mRNA expression may be monitored directly by hybridization to the nucleic acids of the invention. Cell lines are exposed to the agent to be tested under appropriate conditions and time and total RNA or mRNA is isolated by standard procedures such those disclosed in Sambrook *et al.* (1989); Ausubel *et al.*, Current Protocols in Molecular Biology (Greene Publishing Co., NY, 1995); Maniatis *et al.*, Molecular Cloning: A Laboratory Manual (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1982); and Short Protocols in Molecular Biology: A Compendium of Methods from Current Protocols in Molecular Biology (Frederick M. Ausubel *et al.*, April 1999).

Probes to detect differences in RNA expression levels between cells exposed to the agent and control cells may be prepared from the nucleic acids of the invention. It is preferable, but not necessary, to design probes which hybridize only with target nucleic acids under conditions of high stringency. Only highly complementary nucleic acid hybrids form under conditions of high stringency. Accordingly, the stringency of the assay conditions determines the amount of complementarity which should exist between two nucleic acid strands in order to form a hybrid. Stringency should be chosen to maximize the difference in stability between the probe:target hybrid and potential probe:non-target hybrids.

Probes may be designed from the nucleic acids of the invention through methods known in the art. For instance, the G+C content of the probe and the probe length can affect probe binding to its target sequence. Methods to optimize probe specificity are commonly available. See for example, Sambrook *et al.* (1989) or
5 Ausubel *et al.* (Current Protocols in Molecular Biology, Greene Publishing Co., NY, 1995).

Hybridization conditions are modified using known methods, such as those described by Sambrook *et al.* (1989) and Ausubel *et al.* (1995), as suitable for each probe. Hybridization of total cellular RNA or RNA enriched for polyA RNA can be
10 accomplished in any available format. For instance, total cellular RNA or RNA enriched for polyA RNA can be affixed to a solid support and the solid support exposed to at least one probe comprising at least one, or part of one of the nucleic acid sequences of the invention under conditions in which the probe will specifically hybridize.

Alternatively, nucleic acid fragments comprising at least one, or part of one of the
15 sequences of the invention can be affixed to a solid support, such as a porous glass wafer. The glass or silica wafer can then be exposed to total cellular RNA or polyA RNA from a sample under conditions in which the affixed sequences will specifically hybridize. Such glass wafers and hybridization methods are widely available, for example, those disclosed by Beattie (WO 95/11755). By examining for the ability of a
20 given probe to specifically hybridize to an RNA sample from an untreated cell population and from a cell population exposed to the agent, agents which up- or down-regulate the expression of a nucleic acid encoding Dkk, a Dkk interacting protein, and/or LRP5 can be identified.

Microarray technology and transcriptional profiling are examples of methods
25 which can be used to analyze the impact of putative Dkk or Dkk interacting protein modulating compounds. For transcriptional profiling, mRNA from cells exposed *in vivo* to a potential Dkk modulating agent, such as the Dkk interacting proteins identified in the present invention (e.g., those identified in Figure 5), agents which modulate Dkk interacting proteins, and mRNA from the same type of cells that were not exposed to
30 the agent could be reverse transcribed and hybridized to a chip containing DNA from

numerous genes, to thereby compare the expression of genes in cells treated and not treated with the agent. If, for example a putative Dkk modulating agent down-regulates the expression of Dkk in the cells, then use of the agent may be undesirable in certain patient populations. For additional methods of transcriptional profiling and the use of
5 microarrays, refer to, for example, U.S. Patent No. 6,124,120 issued to Lizardi (2000).

Additional methods for screening the impact of Dkk and Dkk interacting protein modulating compounds or the impact of Dkk or Dkk interacting proteins on modulation of LRP5, LRP6, HBM or the Wnt pathway include the use of TaqMan PCR, conventional reverse transcriptase PCR (RT-PCR), changes in downstream surrogate
10 markers (*i.e.*, Wnt responsive genes), and anti-Dkk Western blots for protein detection. Other methods would be readily apparent to the artisan of ordinary skill.

12. Methods to Identify Agents that Modulate at Least One Activity of Dkk, a Dkk Interacting Protein, or LRP5/LRP6/HBM

Another embodiment of the present invention provides methods for identifying agents that modulate at least one activity of Dkk, Dkk interacting proteins, and/or LRP5/LRP6/HBM proteins or preferably which specifically modulate an activity of a Dkk/Dkk interacting protein complex or an LRP5(or LRP6/HBM)/Dkk complex, or a biologically active fragment of Dkk (*e.g.*, comprising the domain which binds
15 LRP5/LRP6/HBM) or a Dkk interacting protein complex. Such methods or assays may utilize any means of monitoring or detecting the desired activity as would be known in the art (See, *e.g.*, Wu *et al.*, *Curr. Biol.* 10:1611-4 (2000); Fedi *et al.*, *J. Biol. Chem.* 274:19465-72 (1991); Grotewold *et al.*, *Mech. Dev.* 89:151-3 (1999); Shibata *et al.*, *Mech. Dev.* 96:243-6 (2000); Wang *et al.*, *Oncogene* 19:1843-8 (2000); and Glinka *et al.*, *Nature* 391:357-62 (1998)). Potential agents which modulate Dkk include, for
20 example, p53, the tumor suppressor protein, which can induce Dkk-1. Damage to DNA has also been observed to up-regulate Dkk-1 expression via a stabilization and activation of p53 (Wang *et al.*, *Oncogene* 19:1843-48 (2000)); and, Shou *et al.*, *Oncogene* 21:878-89 (2002)). Additionally, Fedi *et al.* (1999) purportedly showed that
25 Dkk-1 can block the Wnt2-induced oncogenic transformation of NIH-3T3 cells.
30

Furthermore, it has been suggested that Dkk expression can be modulated by BMP signaling in the developing skeleton (Mukhopadhyay *et al.*, *Dev. Cell.* 1:423-34 (2001); and Grotewold *et al.*, *EMBO J.* 21:966-75 (2002)). Grotewold *et al.* additionally describe altered Dkk expression levels in response to stress signals including UV irradiation and other genotoxic stimuli. They propose that Dkk expression is pro-apoptotic. In animals expressing HBM constructs conferring high bone mass, a reduced osteoblast apoptosis effect was observed. Thus, HBM and HBM-like variants may control/alter Dkk's role in programmed cell death. Other agents which potentially modulate Dkk activity include the Dkk interacting proteins identified in Figure 5.

In one embodiment, the relative amounts of Dkk or a Dkk interacting protein of a cell population that has been exposed to the agent to be tested is compared to an unexposed control cell population. Antibodies can be used to monitor the differential expression of the protein in the different cell populations. Cell lines or populations are exposed to the agent to be tested under appropriate conditions and time. Cellular lysates may be prepared from the exposed cell line or population and a control, unexposed cell line or population. The cellular lysates are then analyzed with the probe, as would be known in the art. See, e.g., Ed Harlow and David Lane, Antibodies: A Laboratory Manual (Cold Spring Harbor, NY, 1988) and Ed Harlow and David Lane, Using Antibodies: A Laboratory Manual (Cold Spring Harbor, NY 1998).

For example, N- and C- terminal fragments of Dkk can be expressed in bacteria and used to search for proteins which bind to these fragments. Fusion proteins, such as His-tag or GST fusion to the N- or C-terminal regions of Dkk (or to biologically active domains of Dkk-1) or a whole Dkk protein can be prepared. These fusion proteins can be coupled to, for example, Talon or Glutathione-Sepharose beads and then probed with cell lysates to identify molecules which bind to Dkk. Prior to lysis, the cells may be treated with purified Wnt proteins, RNA, or drugs which may modulate Wnt signaling or proteins that interact with downstream elements of the Wnt pathway. Lysate proteins binding to the fusion proteins can be resolved by SDS-PAGE, isolated and identified by, for example protein sequencing or mass spectroscopy, as is known in the art. See, e.g., Protein Purification Applications: A Practical Approach (Simon Roe, ed., 2nd ed.

Oxford Univ. Press, 2001) and "Guide to Protein Purification" in *Meth. Enzymology* vol. 182 (Academic Press, 1997).

The activity of Dkk, a Dkk interacting protein, or a complex of Dkk with LRP5/LRP6/HBM may be affected by compounds which modulate the interaction
5 between Dkk and a Dkk interacting protein (such as those shown in Figure 5) and/or Dkk and LRP5/LRP6/HBM. The present invention provides methods and research tools for the discovery and characterization of these compounds. The interaction between Dkk and a Dkk interacting protein and/or Dkk and LRP5/6/HBM may be monitored *in vivo* and *in vitro*. Compounds which modulate the stability of a Dkk -
10 LRP5/LRP6/HBM complex are potential therapeutic compounds. Example *in vitro* methods include: Binding LRP5/6/HBM, Dkk, or a Dkk interacting protein to a sensor chip designed for an instrument such are made by Biacore (Uppsala, Sweden) for the performance of an plasmon resonance spectroscopy observation. In this method, the chip with one of Dkk, a Dkk interacting protein, or LRP5/6 is first exposed to the other
15 under conditions which permit them to form the complex. A test compound is then introduced and the output signal of the instrument provides an indication of any effect exerted by the test compound. By this method, compounds may be rapidly screened. Another, *in vitro*, method is exemplified by the SAR-by-NMR methods (Shuker *et al.*, *Science*. 274:1531-4 (1996)). Briefly, a Dkk-1 binding domain and/or LRP 5 or 6 LBD
20 are expressed and purified as ¹⁵N labeled protein by expression in labeled media. The labeled protein(s) are allowed to form the complex in solution in an NMR sample tube. The heteronuclear correlation spectrum in the presence and absence of a test compound provides data at the level of individual residues with regard to interactions with the test compound and changes at the protein-protein interface of the complex.
25 One of skill in the art knows of many other protocols, e.g. affinity capillary electrophoresis (Okun *et al. J Biol Chem* 276:1057-62 (2001); Vergun and Chu, *Methods*, 19:270-7 (1999)), fluorescence spectroscopy, electron paramagnetic resonance, etc. which can monitor the modulation of a complex and/or measure binding affinities for complex formation.

In vitro protocols for monitoring the modulation of a Dkk/LRP5/LRP6/HBM complex include the yeast two hybrid protocol. The yeast two hybrid method may be used to monitor the modulation of a complex in vivo by monitoring the expression of genes activated by the formation of a complex of fusion proteins of Dkk and LRP ligand binding domains. Nucleic acids according to the invention which encode the interacting Dkk and LRP LBD domains are incorporated into bait and prey plasmids. The Y2H protocol is performed in the presence of one or more test compounds. The modulation of the complex is observed by a change in expression of the complex activated gene. It will be appreciated by one skilled in the art that test compounds can be added to the assay directly or, in the case of proteins, can be coexpressed in the yeast with the bait and prey compounds. Similarly, fusion proteins of Dkk and Dkk interacting proteins can also be used in a Y2H screen to identify other proteins which modulate the Dkk/Dkk interacting protein complex.

Assay protocols such as these may be used in methods to screen for compounds, drugs, treatments which modulate the Dkk/Dkk interacting protein and/or Dkk/LRP5/6 complex, whether such modulation occurs by competitive binding, or by altering the structure of either LRP 5/6 or Dkk at the binding site, or by stabilizing or destabilizing the protein-protein interface. It may be anticipated that peptide aptamers may competitively bind, although induction of an altered binding site structure by steric effects is also possible.

12.1 Antibodies and Antibody Fragments

Polyclonal and monoclonal antibodies and fragments of these antibodies which bind to Dkk or LRP5/LRP6/HBM can be prepared as would be known in the art. For example, suitable host animals can be immunized using appropriate immunization protocols and the peptides, polypeptides or proteins of the invention. Peptides for use in immunization are typically about 8-40 residues long. If necessary or desired, the polypeptide immunogens can be conjugated to suitable carriers. Methods for preparing immunogenic conjugates with carriers such as bovine serum albumin (BSA), keyhole limpet hemocyanin (KLH), or other carrier proteins are well known in the art (See,

Harlow *et al.*, 1988). In some circumstances, direct conjugation using, for example, carbodiimide reagents, may be effective; in other instances linking reagents such as those supplied by Pierce Chemical Co., Rockford, IL, may be desirable to provide accessibility to the polypeptide or hapten. The hapten peptides can be extended at
5 either the amino or carboxy terminus with a cysteine residue or interspersed with cysteine residues, for example, to facilitate linking to a carrier. Administration of the immunogens is conducted generally by injection over a suitable time period and with use of suitable adjuvants, as is generally understood in the art. During the immunization schedule, titers of antibodies are taken to determine adequacy of
10 antibody formation.

Anti-peptide antibodies can be generated using synthetic peptides, for example, the peptides derived from the sequence of any Dkk, including Dkk-1, or LRP5/LRP6/HBM. Synthetic peptides can be as small as 2-3 amino acids in length, but are preferably at least 3, 5, 10, or 15 or more amino acid residues long. Such peptides
15 can be determined using programs such as DNASTar. The peptides are coupled to KLH using standard methods and can be immunized into animals such as rabbits. Polyclonal anti-Dkk or anti-LRP5/LRP6/HBM peptide antibodies can then be purified, for example using Actigel beads containing the covalently bound peptide.

While the polyclonal antisera produced in this way may be satisfactory for some
20 applications, for pharmaceutical compositions, use of monoclonal preparations is preferred. Immortalized cell lines which secrete the desired monoclonal antibodies may be prepared using the standard method of Kohler and Milstein or modifications which effect immortalization of lymphocytes or spleen cells, as is generally known (See, e.g., Harlow *et al.*, 1988 and 1998). The immortalized cell lines secreting the desired
25 antibodies can be screened by immunoassay in which the antigen is the peptide hapten, polypeptide or protein. When the appropriate immortalized cell culture secreting the desired antibody is identified, the cells can be cultured either *in vitro* or by production in ascites fluid.

The desired monoclonal antibodies are then recovered from the culture
30 supernatant or from the ascites supernatant. Fragments of the monoclonal antibodies

which contain the immunologically significant portion can be used as agonists or antagonists of Dkk activity. Use of immunologically reactive fragments, such as the Fab, scFV, Fab', of F(ab')₂ fragments are often preferable, especially in a therapeutic context, as these fragments are generally less immunogenic than the whole immunoglobulin.

The antibodies or fragments may also be produced, using current technology, by recombinant means. Regions that bind specifically to the desired regions of Dkk or LRP5/LRP6/HBM can also be produced in the context of chimeras with multiple species origin. Antibody reagents so created are contemplated for use diagnostically or as stimulants or inhibitors of Dkk activity.

In one embodiment, antibodies against Dkk, bind Dkk with high affinity, i.e., ranging from 10⁻⁵ to 10⁻⁹ M. Preferably, the anti-Dkk antibody will comprise a chimeric, primate, Primatized®, human or humanized antibody. Also, the invention embraces the use of antibody fragments, e.g., Fab's, Fv's, Fab's, F(ab)₂, and aggregates thereof.

Another embodiment contemplates chimeric antibodies which recognize Dkk or LRP5/LRP6/HBM. A chimeric antibody is intended to refer to an antibody with non-human variable regions and human constant regions, most typically rodent variable regions and human constant regions.

A "primatized® antibody" refers to an antibody with primate variable regions, e.g., CDR's, and human constant regions. Preferably, such primate variable regions are derived from an Old World monkey.

A "humanized antibody" refers to an antibody with substantially human framework and constant regions, and non-human complementarity-determining regions (CDRs). "Substantially" refers to the fact that humanized antibodies typically retain at least several donor framework residues (i.e., of non-human parent antibody from which CDRs are derived).

Methods for producing chimeric, primate, primatized®, humanized and human antibodies are well known in the art. See, e.g., U.S. Patent 5,530,101, issued to Queen *et al.*; U.S. Patent 5,225,539, issued to Winter *et al.*; U.S. Patents 4,816,397 and

4,816,567, issued to Boss *et al.* and Cabilly *et al.* respectively, all of which are incorporated by reference in their entirety.

The selection of human constant regions may be significant to the therapeutic efficacy of the subject anti-Dkk or LRP5/LRP6/HBM antibody. In a preferred embodiment, the subject anti-Dkk or LRP5/LRP6/HBM antibody will comprise human, gamma 1, or gamma 3 constant regions and, more preferably, human gamma 1 constant regions.

Methods for making human antibodies are also known and include, by way of example, production in SCID mice, and *in vitro* immunization.

The subject anti-Dkk or LRP5/LRP6/HBM antibodies can be administered by various routes of administration, typically parenteral. This is intended to include intravenous, intramuscular, subcutaneous, rectal, vaginal, and administration with intravenous infusion being preferred.

The anti-Dkk or LRP5/LRP6/HBM antibody will be formulated for therapeutic usage by standard methods, e.g., by addition of pharmaceutically acceptable buffers, e.g., sterile saline, sterile buffered water, propylene glycol, and combinations thereof.

Effective dosages will depend on the specific antibody, condition of the patient, age, weight, or any other treatments, among other factors. Typically effective dosages will range from about 0.001 to about 30 mg/kg body weight, more preferably from about 0.01 to 25 mg/kg body weight, and most preferably from about 0.1 to about 20 mg/kg body weight.

Such administration may be effected by various protocols, e.g., weekly, bi-weekly, or monthly, depending on the dosage administered and patient response. Also, it may be desirable to combine such administration with other treatments.

Antibodies to Dkk-1 interacting proteins, such as those identified in Figure 5, are also contemplated according to the present invention, and can be used similarly to the Dkk-1 antibodies mentioned in the above methodology.

The antibodies of the present invention can be utilized in experimental screening, as diagnostic reagents, and in therapeutic compositions.

12.2 Chemical Libraries

Agents that are assayed by these methods can be randomly selected or rationally selected or designed. As used herein, an agent is said to be randomly selected when the agent is chosen randomly without considering the specific
5 sequences involved in the association of Dkk-1 alone, Dkk-1 interacting proteins alone, or with their associated substrates, binding partners, etc. An example of randomly selected agents is the use of a chemical library or a peptide combinatorial library, or a growth broth of an organism.

10 The agents of the present invention can be, as examples, peptides, small molecules, vitamin derivatives, as well as carbohydrates. A skilled artisan can readily recognize that there is no limit as to the structural nature of the agents of the present invention.

12.3 Peptide Synthesis

15 The peptide agents of the invention can be prepared using standard solid phase (or solution phase) peptide synthesis methods, as is known in the art. In addition, the DNA encoding these peptides may be synthesized using commercially available oligonucleotide synthesis instrumentation and produced recombinantly using standard recombinant production systems. The production of polypeptides using solid phase
20 peptide synthesis is necessitated if non-nucleic acid-encoded amino acids are to be included.

13. Uses for Agents that Modulate at Least One Activity of Dkk, a Dkk Interacting Protein, a Dkk/Dkk Interacting Protein Complex, or a Dkk/LRP5 or 25 Dkk/LRP6 Complex

The proteins and nucleic acids of the invention, such as the proteins or polypeptides containing an amino acid sequence of LRP5, Dkk, and Dkk interacting proteins are involved in bone mass modulation and lipid modulation of other Wnt pathway mediated activity. Agents that modulate (*i.e.*, up and down-regulate) the
30 expression of Dkk or Dkk interacting proteins, or agents, such as agonists and

antagonists respectively, of at least one activity of Dkk or a Dkk interacting protein may be used to modulate biological and pathologic processes associated with the function and activity of Dkk or a Dkk interacting protein.

As used herein, a subject can be preferably any mammal, so long as the mammal is in need of modulation of a pathological or biological process modulated by a protein of the invention. The term "mammal" means an individual belonging to the class *Mammalia*. The invention is particularly useful in the treatment of human subjects.

As used herein, a biological or pathological process modulated by Dkk or a Dkk interacting protein may include binding of Dkk to a Dkk interacting protein, Dkk to LRP5 or LRP6 or release therefrom, inhibiting or activating Dkk or a Dkk interacting protein mRNA synthesis or inhibiting Dkk or Dkk interacting protein modulated inhibition of LRP5 or LRP6 mediated Wnt signaling. Further bone-related markers may be observed such as alkaline phosphatase activity, osteocalcin production, or mineralization.

Pathological processes refer to a category of biological processes which produce a deleterious effect. For example, expression or up-regulation of expression of LRP5 or LRP6 and/or Dkk and/or a Dkk interacting protein may be associated with certain diseases or pathological conditions. As used herein, an agent is said to modulate a pathological process when the agent statistically significantly ($p < 0.05$) alters the process from its base level in the subject. For example, the agent may reduce the degree or severity of the process mediated by that protein in the subject to which the agent was administered. For instance, a disease or pathological condition may be prevented, or disease progression modulated by the administration of agents which reduce or modulate in some way the expression or at least one activity of a protein of the invention.

As LRP5/6 and Dkk are involved both directly and indirectly in bone mass modulation, one embodiment of this invention is to use Dkk or Dkk interacting protein expression as a method of diagnosing a bone condition or disease. Certain markers are associated with specific Wnt signaling conditions (e.g., *TCF/LEF* activation).

Diagnostic tests for bone conditions may include the steps of testing a sample or an

extract thereof for the presence of Dkk or Dkk interacting protein nucleic acids (*i.e.*, DNA or RNA), oligomers or fragments thereof or protein products of TCF/LEF regulated expression. For example, standard *in situ* hybridization or other imaging techniques can be utilized to observe products of Wnt signaling.

5 This invention also relates to methods of modulating bone development or bone loss conditions. Inhibition of bone loss may be achieved by inhibiting or modulating changes in the LRP5/6 mediated Wnt signaling pathway. For example, absence of LRP5 activity may be associated with low bone mass. Increased activity LRP5 may be associated with high bone mass. Therefore, modulation of LRP5 activity will in turn
10 modulate bone development. Modulation of the Dkk/LRP5/6 or Dkk/Dkk interacting protein complex via agonists and antagonists is one embodiment of a method to regulate bone development. Such modulation of bone development can result from inhibition of the activity of, for example, a Dkk/LRP(5/6) protein complex, a Dkk/Dkk interacting protein complex, upregulated transcription of the *LRP5* gene or inhibited
15 translation of Dkk or Dkk interacting protein mRNA.

 The agents of the present invention can be provided alone, or in combination with other agents that modulate a particular pathological process. As used herein, two agents are said to be administered in combination when the two agents are administered simultaneously or are administered independently in a fashion such that
20 the agents will act at the same time.

 The agents of the present invention can be administered via parenteral, subcutaneous (sc), intravenous (iv), intramuscular (im), intraperitoneal (ip), transdermal or buccal routes. Alternatively, or concurrently, administration may be by the oral route. The dosage administered will be dependent upon the age, health, and weight of the
25 recipient, kind of concurrent treatment, if any, frequency of treatment, and the nature of the effect desired.

 The present invention further provides compositions containing one or more agents which modulate expression or at least one activity of a protein of the invention. While individual needs vary, determination of optimal ranges of effective amounts of
30 each component is within the skill of the art. Typical dosages of the active agent which

mediate Dkk or Dkk interacting protein activity comprise from about 0.0001 to about 50 mg/kg body weight. The preferred dosages comprise from about 0.001 to about 50 mg/kg body weight. The most preferred dosages comprise from about 0.1 to about 1 mg/kg body weight. In an average human of 70 kg, the range would be from about 7
5 µg to about 3.5 g, with a preferred range of about 0.5 mg to about 5 mg.

In addition to the pharmacologically active agent, the compositions of the present invention may contain suitable pharmaceutically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically for delivery to the site of action.

10 Suitable formulations for parenteral administration include aqueous solutions of the active compounds in water-soluble form, for example, water-soluble salts. In addition, suspensions of the active compounds as appropriate oily injection suspensions may be administered. Suitable lipophilic solvents or vehicles include fatty oils, for example, sesame oil, or synthetic fatty acid esters, (e.g., ethyl oleate or triglycerides). Aqueous
15 injection suspensions may contain substances which increase the viscosity of the suspension include, for example, sodium carboxymethyl cellulose, sorbitol and/or dextran. Optionally, the suspension may also contain stabilizers. Liposomes and other non-viral vectors can also be used to encapsulate the agent for delivery into the cell.

The pharmaceutical formulation for systemic administration according to the
20 invention may be formulated for enteral, parenteral, or topical (top) administration. Indeed, all three types of formulations may be used simultaneously to achieve systemic administration of the active ingredient.

Suitable formulations for oral administration include hard or soft gelatin capsules, pills, tablets, including coated tablets, elixirs, suspensions, syrups or inhalations and
25 controlled release forms thereof.

Potentially, any compound which binds Dkk or a Dkk interacting protein or modulates the Dkk/LRP5 or Dkk/LRP6 or Dkk/Dkk interacting protein complex may be a therapeutic compound. In one embodiment of the invention, a peptide or nucleic acid aptamer according to the invention is used in a therapeutic composition. Such
30 compositions may comprise an aptamer, or a LRP5 or LRP6 fragment unmodified or

modified. In another embodiment, the therapeutic compound comprises a Dkk-1 interacting protein, or biologically active fragment thereof.

Nucleic acid aptamers have been used in compositions for example by chemical bonding to a carrier molecule such as polyethylene glycol (PEG) which may facilitate uptake or stabilize the aptamer. A di-alkylglycerol moiety attached to an RNA will embed the aptamer in liposomes, thus stabilizing the compound. Incorporating chemical substitutions (i.e. changing the 2'OH group of ribose to a 2'NH in RNA confers ribonuclease resistance) and capping, etc. can prevent breakdown. Several such techniques are discussed for RNA aptamers in Brody and Gold (*Rev. Mol. Biol.* 74:3-13 (2000)).

Peptide aptamers may be used in therapeutic applications by the introduction of an expression vector directing aptamer expression into the affected tissue such as for example by retroviral delivery, by encapsulating the DNA in a delivery complex or simple by naked DNA injection. Or, the aptamer itself or a synthetic analog may be used directly as a drug. Encapsulation in polymers and lipids may assist in delivery. The use of peptide aptamers as therapeutic and diagnostic agents is reviewed by Hoppe-Syler and Butz (*J. Mol. Med.* 78:426-430 (2000)).

In another aspect of the invention. The structure of a constrained peptide aptamer of the invention may be determined such as by NMR or X-ray crystallography. (Cavanagh et al., Protein NMR Spectroscopy: Principles and Practice, Academic Press, 1996; Drenth, Principles of Protein X-Ray Crystallography, Springer Verlag, 1999) Preferably the structure is determined in complex with the target protein. A small molecule analog is then designed according to the positions of functional elements of the 3D structure of the aptamer. (Guidebook on Molecular Modeling in Drug Design, Cohen, Ed., Academic Press, 1996; Molecular Modeling and Drug Design (Topics in Molecular and Structural Biology), Vinter and Gardner Eds., CRC Press, 1994) Thus the present invention provides a method for the design of effective and specific drugs which modulate the activity of Dkk, Dkk interacting proteins, Dkk/Dkk interacting protein complex and the Dkk/LRP complex. Small molecule mimetics of the peptide aptamers of the present invention are encompassed within the scope of the invention.

In practicing the methods of this invention, the compounds of this invention may be used alone or in combination, or in combination with other therapeutic or diagnostic agents. In certain preferred embodiments, the compounds of this invention may be co-administered along with other compounds typically prescribed for these conditions according to generally accepted medical practice. For example, the compounds of this invention can be administered in combination with other therapeutic agents for the treatment of bone loss. Bone loss mediating agents include bone resorption inhibitors such as bisphosphonates (e.g., alendronic acid, clodronic acid, etidronic acid, pamidronic acid, risedronic acid and tiludronic acid), vitamin D and vitamin D analogs, cathepsin K inhibitors, hormonal agents (e.g., calcitonin and estrogen), and selective estrogen receptor modulators or SERMs (e.g., raloxifene). And bone forming agents such as parathyroid hormone (PTH) and bone morphogenetic proteins (BMP).

Additionally contemplated are combinations of agents which regulate Dkk-1 and agents which regulate lipid levels such as HMG-CoA reductase inhibitors (i.e., statins such as Mevacor®, Lipitor® and other inhibitors such as Baycol®, Lescol®, Pravachol® and Zocor®), bile acid sequestrants (e.g., Colestid® and Welchol®), fibric acid derivatives (Atromid-S®, Lopid®, Tricor®), and nicotinic acid.

[0001] The compounds of this invention can be utilized *in vivo*, ordinarily in vertebrates and preferably in mammals, such as humans, sheep, horses, cattle, pigs, dogs, cats, rats and mice, or *in vitro*.

14. Transgenic Animals

Transgenic animal models can be created which conditionally express Dkk and/or LRP5 or LRP6 and/or Dkk interacting proteins, such as those shown in Figure 5. These animals can be used as research tools for the study of the physiological effects of the Dkk-1/Dkk-1 interacting protein interaction and/or the LRP5 / Dkk interaction. Alternatively, transgenic animals can be created which express a transgenic form of Dkk alone or in addition to a transgenic form of HBM or express Dkk interacting proteins alone or in addition to a transgenic form of Dkk. Transgenic animals expressing HBM or LRP5 can be crossed with transgenic animals expressing Dkk or

Dkk interacting proteins to obtain heterozygote as well as homozygote animals which express both desired genes.

Animal models may be created to directly modulate the Dkk/Dkk interacting protein or Dkk/ LRP5 interaction activity *in vivo* to serve as a research tool for determining the efficacy of candidate compounds which modulate the Dkk/Dkk interacting protein or LRP5 / Dkk interaction activity *in vitro*. Animals, such as transgenic mice, can be created using the techniques employed to make transgenic mice that express for example, human Dkk or a Dkk interacting protein, or knockouts (KO), which may be conditional, of the gene encoding mouse Dkk or Dkk interacting protein. Knock-in animals include animals wherein genes have been introduced and animals wherein a gene that was previously knocked-out is reintroduced into the animal. Other transgenic animals can be created with inducible forms of Dkk or a Dkk interacting protein to study the effects of the gene on bone mass development and loss as well as lipid level regulation. These animals can also be used to study long term effects of Dkk or Dkk interacting protein modulation. Transgenic animals may be created to express peptide aptamers, or produce RNA aptamers. The transgenic vectors may direct expression in a tissue specific manner by the use of tissue specific promoters. In a preferred embodiment, a peptide aptamer fusion protein is expressed using a bone specific promoter. Such systems can provide a tissue specific knock-out of Dkk or Dkk interacting protein activity.

General methods for creating transgenic animals are known in the art, and are described in, for example, Strategies in Transgenic Animal Science (Glenn M. Monastersky and James M. Robl eds., ASM Press; Washington, DC, 1995); Transgenic Animal Technology: A Laboratory Handbook (Carl A. Pinkert ed., Academic Press 1994); Transgenic Animals (Louis Marie Houdebine, ed., Harwood Academic Press, 1997); Overexpression and Knockout of Cytokines in Transgenic Mice (Chaim O. Jacob, ed., Academic Press 1994); Microinjection and Transgenesis: Strategies and Protocols (Springer Lab Manual) (Angel Cid-Arregui and Alejandro Garcia-Carranca, eds., Springer Verlag 1998); and Manipulating the Mouse Embryo: A Laboratory Manual (Brigid Hogan *et al.*, eds., Cold Spring Harbor Laboratory Press 1994).

15. Peptide and Nucleotide Aptamers and Peptide Aptamer Mimetics

Another embodiment contemplates the use of peptide and nucleotide aptamer technology to screen for agents which interact with Dkk, which block Dkk from interacting with LRP5 or LRP6, or which block any other Dkk ligand interaction, or which interact with Dkk interacting proteins, such as those shown in Figure 5. Peptide aptamers are molecules in which a variable peptide domain is displayed from a scaffold protein. Thioredoxin A (trxA) is commonly used for a scaffold. The peptide insert destroys the catalytic site of trxA. It is recognized that numerous proteins may also be used as scaffolding proteins to constrain and/or present a peptide aptamer. Other scaffold proteins that could display a constrained peptide aptamer could include staphylococcal nuclease, the protease inhibitor eglin C, the *Streptomyces tendea* alpha-amylase inhibitor Tendamistat, Sp1, and green fluorescent protein (GFP) (reviewed in Hoppe-Seyler *et al.*, *J. Steroid Biochem Mol. Biol.* 78:105-11 (2001)), and the S1 nuclease from *Staphylococcus* or M13 for phage display. Any molecule to which the aptamer could be anchored and presented in its bioactive conformation would be suitable.

Aptamers can then specifically bind to a given target protein *in vitro* and *in vivo* and have the potential to selectively block the function of their target protein. Peptide aptamers are selected from randomized expression libraries on the basis of their *in vivo* binding capacity to the desired target protein. Briefly, a target protein (e.g., Dkk, a Dkk interacting protein, or LRP5/6) is linked to a heterologous DNA binding domain (BD) and expressed as bait in a yeast test strain. Concomitantly, a library coding for different peptides (e.g., 16-mers) of randomized sequence inserted in a scaffold protein sequence, which are linked to a heterologous transcriptional activation domain (AD) is expressed as prey. If a peptide binds to a target protein, a functional transcription factor is reconstituted, in which the BD and AD are bridged together by interacting proteins. This transcription factor is then able to activate the promoter of a marker gene which can be monitored by colorimetric enzymatic assays or by growth selection. Additional variation, methods of preparing and screening methodologies are described in, for example, Hoppe-Seyler *et al.*, *J. Mol. Med.* 78: 426-430 (2000).

Nucleotide aptamers are described for example in Brody *et al.*, *Trends Mol. Biotechnol.* 74: 5-13 (2000). Additional methods of making and using nucleotide aptamers include SELEX, *i.e.*, Systematic Evolution of Ligands by Exponential Enrichment. SELEX is a process of isolating oligonucleotide ligands of a chosen target molecule (see Tuerk and Gold, *Science* 249:505-510 (1990); U.S. Pat. Nos. 5,475,096, 5,595,877, and 5,660,985). SELEX, as described in Tuerk and Gold, involves admixing the target molecule with a pool of oligonucleotides (*e.g.*, RNA) of diverse sequences; retaining complexes formed between the target and oligonucleotides; recovering the oligonucleotides bound to the target; reverse-transcribing the RNA into DNA; amplifying the DNA with polymerase chain reactions (PCR); transcribing the amplified DNA into RNA; and repeating the cycle with ever increasing binding stringency. Three enzymatic reactions are required for each cycle. It usually takes 12-15 cycles to isolate aptamers of high affinity and specificity to the target. An aptamer is an oligonucleotide that is capable of binding to an intended target substance but not other molecules under the same conditions.

In another reference, Bock *et al.*, *Nature* 355:564-566 (1990), describe a different process from the SELEX method of Tuerk and Gold in that only one enzymatic reaction is required for each cycle (*i.e.*, PCR) because the nucleic acid library in Bock's method is comprised of DNA instead of RNA. The identification and isolation of aptamers of high specificity and affinity with the method of Bock *et al.* still requires repeated cycles in a chromatographic column.

Other nucleotide aptamer methods include those described by Conrad *et al.*, *Meth. Enzymol.* 267:336-367 (1996). Conrad *et al.* describe a variety of methods for isolating aptamers, all of which employ repeated cycles to enrich target-bound ligands and require a large amount of purified target molecules. More recently described methods of making and using nucleotide aptamers include, but are not limited to those described in U.S. Patent Nos. 6,180,348; 6,051,388; 5,840,867; 5,780,610, 5,756,291 and 5,582,981.

Potentially, any compound which binds Dkk or a Dkk interacting protein or modulates the Dkk/Dkk interacting protein or Dkk/LRP5 or Dkk/LRP6 complex may be

a therapeutic compound. In one embodiment of the invention, a peptide or nucleic acid aptamer according to the invention is used in a therapeutic composition. Such compositions may comprise an aptamer, or a LRP5 or LRP6 fragment unmodified or modified.

5 Nucleic acid aptamers have been used in compositions for example by chemical bonding to a carrier molecule such as polyethylene glycol (PEG) which may facilitate uptake or stabilize the aptamer. A di-alkylglycerol moiety attached to an RNA will embed the aptamer in liposomes, thus stabilizing the compound. Incorporating chemical substitutions (*i.e.*, changing the 2'-OH group of ribose to a 2'-NH in RNA
10 confers ribonuclease resistance) and capping, etc. can prevent breakdown. Several such techniques are discussed for RNA aptamers in Brody and Gold *Rev. Mol. Biol.* 74:3-13 (2000).

Peptide aptamers may be used in therapeutic applications by the introduction of an expression vector directing aptamer expression into the affected tissue such as for
15 example by retroviral delivery, by encapsulating the DNA in a delivery complex or simply by naked DNA injection. Or, the aptamer itself or a synthetic analog may be used directly as a drug. Encapsulation in polymers and lipids may assist in delivery. The use of peptide aptamers as therapeutic and diagnostic agents is reviewed by Hoppe-Syler and Butz *J. Mol. Med.* 78:426-430 (2000).

20 In another aspect of the invention, the structure of a constrained peptide aptamer of the invention may be determined such as by NMR or X-ray crystallography. (Cavanagh et al., Protein NMR Spectroscopy : Principles and Practice, Academic Press, 1996; Drenth, Principles of Protein X-Ray Crystallography, Springer Verlag, 1999) Preferably the structure is determined in complex with the target protein. A
25 small molecule analog is then designed according to the positions of functional elements of the 3D structure of the aptamer. (Guidebook on Molecular Modeling in Drug Design, Cohen, Ed., Academic Press, 1996; Molecular Modeling and Drug Design (Topics in Molecular and Structural Biology), Vinter and Gardner Eds., CRC Press, 1994) Thus, a method is provided for the design of effective and specific drugs which
30 modulate the activity of Dkk, Dkk interacting proteins, Dkk/Dkk interacting protein

complex, and the Dkk/LRP complex. Small molecule mimics of the peptide aptamers of the present invention are also encompassed within the scope of the invention.

16. Alternative Variants of LRP5/LRP6 Having HBM Activity

5 A structural model of the LRP5/Zmax1 first beta-propeller module was generated based on a model prediction in Springer et al., (1998) *J. Molecular Biology*, 283:837-862. Based on the model, certain amino acid residues were identified as important variants of LRP5/HBM/Zmax1. The following three categories provide examples of such variants:

10 The shape of the beta-propeller resembles a disk with inward-sloping sides and a hole down the middle. Residue 171 is in a loop on the outer or top surface of the domain in blade 4 of propeller module 1. Thus, variants comprising changed residues in structurally equivalent positions in other blades; as well as residues that are slightly more interior to the binding pocket, but still accessible to the surface, are important
15 embodiments of the present invention for the study of bone mass modulation by LRP5/HBM, for the development of pharmaceuticals and treatments of bone mass disorders, and for other objectives of the present invention. The following are examples of such variants:

20 A214V (a position equivalent to 171 in blade 5; alanine is not conserved in other propellers),

E128V (a position equivalent to 171 in blade 3; glutamate is not conserved in other propellers),

A65V (a position equivalent to 171 in blade 2; alanine is conserved in propellers 1-3 but not 4),

25 G199V (an accessible interior position in blade 5; glycine is conserved in propellers 1-3 but not 4), and

M282V (accessible interior position in blade 1; methionine is conserved in propellers 1-3 but not 4).

30 LRP5/Zmax1 has four beta-propeller structures; the first three beta-propeller modules conserve a glycine in the position corresponding to residue 171 in human

LRP5/Zmax1. Therefore, variants bearing a valine in the equivalent positions in the other propellers are important embodiments of the present invention. The following variants are useful for the study of bone mass modulation by LRP5/HBM, for the development of pharmaceuticals and treatments of bone mass disorders, and for other objectives of the present invention: G479V, G781V, and Q1087V.

The G171V HBM polymorphism results in "occupied space" of the beta-propeller 1, with the side-chain from the valine residue sticking out into an open binding pocket and potentially altering a ligand/protein interaction. The glycine residue is conserved in LRP5/Zmax1 propellers 1, 2 and 3 but is a glutamine in propeller 4. Therefore, the following variants of LRP5/HBM are important embodiments of the present invention for the study of bone mass modulation by LRP5/HBM, for the development of pharmaceuticals and treatments of bone mass disorders, and for other objectives of the present invention:

G171K (which introduces a charged side-chain),
G171F (which introduces a ringed side-chain),
G171I (which introduces a branched side-chain), and
G171Q (which introduces the propeller 4 residue).

Furthermore, LRP6 is the closest homolog of LRP5/Zmax1. LRP6 has a beta-propeller structure predicted to be similar, if not identical to Zmax1. The position corresponding to glycine 171 in human LRP5/Zmax1 is glycine 158 of human LRP6. Thus, corresponding variants of LRP6 are an important embodiment of the present invention for the study of the specificity of LRP5/Zmax1 versus its related family member, for the development of pharmaceuticals and treatments of bone mass disorders, and for other objectives of the present invention. Specifically, for example, a glycine to valine substitution at the structurally equivalent position, residue 158, of human LRP6 and similar variants of other species' LRP6 homologs represent important research tools.

Site-directed mutants of LRP5 were generated in the full-length human LRP5 cDNA using the QuikChange XL-Site-Directed Mutagenesis Kit (catalog #200516,

Stratagene, La Jolla, CA) following the manufacturer's protocol. The mutant sequences were introduced using complementary synthetic oligonucleotides:

A65V: TGGTCAGCGGCCTGGAGGATGTGGCCGCAGTGGACTTCC (SEQ ID NO:129) and

5 GGAAGTCCACTGCGGCCACATCCTCCAGGCCGCTGACCA (SEQ ID NO:130)

E128V: AAGCTGTACTGGACGGACTCAGTGACCAACCGCATCGAGG (SEQ ID NO:131) and

10 CCTCGATGCGGTTGGTCACTGAGTCCGTCCAGTACAGCTT (SEQ ID NO:132)

G171K: ATGTACTGGACAGACTGGAAGGAGACGCCCCGGATTGAGCG (SEQ ID NO: 133) and

CGCTCAATCCGGGGCGTCTCCTTCCAGTCTGTCCAGTACAT (SEQ ID NO:134)

15 G171F: ATGTACTGGACAGACTGGTTTGAGACGCCCCGGATTGAGCG (SEQ ID NO:135) and

CGCTCAATCCGGGGCGTCTCAAACCAGTCTGTCCAGTACAT (SEQ ID NO:136)

20 G171I: ATGTACTGGACAGACTGGATTGAGACGCCCCGGATTGAGCG (SEQ ID NO:137) and

CGCTCAATCCGGGGCGTCTCAATCCAGTCTGTCCAGTACAT (SEQ ID NO:138)

G171Q: ATGTACTGGACAGACTGGCAGGAGACGCCCCGGATTGAGCG (SEQ ID NO:139) and

25 CGCTCAATCCGGGGCGTCTCCTGCCAGTCTGTCCAGTACAT (SEQ ID NO:140)

G199V: CGGACATTTACTGGCCCAATGTACTGACCATCGACCTGGAGG (SEQ ID NO:141) and

30 CCTCCAGGTCGATGGTCAGTACATTGGGCCAGTAAATGTCCG (SEQ ID NO:142)

A214V: AGCTCTACTGGGCTGACGTCAAGCTCAGCTTCATCCACCG (SEQ ID NO: 143) and

CGGTGGATGAAGCTGAGCTTGACGTCAGCCCAGTAGAGCT (SEQ ID NO:144)

5 M282V: GAGTGCCCTCTACTCACCCGTGGACATCCAGGTGCTGAGCC (SEQ ID NO:145) and

GGCTCAGCACCTGGATGTCCACGGGTGAGTAGAGGGCACTC (SEQ ID NO:146)

10 G479V: CATGTACTGGACAGACTGGGTAGAGAACCCTAAAATCGAGTGTGC (SEQ ID NO:147) and

GCACACTCGATTTTAGGGTTCTCTACCCAGTCTGTCCAGTACATG (SEQ ID NO:148)

G781V: CATCTACTGGACCGAGTGGGTCTGGCAAGCCGAGGATCGTGCG (SEQ ID NO:149) and

15 CGCACGATCCTCGGCTTGCCGACCCACTCGGTCCAGTAGATG (SEQ ID NO:150)

Q1087V: GTACTTCACCAACATGGTGGACCGGGCAGCCAAGATCGAACG (SEQ ID NO:151) and

20 CGTTCGATCTTGGCTGCCCCGGTCCACCATGTTGGTGAAGTAC (SEQ ID NO:152)

LRP6 G158V:

GTACTGGACAGACTGGGTAGAAGTGCCAAAGATAGAACGTGC (SEQ ID NO:153) and

25 GCACGTTCTATCTTTGGCACTTCTACCCAGTCTGTCCAGTAC (SEQ ID NO:154).

All constructs were sequence verified to ensure that only the engineered modification was present in the gene. Once verified, each variant was functionally evaluated in the TCF-luciferase assay in U2OS cells (essentially as described in Example 7. Other functional evaluations could also be performed, such as the *Xenopus* embryo assay (essentially as described in Example 6), or other assays to evaluate Wnt

signaling, Dkk modulation, or anabolic bone effect. Binding of these mutants to Dkk, LRP-interacting proteins, Dkk-interacting proteins, or peptide aptamers to any of the preceding could also be investigated in a variety of ways such as in a two-hybrid system (such as in yeast as described in this application), or other methods.

5 Figure 24 shows the effects of the G171F mutation in propeller 1 of LRP5. This mutation is at the same position as HBM's G171V substitution. Expression of G171F results in an HBM effect. That is, in the presence of Wnt, G171F is able to activate the TCF-luciferase reporter construct. In fact, it may activate the reporter to a greater extent than either LRP5 or HBM. Furthermore, in the presence of Dkk1 and Wnt1,
10 G171F is less susceptible than LRP5 to modulation by Dkk. These data exemplify that the G171F variant modulates Wnt signaling in a manner similar to HBM. In addition, this data confirms that HBM's valine residue at 171 is not the only modification at 171 that can result in an HBM effect. Together these data support an important role for LRP5 propeller 1 in modulating Wnt pathway activity; in responding to Dkk modulation;
15 and, in the ability to generate an HBM effect.

 Figure 25 shows the effects of the M282V mutation in propeller 1 of LRP5. M282 expression results in an HBM-effect. That is, in the presence of Wnt, M282 is able to activate the TCF-luciferase reporter construct. Furthermore, in the presence of Dkk1 and Wnt1, M282V is less susceptible than LRP5 to modulation by Dkk. These
20 data show that the M282V variant modulates Wnt signaling in a manner similar to HBM. In addition, this data confirms that modifications of other residues in propeller 1 of LRP5 can result in an HBM effect.

 These data support an "occupied space" model of the HBM mutation in propeller 1 and show that multiple mutations of propeller 1 are capable of generating an HBM
25 effect; the original G171V HBM mutation is not unique in this ability. Moreover, various perturbations in propeller 1 can modulate Dkk activity.

 These data illustrate the molecular mechanism of Dkk modulation of LRP signaling. Using the methods disclosed herein and in U.S. Application 60/290,071, generation of a comprehensive mutant panel will reveal residues in LRP that function in
30 Dkk modulation of Wnt signaling. Such variants of LRP5 and LRP6 that modulate Dkk

activity and the residues which distinguish them from LRP5 and LRP6 are points for therapeutic intervention by small molecule compound, antibody, peptide aptamer, or other agents. Furthermore, models of each HBM-effect mutation/polymorphism may be used in rational drug design of an HBM mimetic agent.

5 These are only a few illustrative examples presented to better describe the present invention. Variants of LRP5 which have demonstrated HBM activity in assays include G171F, M282V, G171K, G171Q and A214V. Clearly, other variants may be contemplated within the scope of the present invention. Furthermore, wherever HBM is recited in the methods of the invention, it should be understood that any such
10 alternative variant of LRP which demonstrates HBM biological activity is also encompassed by those claims.

17. Screening Assays

 The two-hybrid system is extremely useful for studying protein:protein
15 interactions. See, e.g., Chien *et al.*, *Proc. Natl Acad. Sci. USA* 88:9578-82 (1991); Fields *et al.*, *Trends Genetics* 10:286-92 (1994); Harper *et al.*, *Cell* 75:805-16 (1993); Vojtek *et al.*, *Cell* 74:205-14 (1993); Luban *et al.*, *Cell* 73:1067-78 (1993); Li *et al.*, *FASEB J.* 7:957-63 (1993); Zang *et al.*, *Nature* 364:308-13 (1993); Golemis *et al.*, *Mol. Cell. Biol.* 12:3006-14 (1992); Sato *et al.*, *Proc. Natl Acad. Sci. USA* 91:9238-42 (1994);
20 Coghlan *et al.*, *Science* 267:108-111 (1995); Kalpana *et al.*, *Science* 266:2002-6 (1994); Helps *et al.*, *FEBS Lett.* 340:93-8 (1994); Yeung *et al.*, *Genes & Devel.* 8:2087-9 (1994); Durfee *et al.*, *Genes & Devel.* 7:555-569 (1993); Paetkau *et al.*, *Genes & Devel.* 8:2035-45; Spaargaren *et al.*, 1994 *Proc. Natl. Acad. Sci. USA* 91:12609-13 (1994); Ye *et al.*, *Proc. Natl Acad. Sci. USA* 91:12629-33 (1994); and U.S. Patent Nos.
25 5,989,808; 6,251,602; and 6,284,519.

 Variations of the system are available for screening yeast phagemid (see, e.g., Harper, Cellular Interactions and Development: A Practical Approach, 153-179 (1993); Elledge *et al.*, *Proc. Natl Acad. Sci. USA* 88:1731-5 (1991)) or plasmid (Bartel, 1993 and Bartel, *Cell* 14:920-4 (1993)); Finley *et al.*, *Proc. Natl Acad. Sci. USA* 91:12980-4

(1994)) cDNA libraries to clone interacting proteins, as well as for studying known protein pairs.

The success of the two-hybrid system relies upon the fact that the DNA binding and polymerase activation domains of many transcription factors, such as GAL4, can be separated and then rejoined to restore functionality (Morin *et al.*, *Nuc. Acids Res.* 21:2157-63 (1993)). While these examples describe two-hybrid screens in the yeast system, it is understood that a two-hybrid screen may be conducted in other systems such as mammalian cell lines. The invention is therefore not limited to the use of a yeast two-hybrid system, but encompasses such alternative systems.

Yeast strains with integrated copies of various reporter gene cassettes, such as for example GAL.fwdarw.LacZ, GAL.fwdarw.HIS3 or GAL.fwdarw.URA3 (Bartel, in Cellular Interactions and Development: A Practical Approach, 153-179 (1993); Harper *et al.*, *Cell* 75:805-16 (1993); Fields *et al.*, *Trends Genetics* 10:286-92 (1994)) are co-transformed with two plasmids, each expressing a different fusion protein. One plasmid encodes a fusion between protein "X" and the DNA binding domain of, for example, the GAL4 yeast transcription activator (Brent *et al.*, *Cell* 43:729-36 (1985); Ma *et al.*, *Cell* 48:847-53 (1987); Keegan *et al.*, *Science* 231:699-704 (1986)), while the other plasmid encodes a fusion between protein "Y" and the RNA polymerase activation domain of GAL4 (Keegan *et al.*, 1986). The plasmids are transformed into a strain of the yeast that contains a reporter gene, such as lacZ, whose regulatory region contains GAL4 binding sites. If proteins X and Y interact, they reconstitute a functional GAL4 transcription activator protein by bringing the two GAL4 components into sufficient proximity to activate transcription. It is well understood that the role of bait and prey proteins may be alternatively switched and thus the embodiments of this invention contemplate and encompass both alternative arrangements.

Either hybrid protein alone must be unable to activate transcription of the reporter gene, the DNA-binding domain hybrid, because it does not provide an activation function, and the activation domain hybrid, because it cannot localize to the GAL4 binding sites. Interaction of the two test proteins reconstitutes the function of GAL4 and results in expression of the reporter gene. The reporter gene cassettes

consist of minimal promoters that contain the GAL4 DNA recognition site (Johnson *et al.*, *Mol. Cell. Biol.* 4:1440-8 (1984); Lorch *et al.*, *J. Mol. Biol.* 186:821-824 (1984)) cloned 5' to their TATA box. Transcription activation is scored by measuring either the expression of β -galactosidase or the growth of the transformants on minimal medium lacking the specific nutrient that permits auxotrophic selection for the transcription product, e.g., URA3 (uracil selection) or HIS3 (histidine selection). See, e.g., Bartel, 1993; Durfee *et al.*, *Genes & Devel.* 7:555-569 (1993); Fields *et al.*, *Trends Genet.* 10:286-292 (1994); and U.S. Pat. No. 5,283,173.

Generally, these methods include two proteins to be tested for interaction which are expressed as hybrids in the nucleus of a yeast cell. One of the proteins is fused to the DNA-binding domain (DBD) of a transcription factor and the other is fused to a transcription activation domain (AD). If the proteins interact, they reconstitute a functional transcription factor that activates one or more reporter genes that contain binding sites for the DBD. Exemplary two-hybrid assays which have been used for Dkk-1 or Dkk-1/LRP5 are presented in the Examples below.

Additional methods of preparing two hybrid assay systems for Dkk-1 interactors would be evident to one of ordinary skill in the art. See for example, Finley *et al.*, "Two-Hybrid Analysis of Genetic Regulatory Networks," in The Yeast Two-Hybrid System (Paul L. Bartel *et al.*, eds., Oxford, 1997); Meijia Yang, "Use of a Combinatorial Peptide Library in the Two-Hybrid Assay," in The Yeast Two-Hybrid System (Paul L. Bartel *et al.*, eds., Oxford, 1997); Gietz *et al.*, "Identification of proteins that interact with a protein of interest: Applications of the yeast two-hybrid system," *Mol. & Cell. Biochem.* 172:67-9 (1997); K. H. Young, "Yeast Two-Hybrid: So Many Interactions,(in) so Little Time," *Biol. Reprod.* 58:302-311 (1998); R. Brent *et al.*, "Understanding Gene and Allele Function with Two-Hybrid Methods," *Annu. Rev. Genet.* 31:663-704 (1997). It will be appreciated that protein networks can be elucidated by performing sequential screens of activation domain-fusion libraries.

Without further description, it is believed that one of ordinary skill in the art can, using the preceding description and the following illustrative examples, make and utilize the compounds of the present invention and practice the claimed methods. The

following working examples therefore, specifically point out preferred embodiments of the present invention, and are not to be construed as limiting in any way the remainder of the disclosure.

5

EXAMPLES

The present invention is described by reference to the following Examples, which are offered by way of illustration and are not intended to limit the invention in any manner. Standard techniques well-known in the art or the techniques specifically described below were utilized.

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For routine practice of the protocols referenced below, one of skill in the art is directed to the references cited in this application as well as the several Current Protocol guides, which are continuously updated, widely available and published by John Wiley and Sons, (New York). In the life sciences, Current Protocols publishes comprehensive manuals in Molecular Biology, Immunology, Human Genetics, Protein Science, Cytometry, Neuroscience, Pharmacology, Cell Biology, Toxicology, and Nucleic Acid Chemistry. Additional sources are known to one of skill in the art.

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Example 1

Yeast Two Hybrid Screen Using LRP5 Ligand Binding Domain (LBD) Bait Sequences

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In a screen against human osteoblast library (*i.e.*, HOB03C5, a custom Gibco generated Y2H compatible cDNA library from a human osteoblast cell line as described by Bodine and Komm, *Bone* 25:535-43 (1999)), an interaction with Dkk-1 was identified. The LRP5 ligand binding domain (LBD) baits used for this screen are depicted in Figures 2B and C. The basic protocol is as follows:

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An overnight culture of the yeast strain containing the bait of interest is grown in 20 ml of appropriate selective medium containing 2% glucose at 30°C. The overnight culture is diluted by a 10 fold factor into YPDmedia supplemented with 40 mg/l of adenine, and grown for 4 hours at 30°C.

For each mating event, an aliquot of the frozen prey library is grown in 150 ml YAPD medium for 5 hours at 30°C.

30

Appropriate volumes calculated by measuring the OD600 of each culture are combined into a tube. The number of diploids to be screened is typically ten times the number of clones originally present in the prey library of interest. Assuming a mating efficiency of 20% minimum, fifty times (*i.e.*, ten times coverage multiplied by 20% mating efficiency) as many haploid cells containing the bait and as many cells containing the prey are used in any given mating event. The mixture is filtered over a 47 mm, 0.45 mm sterile Metrical filter membrane (Gelman).

Using sterile forceps, the filter is transferred onto a 100 mm² YAPD agar plate with the cell side up, removing all air bubbles underneath the filter. The plate is incubated overnight at room temperature.

The filter is transferred into a 50 ml Falcon tube using sterile forceps and 10 ml SD medium containing 2% glucose are added to resuspend the cells. The filter, once free of cells, is removed and the cell suspension is spun for 5 min. at 2,000 xg.

The cells are resuspended in 10 ml SD medium containing 2% glucose. An aliquot of 100 μ l is set aside for titration.

The cells are plated onto large square plates containing appropriate selective media and incubated at 30°C for three to five days.

To calculate the mating efficiency and to determine the total number of diploid cells screened, the 100 μ l aliquot set aside for titration is diluted and plated onto different selective media. The mating efficiency is calculated by dividing the number of diploids/ml by the lowest number of haploids/ml, either bait or prey, and multiplied by 100. For example, if 2 million diploids were obtained by mating 10 million of haploids containing a bait and 12 million of haploids containing a prey, then the mating efficiency is calculated by dividing 2 million by 10 million, which equals 0.2 and multiplied by 100 which equals 20%. Typical mating efficiencies under the above conditions are within about 20 to about 40%. The total number of diploids screened in a mating event is obtained by multiplying the number of diploids/ml by the total number of ml plated, typically about 10.

Isolation of colonies containing pairs of interacting proteins.

Yeast colonies from the interaction selection (large square) plates are picked with a sterile toothpick and patched onto plates containing the appropriate selective media and incubated at 30°C for two days.

5 To further ensure purity of the yeast, the plates are replicated onto another plate containing the same media and incubated at 30°C for another two days.

Yeast patches are scraped using a sterile toothpick and placed into a 96-well format plate containing 100 μ l SD –L –W –H with 2% glucose liquid medium.

10 Half the volume of the plate is transferred to a 96-well plate containing 50 μ l of 40% glycerol for storage. The other half is set aside for replication and galactosidase-activity assay (see below).

Cells are replicated onto a SD –L –W –H plate with 2% glucose plate to create a master plate, and incubated two days at 30°C. The master plate is replicated onto different selective media to score the strength of each interaction.

15 Cells are also replicated onto media selecting for the prey vector only for colony PCR and incubated two days at 30°C.

Galactosidase activity assay

20 Ten microliters from the 96-well plate (set aside from above) are transferred into another 96-well plate containing 100 μ l SD and 2% glucose media. The cell density is measured at OD₆₀₀ using a spectrophotometer, the OD₆₀₀ is usually between 0.03 and 0.1. Fifty microliters of Galactosidase reaction mixture (Tropix) are added to microplates (Marsh) specifically designed for the luminometer (Hewlett Packard Lumicount). Fifty microliters of the diluted cells are then added and mixed by pipetting.

25 The reaction is incubated sixty to one hundred twenty minutes at room temperature. Relative Light Units (RLUs) are read by the luminometer. Each plate contains a negative control, constituted by diploid yeast containing the bait of interest and an empty prey vector. To be scored as positive, the diploids tested have to have an RLU number at least twice as high as the negative control.

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Example 2

Minimum interaction domain mapping

Further analysis of yeast two hybrid (Y2H) interacting proteins includes the dissection of protein motifs responsible for the interaction. Sequence alignment of multiple clones identified in the Y2H screens can help identify the smallest common region responsible for the interaction. In the absence of appropriate clones, deletion mapping of interacting domains is necessary.

PCR primers containing restriction sites suitable for cloning are designed to cover multiple sub-domains of the protein of interest (bait or prey). The methods involved in cloning, sequencing, yeast transformation, mating, and scoring of interactions are readily performed by one of ordinary skill in the art of molecular biology and genetic engineering.

Materials and Methods

Minimum interaction domain: primers were designed for PCR of the Dkk-1 clone isolated by screening a primary osteoblast cell strain (HOB03C5) library with pooled Zmax1/LRP5 ligand binding domain (LBD) baits: LBD1 (Leu969-Pro1376) and LBD4 (Arg1070-Pro1376). The primers, which are presented in 5' to 3' orientation, were as follows:

<u>SEQ ID NO</u>	<u>Primer</u>	<u>Sequence</u>
155	Forward 1	TTTTTTGTCGACCAATTCCAACGCTATCAAG
156	Forward 2	TTTTTTGTCGACCTGCGCTAGTCCCACCCGC
157	Forward 3	TTTTTTGTCGACCGTGTCTTCTGATCAAAATC
158	Forward 4	TTTTTTGTCGACCGGACAAGAAGGTTCTGTTTG
159	Reverse 1	TTTTTTGCGGCCGCTTATTTGGTGTGATACATTTTGTG
160	Reverse 2	TTTTTTGCGGCCGCTTAGCAAGACAGACCTTCTCC
161	Reverse 3	TTTTTTGCGGCCGCTTAGTGTCTCTGACAAGTGTG

PCR was performed using PfuTurbo® polymerase (Stratagene). The PCR products were gel purified, digested with *Sall*/ *NotI* and ligated to pPC86 (Gibco/BRL) which had been linearized with *Sall*/*NotI*. Clones were recovered and sequenced to ascertain that the structure was as expected and that the Gal4 activation domain and Dkk-1 were in-frame. The ORF of Dkk-1 was Met1-His266, as in human Dkk-1 (GenBank Accession No. XM_005730).

The clones used were as follows: D5 (F1/R3: Asn34-His266), D4 (F1/R2: Asn34-Cys245), D3 (F1/R1: Asn34-Lys182), D9 (F2/R3: Cys97-His266), D12 (F3/R3, Val139-His266), D14 (F4/R3: Gly183-His266), D8 (F2/R2: Cys97-Cys245), and D11 (F3/R2: Val139-Cys245). F1, F2, F3 and F4 refer respectively to Forward primers 1, 2, 3 and 4. R1, R2 and R3 refer respectively to reverse primers 1, 2 and 3.

These clones were transformed into yeast and mated with each of three yeast strains containing pDBleu (Gibco/BRL), pDBleuLBD1, and pDBleuLBD4. Positive interactions were detected by growth of the hybrids on appropriate selective media.

Results

Minimum interaction domain: Figure 6 shows that while growth was observed in diploids of D4, D5, D8, D9, and D12, no growth was observed in hybrids of D3, D11, and D12. Carboxy terminal (C-terminal) deletions indicated that while the C-terminal amino acids of Dkk-1 containing the potential N-glycosylation site (Arg246-His266) are not required for interaction with Zmax1/LRP5 LBD baits, the Cys2 domain, Gly183-Cys245, is required. N-terminal deletions also demonstrated that the region between the two cysteine domains, *i.e.* Val139 to Lys182, is also required. Two minimum interaction domain constructs were isolated: D12 (Val139-His266) and D8 (Cys97-Cys245). Similar constructs could be prepared for Dkk-1 interactors.

Example 3

Yeast-2 Hybrid screen for peptide aptamer sequences to Dkk-1

Peptide aptamer library construction

A peptide aptamer library, Tpep, was constructed, which provides a means to identify chimeric proteins that bind to a protein target (or bait) of interest using classic yeast two hybrid (Y2H) assays. The Tpep library is a combinatorial aptamer library composed of constrained random peptides, expressed within the context of the disulfide loop of *E. coli* thioredoxin (trxA), and as C-termini fusion to the *S. cerevisiae* Gal4 activation domain. The Tpep library was generated using a restriction enzyme modified recombinant Y2H prey vector, pPC86 (Gibco), which contains the trxA scaffold protein.

Generation of aptamer-encoding sequences

Aptamer-encoding sequences were produced as follows. DNA encoding random stretches of approximately sixteen amino acids surrounded by appropriate restriction sites were generated by semi-random oligonucleotide synthesis. The synthetic oligonucleotides were PCR-amplified, restriction digested, and cloned into the permissive sites within the trxA scaffold protein. The cloning strategy was to insert the random oligonucleotide sequence is in-frame with the scaffold protein coding sequence, resulting in expression of a scaffold protein-aptamer chimera. The scaffold protein is itself in-frame with the activation domain of Gal4, within the pPC86 vector that is appropriate for the aptamer to be expressed and functional in a regular Y2H assay. Additional methods of preparing aptamers would be apparent to the skilled artisan.

Generation of a permissive recombinant pPC86 vector containing the trxA coding sequence

First the *RsrII* restriction site located within the Gal4 activation domain of pPC86 (Gibco) was eliminated by site-directed mutagenesis (Quickchange™ kit, Stratagene). The amino acid sequence of the Gal4 activation domain was unchanged by this modification. The strength of different control interactions was verified to be unchanged by the modification.

Second, the *E. coli* trxA coding sequence was cloned into the *Sall* and *NotI* sites of the *RsrII*-modified pPC86. *EcoRI* and *SpeI* sites were then introduced within the trxA

*Rsr*II site. The oligonucleotides encoding the peptide aptamers were cloned into the *Eco*RI and *Spe*I sites of the resulting vector.

Example 4

Yeast-2 Hybrid screen for Dkk-1 interacting proteins

A Dkk-1 bait sequence was utilized in a yeast two hybrid screen to identify Dkk-1 interacting proteins. The procedure for the Y2H was carried out similarly to that employed in Example 1, except that the Dkk-1 bait from Figure 2C was used instead of LRP baits. The screen was performed using Hela and fetal brain libraries (Invitrogen Corporation, Carlsbad, CA). Multiple libraries were used to identify additional Dkk-1 interacting proteins and to confirm interactions found in other libraries.

The list of Dkk-1 interacting proteins uncovered in these Y2H screens are listed in Figure 5.

The interacting proteins identified in the Dkk-1 bait screen can be used in other Y2H screens with LRP baits and other Dkk-1 interacting proteins to determine more complex interactions which may modulate Dkk-1/LRP interactions and/or Wnt signaling.

Example 5

Generation of antibodies

In each of the following antibody-generating examples, the synthesis of these linear peptides is followed by injection into two New Zealand Rabbits. Subsequent boosts and bleeds are taken according to a standard ten-week protocol. The end-user receives back 5 mgs of peptide, aliquots of pre-bleeds, roughly 80 ml of crude sera from each of the two rabbits and, and ELISA titration data is obtained.

Generation of LRP5 Polymorphism-specific antibodies

Antibodies were generated to the following peptides to obtain antibodies which distinguish the HBM polymorphism versus wild-type LRP5/Zmax: MYWTDWVETPRIE

(SEQ ID NO:123) (mutant peptide) and MYWTDWGETPRIE (SEQ ID NO:124) (wild-type peptide for negative selection). Immunofluorescence data confirmed that the antibody, after affinity purification, is specific for HBM and does not recognize LRP5 (Figure 17).

5

Generation of LRP5 Monospecific antibodies

LRP5 monospecific polyclonal antibodies were generated to the following amino acid sequences of LRP5: Peptide 1 (a.a. 265-277) - KRTGGKRKEILSA (SEQ ID NO:125), Peptide 2 (a.a. 1178-1194) - ERVEKTTGDKRTRIQGR (SEQ ID NO:126), and Peptide 3 (a.a. 1352-1375) - KQQCDSFPDCIDGSDE (SEQ ID NO:127).

10

Immunofluorescence confirmed that the antibody generated detects LRP5.

Generation of Dkk-1 monospecific polyclonal antibodies

Dkk-1 monospecific polyclonal antibodies were generated to the following amino acid sequences of Dkk-1: Peptide 1 (a.a. 71-85) - GNKYQTIDNYQPYPYPC (SEQ ID NO:118), Peptide 2 (a.a. 165-186) - LDGYSRRTTLSSKMYHTKGQEG (SEQ ID NO:119), Peptide 3 (a.a. 246-266) - RIQKDHHQASNSSLHTCQRH (SEQ ID NO:120), Peptide 4 (a.a. 147-161) - RGEIETITESFGND (SEQ ID NO:121), and Peptide 5 (232-250) - EIFQRCYCGEGLSCRIQKD (SEQ ID NO:122) of human Dkk-1.

15

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Figure 26 shows the location of the various peptides selected, their relationship to the Dkk-1 amino acid sequence and polyclonal antibodies generated.

Western blots demonstrated that the antibodies generated against peptides 2 (Antibody #5521) (Figure 27) and 4 (Antibody #74397) (Figure 28) are specific toward Dkk-1. Figure 27 shows Western blots using 500 μ l of conditioned medium (CM) from non-transfected 293 cells or from 293 cells transfected with Dkk1-V5 that were immunoprecipitated by anti-V5 antibody. Bead elutes were separated by non-reducing SDS-PAGE (lanes #4, 5 of Figure 27). 20 μ l of conditioned medium from both samples (lanes #2, 3 of Figure 27) and from Dkk1-AP transfected 293 cells (lane #6 of Figure 27) were additionally separated on the gel. The Western was performed using

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antibodies Anti-V5/AP (1:10,000) and Ab#5521 (10 μ g/ml). Ab#5521 detected Dkk1-V5 and Dkk1-AP from conditioned medium.

Figure 28 shows Western blot results using Ab#74397. Anti-V5/AP was tested at a 1:4000 dilution and Ab#74397 was tested at a 1:500 dilution. Ab#74397 was able to detect Dkk1-V5 in both conditioned medium and immunoprecipitated conditioned medium.

The results obtained with antibodies #5521 and #74397 are summarized in the following table:

Rabbit No.	Peptide Position	Peptide Sequence	Purified (Y/N)	Western	Immuno-precipitation	Location
5521	165-186	LDGYSR RTTLSSK MYHTKG QEG	Y (Protein G purified)	Y	N/A	Between Cy1 and Cys2 domain
74397	147-161	RGEIETI TESFGN D	N	Y	N/A	Between Cy1 and Cys2 domain

Example 6

Effects of exogenous Dkk-1 on Wnt-mediated signaling in the *Xenopus* embryo assay

Xenopus embryos are an informative and well-established *in vivo* assay system to evaluate the modulation of Wnt signaling (McMahon *et al.*, *Cell* 58: 1075-84 (1989); Smith and Harland, 1991; reviewed in Wodarz and Nusse 1998).

Modification of the Wnt signaling pathway can be visualized by examining the embryos for a dorsalization phenotype (duplicated body axis) after RNA injection into the ventral blastomere at the 4- or 8-cell stage. On the molecular level, phenotypes can be analyzed by looking for expression of various marker genes in stage 10.5 embryos. Such markers would include general endoderm, mesoderm, and ectoderm markers as well as a variety of tissue-specific transcripts.

Analysis can be done by RT-PCR/TaqMan® and can be done on whole embryo tissue or in a more restricted fashion (microdissection). Because this system is very flexible and rapid, by injecting combinations of transcripts, such as HBM and different Wnts or Wnt antagonists, the mechanism of HBM in the Wnt pathway can thereby be dissected. Furthermore, investigations are conducted to determine whether Zmax/LRP5 and HBM differentially modulate Wnt signaling either alone, or in combination with other components. Previous studies have demonstrated that LRP6 alone or LRP5 + Wnt5a were able to induce axis duplication (dorsalization) in this system (Tamai *et al.*, *Nature* 407: 530-35 (2000)).

Constructs for *Xenopus* Expression (Vector pCS2⁺)

Constructs were prepared using the vector pCS2⁺. DNA inserts were subcloned in the sense orientation with respect to the vector SP6 promoter. The pCS2⁺ vector contains an SV40 virus polyadenylation signal and T3 promoter sequence (for generation of antisense mRNA) downstream of the insert.

Full length Zmax/LRP5 and HBM ORF cDNA: Insert cDNA was isolated from the full length cDNA retrovirus constructs (with optimized Kozak sequences) by *Bgl*II-*Eco*RI digestion and subcloned into the *Bam*HI-*Eco*RI sites of the pCS2⁺ vector.

Full length XWnt8: This cDNA was PCR amplified from a *Xenopus* embryo cDNA library using oligos 114484 (SEQ ID NO:162) (5'-CAGTGAATTCACCATGCAAAACACCACTTTGTTC-3') and 114487 (SEQ ID NO:163) (5'-CAGTTGCGGCCGCTCATCTCCGGTGGCCTCTG-3'). The oligos were designed to amplify the ORF with a consensus Kozak sequence at the 5' end as determined from GenBank #X57234. PCR was carried out using the following conditions: 96°C, 45 sec.; 63°C, 45 sec.; 72°C, 2 min. for 30 cycles. The resulting PCR product was purified, subcloned into pCRII-TOPO (Invitrogen Corp.), sequence verified, and digested with *Bam*HI/*Xho*I. This insert was subcloned into the vector at the *Bam*HI-*Xho*I sites.

Full length Wnt5a: A murine Wnt5a cDNA clone was purchased from Upstate Biotechnology (Lake Placid, NY) and subcloned into the *Eco*RI site of the vector. Sequencing confirmed insert orientation.

Full length human Dkk-1: A human cDNA with GenBank accession number AF127563 was available in the public database. Using this sequence, PCR primers were designed to amplify the open reading frame with a consensus Kozak sequence immediately upstream of the initiating ATG. Oligos 117162 (SEQ ID NO:164) (5'-
5 CAATAGTCGACGAATTCACCATGGCTCTGGGCGCAGCGG-3') and 117163 (SEQ ID NO:165) (5'-GTATTGCGGCCGCTCTAGATTAGTGTCTCTGACAAGTGTGAA-3') were used to screen a human uterus cDNA library by PCR. The resulting PCR product was purified, subcloned into pCRII-TOPO (Invitrogen Corp.), sequence verified, and digested with *EcoRI/XhoI*. This insert was subcloned into the pCS2⁺ vector at the
10 *EcoRI-XhoI* sites.

Full length human Dkk-2: A full length cDNA encoding human Dkk-2 was isolated to investigate the specificity of the Zmax/LRP5/HBM interaction with the Dkk family of molecules. Dkk-1 was identified in yeast as a potential binding partner of Zmax/LRP5/HBM. Dkk-1 has also been shown in the literature to be an antagonist of
15 the Wnt signaling pathway, while Dkk-2 is not (Krupnik *et al.*, 1999). The Dkk-2 full length cDNA serves as a tool to discriminate the specificity and biological significance of Zmax/LRP5/HBM interactions with the Dkk family (e.g., Dkk-1, Dkk-2, Dkk-3, Dkk-4, Soggy, their homologs and variant, etc.). A human cDNA sequence for Dkk-2 (GenBank Accession No. NM_014421) was available in the public database. Using this
20 sequence, PCR primers were designed to amplify the open reading frame with a consensus Kozak sequence immediately upstream of the initiating ATG. Oligos 51409 (SEQ ID NO:166) (5'-CTAACGGATCCACCATGGCCGCGTTGATGCGG-3') and 51411 (SEQ ID NO:167) (5'-GATTCTGAATTCTCAAATTTCTGACACACATGG-3') were used to screen human embryo and brain cDNA libraries by PCR. The resulting
25 PCR product was purified, subcloned into pCRII-TOPO, sequence verified, and digested with *BamHI/EcoRI*. This insert was subcloned into the pCS2⁺ vector at the *BamHI-EcoRI* sites.

Full length LRP6 was isolated from the pED6dpc4 vector by *XhoI-XbaI* digestion. The full length cDNA was reassembled into the *XhoI-XbaI* sites of pCS2⁺. Insert
30 orientation was confirmed by DNA sequencing.

mRNA Synthesis and Microinjection Protocol

mRNA for microinjection into *Xenopus* embryos is generated by *in vitro* transcription using the cDNA constructs in the pCS2⁺ vector described above as template. RNA is synthesized using the Ambion mMessage mMachine high yield capped RNA transcription kit (Cat. #1340) following the manufacturer's specifications for the Sp6 polymerase reactions. RNA products were brought up to a final volume of 50 µl in sterile, glass-distilled water and purified over Quick Spin Columns for Radiolabelled RNA Purification G50-Sephadex (Roche, Cat. #1274015) following the manufacturer's specifications. The resulting eluate was finally extracted with phenol:chloroform:isoamyl alcohol and isopropanol precipitated using standard protocols (Sambrook *et al.*, 1989). Final RNA volumes were approximately 50 µl. RNA concentration was determined by absorbance values at 260 nm and 280 nm. RNA integrity was visualized by ethidium bromide staining of denaturing (formaldehyde) agarose gel electrophoresis (Sambrook *et al.*, 1989). Various amounts of RNA (2 pg to 1 ng) are injected into the ventral blastomere of the 4- or 8-cell *Xenopus* embryo. These protocols are described in Moon *et al.*, *Technique-J. of Methods in Cell and Mol. Biol.* 1: 76-89 (1989), and Peng, *Meth. Cell. Biol.* 36: 657-62 (1991).

Screening for Duplicated Body Axis

In vitro transcribed RNA is purified and injected into a ventral blasomere of the 4- or 8-cell *Xenopus* embryo (approx. 2 hours post-fertilization). At stage 10.5 (approx. 11 hours post-fertilization), the injected embryos are cultured for a total of 72 hours and then screened for the presence of a duplicated body axis (dorsalization) (Figure 7). Using XWnt8-injected (2-10 pg) as a positive control (Christian *et al.* (1991)) and water-injected or non-injected embryos as negative controls, we replicated the published observation that Zmax(LRP5) + Wnt5a (500 and 20 pg, respectively) could induce axis duplication. Wnt5a (20 pg) alone could not induce axis duplication (as previously reported by Moon *et al.* (1993)). We have also injected GFP RNA (100-770 pg) as a negative control to show that the amount of RNA injected is not perturbing embryo development (not shown). Strikingly, HBM + Wnt5a (500 and 20 pg, respectively)

yielded an approximately 3.5 fold more robust response of the phenotype ($p=0.043$ by Fisher's exact test) compared to Zmax(LRP5) + Wnt5a, suggesting that the HBM mutation is activating the Wnt pathway (Figures 8 and 9). The HBM/Wnt5a embryos also appear to be more "anteriorized" than the Zmax(LRP5)/Wnt5a embryos, again suggestive of a gain-of-function mutation.

The role of Dkk-1 as a modulator of Zmax/LRP5- and HBM-mediated Wnt signaling was investigated. Literature reports have previously characterized *Xenopus* and murine Dkk-1 as antagonists of the canonical Wnt pathway in the *Xenopus* system (Glinka *et al.*, *Nature* 391:357-362 (1998)). Using the human Dkk-1 construct, a dose-response assay was performed to confirm that our construct was functional and to identify the optimal amount of RNA for microinjection. Using 250 pg/embryo of hDkk-1 RNA, over 90% ($p<0.001$) of the embryos were observed to display enlarged anterior structures (big heads) as anticipated from the published reports (Figure 10).

The mechanism of hDkk-1 modulation of Wnt signaling in the presence of Zmax/LRP5 or HBM was also investigated. Without any hDkk-1 present, it was confirmed that HBM + Wnt5a was a more potent activator of Wnt signaling than Zmax/LRP5 + Wnt5a ($p<0.05$). Interestingly, in the presence of hDkk-1 (250 pg), Zmax/LRP5-mediated Wnt signaling was repressed ($p<0.05$) but hDkk-1 was unable to repress HBM-mediated Wnt signaling ($p<0.01$) (Figure 11). The specificity of this observation can be further addressed by investigating other members of the Dkk family, other Wnt genes, LRP6, additional Zmax/LRP5 mutants, and the peptide aptamers.

Example 7

Effects of exogenous Dkk and LRP5 on Wnt signaling in the TCF-luciferase Assay

Wnt activity can be antagonized by many proteins including secreted Frizzled related proteins (SFRPs), Cerberus, Wnt Inhibitory Factor-1 and Dkk-1 (Krupnik *et al.*, 1999). The Dkk family of proteins consists of Dkk-1-4 and Soggy, a Dkk-3-like protein. Dkk-1 and Dkk-4 have been shown to antagonize Wnt mediated *Xenopus* embryo development, whereas Dkk-2, Dkk-3, and Soggy do not. Unlike many of these proteins

that antagonize Wnt activity by directly interacting with Wnt proteins, Dkk-1 acts by binding to two recently identified Wnt coreceptors, LRP5 and LRP6. (Mao *et al.*, 2001; Bafico *et al.*, 2001). The details of this interaction have been examined by the present inventors and Mao *et al.* using deletion constructs of LRP6, which demonstrated that EGF repeats 3 and 4 are important for Dkk-1 interaction. Accordingly, the activity of two Dkk proteins, Dkk-1 and Dkk-2, were investigated with various Wnt members, LRP5, LRP6, and the mutant form of LRP5, designated HBM. The present invention explores whether there is any functional difference between LRP5 and HBM with regard to Dkk action on Wnt mediated signaling. Various reagents were developed, including Dkk-1 peptides, constrained LRP5 peptide aptamers, constrained Dkk-1 peptide aptamers and polyclonal antibodies to Dkk-1 (in Example 5 above) to identify factors that mimic HBM mediated Wnt signaling.

Methods

Various LRP5 constrained peptides were developed. Specifically, four peptides that interact with the LBD of LRP5 (Figure 4, constructs OST259-262 in Figure 12) and three peptides that interact with the cytoplasmic domain of LRP5 (constructs OST266-OST268 in Figure 12). In addition two Dkk-1 peptides were developed: constructs OST264 and OST265 in Figure 12, corresponding to Dkk-1 amino acids 139-266 and 96-245, containing the smallest region of Dkk-1 that interacts with LRP5 (Figure 6). The cDNA clones encoding the LRP5 LBD interacting peptides and the Dkk-1 peptides were subcloned into pcDNA3.1 with the addition of a Kozak and signal sequence to target the peptide for secretion. The constructs encoding the three peptides interacting with the cytoplasmic domain of LRP5 were also subcloned into pcDNA3.1. However, these latter constructs do not contain a signal sequence.

HOB-03-CE6 osteoblastic cells developed by Wyeth Ayerst (Philadelphia, PA) were seeded into 24-well plates at 150,000 cells per well in 1 ml of the growth media (D-MEM/F12 phenol red-free) containing 10% (v/v) heat-inactivated FBS, 1X penicillin streptomycin, and 1X Glutamax-1, and incubated overnight at 34°C. The following day, the cells were transfected using Lipofectamine 2000® (as described by the

manufacturer, Invitrogen) in OptiMEM (Invitrogen) with 0.35 μ g /well of LRP5, HBM, or control plasmid DNA (empty vector pcDNA3.1) and either Wnt1 or Wnt3a plasmid DNA. Similar experiments were performed with LRP6 plasmid DNA (0.35 μ g/well) or a control pEDdpc4 empty vector. Furthermore, each of these groups were then divided
5 into three groups, those receiving 0.35 μ g/well Dkk-1, Dkk-2, or pcDNA3.1 control DNA. All wells were transfected with 0.025 μ g/well of CMV beta-galactosidase plasmid DNA and 0.35 μ g/well 16X TCF(AS)-luciferase reporter DNA (developed by Ramesh Bhat, Wyeth-Ayerst (Philadelphia, PA)). After 4 hours of incubation, the cells were rinsed and 1 ml of fresh growth media was added to each well. The cells were cultured overnight
10 at 34°C, followed by a wash and a change of media. Cells were cultured for an additional 18-24 hours at 37°C. Cells were then lysed with 50 μ l/well of 1X lysis buffer. The extracts were assayed for beta-galactosidase activity (Galacto Reaction Buffer Diluent & Light Emission Accelerator, Tropix) using 5 μ l extract + 50 μ l beta-galactosidase diluent and luciferase activity (Luciferase Assay Reagent, Promega)
15 using 20 μ l extract.

U2OS human osteosarcoma cells were also utilized. U2OS cells (ATCC) were seeded into 96-well plates at 30,000 cells per well in 200ul of the growth media (McCoy's 5A) containing 10% (v/v) heat-inactivated FBS, 1X penicillin streptomycin, and 1X Glutamax-1, and incubated overnight at 37°C. The following day, the media
20 was replaced with OptiMEM (Invitroge) and cells were transfected using Lipofectamine 2000® (as described by the manufacturer, Invitrogen) with 0.005 μ g/well of LRP5, HBM, LRP6 or control plasmid DNA (empty vector pcDNA3.1) and either Wnt1 (.0025ug/well) or Wnt3a (.0025ug/well) plasmid DNA. In addition, the 16x-(AS) TCF-TK-firefly-luciferase (Ramesh Bhat, WHRI, Wyeth) and control TK-renilla luciferase (Promega
25 Corp.) were co-transfected at 0.3ug/well and 0.06ug/well respectively in all experiments. Furthermore, each of these groups was then divided into different groups, those receiving 0.05ug/well Dkk-1, Dkk-2, Dkk3, Dkk1-Alkaline Phosphatase (AP), mutant Dkk-1 (C220A), Soggy or pcDNA3.1 control DNA. In other experiments, cells were co-transfected with 0.005 μ g/well of LRP5, 0.0025ug/well of Wnt1 or Wnt3a (using
30 0.0025 μ g/well of a control pcDNA3.1) with LRP5-interacting aptamers (0.05ug/well).

Cells were cultured for an additional 18-20 hours at 37°C. Culture medium was removed. Cells were cultured for an additional 18-20 hours at 37°C. Culture medium was removed. Cells were then lysed with 100 μ l/well of 1X Passive Lysis Buffer (PLB) of Dual Luciferase Reagent kit (DLR-kit-Promega Corp.) 20 μ l of the lysates were
5 combined with LARII reagent of DLR-kit and assayed for TCF-firefly luciferase signal in Top Count (Packard) instrument. After measuring the Firefly readings, 100 μ l of the "Stop and Glo" reagent of DLR kit that contains a quencher and a substrate for renilla luciferase was added into each well. Immediately the renilla luciferase reading was measured using the Top Count (Packard) Instrument. The ratios of the TCF-firefly
10 luciferase to control renilla readings were calculated for each well and the mean ratio of triplicate or more wells was expressed in all data.

Results

The results of these experiments demonstrate that Dkk-1, in the presence of
15 Wnt1 and LRP5, significantly antagonized TCF-luciferase activity (Figure 14). In marked contrast, Dkk-1 had no effect on HBM/Wnt1 mediated TCF-luciferase activity (Figure 14). In similar experiments, Dkk-1 was also able to antagonize LRP5/Wnt3a but not HBM/Wnt3a mediated TCF-luciferase activity (Figure 15). These results indicate that the HBM mutation renders Dkk-1 inactive as an antagonist of Wnt1 and
20 Wnt3a signaling in HOB03CE6 osteoblastic cells. In other experiments with Wnt1, Dkk-1 had no effect on LRP5 or HBM mediated TCF-luciferase activity (Figure 14). In contrast, with either LRP5 or HBM in the presence of Wnt3a, Dkk-2 was able to antagonize the TCF-luciferase activity (Figure 15). These latter results indicate that the HBM mutation has no effect on Dkk-2 action in the presence of Wnt3a. Experiments
25 were also performed using the closely related LRP6 cDNA in HOB-03-CE6 cells. In these experiments, LRP6/Wnt1 and LRP6/Wnt3a mediated TCF-luciferase were regulated in the same manner as LRP5. Specifically, Dkk-1 antagonized LRP6/Wnt1 mediated TCF-luciferase activity, whereas Dkk-2 had no effect (Figure 14). However, similar to the action of Dkk-2 with LRP5/Wnt3a, Dkk-2 was able to antagonize
30 LRP6/Wnt3a mediated TCF-luciferase activity (Figure 15).

The results in the U2OS cells show a robust effect of the OST262 LRP5 peptide aptamer activation of Wnt signaling in the presence of Wnt3a (Figure 16). These functional results are confirmed by the results shown below in Example 11 using LRP5 peptide aptamers in the Xenopus assay. Such results affirmatively demonstrate that the effects of small molecules on LRP5/LRP6/HBM signaling can be detected using the TCF-luciferase assay.

These data demonstrate that there is a functional difference between LRP5 and HBM regarding the ability of Dkk-1 to antagonize Wnt1 and Wnt3a signaling. These data and previous data showing that Dkk-1 directly interacts with LRP5 suggests that the inability of Dkk-1 to antagonize HBM/Wnt signaling may in part contribute to the HBM phenotype. These experiments further demonstrate the ability to test various molecules (e.g., small molecules, aptamers, peptides, antibodies, LRP5 interacting proteins or Dkk-1 interacting proteins, and the like) for a LRP5 ligand that mimics HBM mediated Wnt signaling or factors that block Dkk-1 interaction with LRP5.

Example 8

Yeast-2 Hybrid Interaction Trap

Small molecule inhibitors (or partial inhibitors) of the Dkk-LRP interaction may be an excellent osteogenic therapeutic. One way to investigate this important protein-protein interaction is using Y2H techniques substantially as described above and as is well known in the art. Regions of LRP5, such as LRP5 LBD, have been found to functionally interact with Dkk. This interaction is quantitated using a reporter element known in the art, e.g., LacZ or luciferase, which is only activated when bait and prey interact. The Y2H assay is used to screen for compounds which modulate the LRP-Dkk interaction. Such a modulation would be visualized by a reduction in reporter element activation signifying a weaker or disrupted interaction, or by an enhancement of the reporter element activation signifying a stronger interaction. Thus, the Y2H assay can be used as a high-throughput screening technique to identify compounds which disrupt or enhance Dkk interaction with LRP5/LRP6/HBM, which may serve as potential therapeutics.

For example, the Interaction Trap methodology can be used as follows. The LRP5 LBD, for example, was fused with LexA and Dkk-1 was fused with either Gal4-AD or B42. With the LRP5LBD-LexA bait and the Gal4AD-Dkk prey, over a 20-fold activation of a lacZ reporter (under the control of a single LexA operator) was detected over the background. Using a Dkk-1 mutant (C220A) that is unable to bind to LRP, the interaction was reduced in yeast, showing the specificity of this interaction and system (Figure 18). As a result, small molecules may be identified that modulate this interaction between LRP and Dkk.

Example 9

Cell-Based Functional High-Throughput Assay

To develop a high throughput assay, the TCF-luciferase assay described in Example 7 was modified utilizing low level expression of endogenous LRP5/6 in U2OS and HEK293 cells. However, HOB-03-CE6 cells and any other cells which show a differential response to Dkk depending on whether LRP5, LRP6 or HBM are expressed. Using U2OS (human osteosarcoma) and HEK293 (ATCC) cells, the TCF-luciferase and tk-Renilla reporter element constructs were co-transfected along with Wnt3a/1 and Dkk. Wnt3a alone, by using endogenous LRP5/6, was able to stimulate TCF reporter gene activation. When Dkk, is co-transfected with Wnt3a/Wnt 1 and reporters (TCF-luci and tk-Renilla), Dkk represses reporter element activity. In addition, the TCF-luci signal is activated by Wnt3a/Wnt1 can be repressed by the addition of Dkk-enriched conditioned media to the cells containing Wnt3a/Wnt1 and reporters. The assay is further validated by the lack of TCF-reporter inhibition by a point mutant construct (C220A) of Dkk1.

The Dkk-mediated repression of the reporter is dependent upon the concentration of transfected Dkk cDNA or on the amount of Dkk-conditioned media added. In addition, the Dkk-mediated reporter suppression can be altered by the co-transfection of LRP5, LRP6, and HBM cDNAs in the U2OS or HEK293 cells. In general, U2OS cells show greater sensitivity to Dkk-mediated reporter suppression than that in HEK-293 cells. In U2OS cells, the transfection of LRP5/LRP6/HBM cDNA leads

to moderate activation of TCF-luci in the absence of Wnt3a/Wnt1 transfection. This activation presumably utilizes the endogenous Wnts present in U2OS cells. Under this condition, Dkk1 can repress TCF-luci and shows a differential signal between LRP5 and HBM. By co-transfecting Wnt3a/Wnt1, there is a generalized increase in the TCF-luci signal in the assay. Further, one can detect Dkk-mediated differential repression of the reporter due to LRP5 and HBM cDNA expression as well as between LRP5 and LRP6 cDNA. The repression is maximal with LRP6, moderate with LRP5, and least with HBM cDNA expression. In addition, the assay can detect the functional impact of the LRP5 interacting peptide aptamers (Figure 4), Dkk1 interacting aptamers and binding domains of Dkk-1 (Figure 6; OST264 and OST265 of Figures 12 and 13).

Using this system with a suppressed Wnt-TCF signal due to the presence of both Dkk and Wnt3a, one can screen for compounds that could alter Dkk modulation of Wnt signaling, by looking for compounds that activate or the TCF-luciferase reporter, and thereby relieve the Dkk-mediated repression of the Wnt pathway. Such compounds identified may potentially serve as HBM-mimetics and be useful, for example, as osteogenic therapeutics. Data generated from this high throughput screen are demonstrated in Figures 19-21. Figure 19 shows that Dkk1 represses Wnt3a-mediated signaling in U2OS bone cells. Figure 20 demonstrates the functional differences between LRP5, LRP6, and HBM. Dkk-1 represses LRP6 and LRP5 but has little or no effect on HBM-generated Wnt1 signaling in U2OS cells. Figure 21 demonstrates the differential effects of various Dkk family members and modified Dkks, including Dkk-1, a mutated Dkk-1 (C220A), Dkk-1-AP (modified with alkaline phosphatase), Dkk-3, and Soggy.

Example 10

DKK/LRP5/6/HBM ELISA Assay

A further method to investigate Dkk binding to LRP is via ELISA assay. Two possible permutations of this assay are exemplified. LRP5 is immobilized to a solid surface, such as a tissue culture plate well. One skilled in the art will recognize that other supports such as a nylon or nitrocellulose membrane, a silicon chip, a glass slide,

beads, etc. can be utilized. In this example, the form of LRP5 used is actually a fusion protein where the extracellular domain of LRP5 is fused to the Fc portion of human IgG. The LRP5-Fc fusion protein is produced in CHO cell extracts from stable cell lines. The LRP5-Fc fusion protein is immobilized on the solid surface via anti-human Fc antibody
5 or by Protein-A or Protein G-coated plates, for example. The plate is then washed to remove any non-bound protein. Conditioned media containing secreted Dkk protein or secreted Dkk-epitope tagged protein (or purified Dkk or purified Dkk-epitope tagged protein) is incubated in the wells and binding of Dkk to LRP is investigated using antibodies to either Dkk or to an epitope tag. Dkk-V5 epitope tagged protein would be
10 detected using an alkaline phosphatase tagged anti-V5 antibody.

Alternatively, the Dkk protein could be directly fused to a detection marker, such as alkaline phosphatase. Here the detection of the Dkk-LRP interaction can be directly investigated without subsequent antibody-based experiments. The bound Dkk is detected in an alkaline phosphatase assay. If the Dkk-alkaline phosphatase fusion
15 protein is bound to the immobilized LRP5, alkaline phosphatase activity would be detected in a colorimetric readout. As a result, one can assay the ability of small molecule compounds to alter the binding of Dkk to LRP using this system. Compounds, when added with Dkk (or epitope-tagged Dkk) to each well of the plate, can be scored for their ability to modulate the interaction between Dkk and LRP based
20 on the signal intensity of bound Dkk present in the well after a suitable incubation time and washing. The assay can be calibrated by doing cold competition experiments with unlabeled Dkk or with a second type of epitope-tagged Dkk. Any small molecule that is able to modulate the Dkk-LRP interaction may be a suitable therapeutic candidate, more preferably an osteogenic therapeutic candidate.

Example 11

Functional Evaluation of Peptide Aptamers in Xenopus

The constrained peptide aptamers constructs OST258-263 (where 258 contains the signal sequence by itself and 263 contains an irrelevant constrained peptide)
30 (Figures 12 and 13) were used to generate RNA substantially as described in Example

7, except the vector was linearized by restriction endonuclease digestion and RNA was generated using T7 RNA polymerase.

Aptamer RNA was injected at 250 pg per blastomere using the protocol of Example 7. Wnt signaling was activated, as visualized by embryo dorsalization (duplicated body axis) with aptamers 261 and, more strongly, 262. The results of this assay are shown in Figures 22 and 23. These results suggest that aptamers 261 and 262 are able to activate Wnt signaling possibly by binding to the LBD of LRP, thereby preventing the modulation of LRP-mediated signaling by Dkk.

The aptamers of the present invention can serve as HBM-mimetics. In the *Xenopus* system they are able to induce Wnt signaling all by themselves. They may also serve as tools for rational drug design by enhancing the understanding of how peptides are able to interact with LRP and modulate Wnt signaling at the specific amino acid level. Thus, one would be able to design small molecules to mimic their effects as therapeutics. In addition, the aptamers identified as positives in this assay may be used as therapeutic molecules themselves.

Example 12

Homogenous Assay

An excellent method to investigate perturbations in protein-protein interactions is via Fluorescence Resonance Energy Transfer (FRET). FRET is a quantum mechanical process where a fluorescent molecule, the donor, transfers energy to an acceptor chromophore molecule which is in close proximity. This system has been successfully used in the literature to characterize the intermolecular interactions between LRP5 and Axin (Mao et al., *Molec. Cell Biol.* 7:801-809). There are many different fluorescent tags available for such studies and there are several ways to fluorescently tag the proteins of interest. For example, CFP (cyan fluorescent protein) and YFP (yellow fluorescent protein) can be used as donor and acceptor, respectively. Fusion proteins, with a donor and an acceptor, can be engineered, expressed, and purified.

For instance, purified LRP protein, or portions or domains thereof, fused to CFP and purified Dkk protein, or portions or domains thereof that interact with Dkk or LRP

respectively, fused to YFP can be generated and purified using standard approaches. If LRP-CFP and Dkk-YFP are in close proximity, the transfer of energy from CFP to YFP will result in a reduction of CFP emission and an increase in YFP emission. Energy is supplied with an excitation wavelength of 450 nm and the energy transfer is recorded at emission wavelengths of 480 nm and 570 nm. The ratio of YFP emission to CFP emission provides a guage for changes in the interaction between LRP and Dkk. This system is amenable for screening small molecule compounds that may alter the Dkk-LRP protein-protein interaction. Compounds that disrupt the interaction would be identified by a decrease in the ratio of YFP emission to CFP emission. Such compounds that modulate the LRP-Dkk interaction would then be considered candidate HBM mimetic molecules. Further characterization of the compounds can be done using the TCF-luciferase or Xenopus embryo assays to elucidate the effects of the compounds on Wnt signaling.

While the above example describes a cell-free, solution-phase assay using purified components, a similar cell-based assay could also be performed. For example, LRP-CFP fusion protein can be expressed in cells. The Dkk-YFP fusion protein then could be added to the cells either as purified protein or as conditioned media. The interaction of LRP and Dkk is then monitored as described above.

All references cited herein are hereby incorporated by reference in their entirety for all purposes. The following applications are also incorporated by reference in their entirety herein for all purposes: U.S. Application No. 60/290,071, filed May 11, 2001; U.S. Application No. 09/544,398, filed on April 5, 2000; U.S. Application No. 09/543,771, filed April 5, 2000; 09/578,900; U.S. Application No. 09/229,319, filed January 13, 1999; U.S. Provisional Application 60/071,449, filed January 13, 1998; and International Application PCT/US00/16951, filed June 21, 2000; International PCT Application entitled "HBM Variants That Modulate Bone Mass and Lipid Levels," filed May 13, 2002; and International PCT Application entitled "Transgenic Animal Model of Bone Mass Modulation," filed May 13, 2002. Additionally, this application claims priority to U.S. provisional applications 60/291,311, filed May 17, 2001; 60/353,058, filed

February 1, 2002; and 60/361,293, filed March 4, 2002; the texts of which are herein incorporated by reference in their entirety for all purposes.

CLAIMS

We claim:

- 5 1. A method of regulating LRP5, LRP6, or HBM activity in a subject comprising administering a composition which modulates a Dkk activity in an amount effective to regulate LRP5, LRP6, or HBM activity.
- 10 2. The method of any of Claims 1, 24, 28, 33, 36, 37, 48, 64, 65, 93, 98, 101, 105, 107, 111, or 112, wherein the Dkk is Dkk-1.
- 15 3. The method of any of Claims 1, 24, 28, or 33, wherein the Dkk is Dkk-1 and the Dkk activity is inhibited.
- 20 4. The method of Claims 1 or 24, wherein the Dkk activity modulates bone mass and/or lipid levels.
- 25 5. The method of Claim 4, wherein bone mass is increased and/or lipid levels are decreased.
- 30 6. The method of Claim 5, wherein the increase in bone mass is determined via one or more of a decrease in fracture rate, an increase in bone strength, an increase in bone density, an increase in bone mineral density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density, an increase in bone diameter, and an increase in inorganic bone content.
7. The method of any of Claims 1, 24, 28, or 33, wherein said composition comprises one or more compounds selected from the group consisting of Dkk interacting proteins, or a Dkk-binding fragment thereof.

8. The method of any of Claims 1, 24, 28, or 33, wherein said composition comprises an antisense, a siRNA, or shRNA molecule which recognizes and binds to a nucleic acid encoding one or more Dkk interacting proteins.

5

9. The method of any of Claims 1, 24, 28, or 33, and wherein said composition comprises a Dkk peptide aptamer.

10

10. The method of any of Claims 1, 24, 28, or 33, wherein said composition comprises a mimetic of a Dkk peptide aptamer.

11. The method of any of Claims 1, 24, 28, or 33, wherein said composition inhibits Dkk binding to LRP5, LRP6, or HBM.

15

12. The method of any of Claims 1, 24, 28, or 33, wherein said composition enhances binding of Dkk to LRP5, LRP6, or HBM.

13. The method of any of Claims 1, 24, 28, or 33, wherein said composition comprises a Dkk interacting protein peptide aptamer.

20

14. The method of any of Claims 1, 24, 28, or 33, wherein said composition comprises a mimetic of a Dkk interacting protein peptide aptamer.

25

15. The method of any of Claims 1, 24, 28 or 33, wherein said composition inhibits Dkk interacting protein or Dkk-binding fragment thereof binding to Dkk.

30

16. The method of any of Claims 1, 24, 28, or 33, wherein said composition enhances binding of Dkk interacting protein or Dkk-binding fragment thereof to Dkk.

17. The method of any of Claims 1, 24, 28, or 33, wherein said subject is a vertebrate or an invertebrate organism.

5 18. The method of any of Claims 1, 24, 28, or 33, wherein said subject is a mammal.

10 19. The method of any of Claims 1, 24, 28, or 33, wherein said subject is a canine, a feline, an ovine, a primate, an equine, a porcine, a caprine, a camelid, an avian, a bovine, or a rodent.

20. The method of Claim 19, wherein said primate is a human.

15 21. The method of any of Claims 1, 24, 28, or 33, wherein said composition comprises an LRP5 peptide aptamer.

22. The method of Claim 21, wherein said peptide aptamer is OST262 (SEQ ID NO:208).

20 23. The method of any of Claims 1, 24, 28 or 33, wherein the composition comprises an LRP5 antibody or an immunologically active fragment thereof.

25 24. A method of regulating Dkk-Wnt pathway activity in a subject comprising administering a composition which modulates Dkk activity in an amount effective to regulate Dkk-Wnt pathway activity.

25. The method of Claims 24, 101, or 107, wherein the Wnt is one or more of Wnt1-Wnt19.

26. The method of Claim 25, wherein the Wnt is Wnt1, Wnt3, Wnt3a, or Wnt10b.

5 27. The method of Claim 24 wherein said composition which modulates Dkk activity or modulates Dkk interaction with LRP5/LRP6/HBM is administered in an amount effective to modulate Wnt signaling.

10 28. A method of modulating bone mass in a subject comprising administering to the subject a composition which modulates Dkk activity or Dkk interaction with LRP5, LRP6, or HBM in an amount effective to modulate bone mass in the subject.

29. The method of Claim 28, wherein bone mass is increased.

15 30. The method of the previous claim, wherein the increase in bone mass is determined via one or more of a decrease in fracture rate, an increase in bone strength, an increase in bone density, an increase in bone mineral density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density, an increase in bone diameter, and an increase in
20 inorganic bone content.

31. The method of Claims 28 or 36, wherein said subject has a bone mass disorder selected from the group consisting of a bone development disorder, a bone fracture, age-related loss of bone, chondrodystrophy, drug-
25 induced bone disorder, high bone turnover, hypercalcemia, hyperostosis, osteogenesis imperfecta, osteomalacia, osteomyelitis, osteoporosis, Paget's disease, osteoarthritis, and rickets.

32. The method of Claim 28, wherein the composition which
30 modulates Dkk activity or Dkk interaction with LRP5, LRP6, or HBM is

administered in an amount effective to modulate the amount of trabecular and/or cortical tissue.

5 33. A method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates Dkk activity or Dkk interaction with LRP5, LRP6, or HBM in an amount effective to modulate lipid levels in the subject.

10 34. The method of Claim 33, wherein lipid levels are decreased.

 35. The method of Claim 33 or 36, wherein the subject has a lipid-modulated disorder and wherein the lipid-modulated disorder is selected from the group consisting of a cardiac condition, atherosclerosis, familial lipoprotein lipase deficiency, familial apoprotein CII deficiency, familial type 3
15 hyperlipoproteinemia, familial hypercholesterolemia, familial hypertriglyceridemia, multiple lipoprotein-type hyperlipidemia, elevated lipid levels due to dialysis and/or diabetes, and elevated lipid levels of unknown etiology.

20 36. A method of diagnosing low or high bone mass and/or high or low lipid levels in a subject comprising examining expression of Dkk, LRP5, LRP6, HBM, or and HBM-like variant in the subject and determining whether Dkk, LRP5, LRP6, HBM, or an HBM-like variant is over- or under-expressed to
25 determine whether subject has (a) high or low bone mass and/or (b) has high or low lipid levels.

 37. A method of screening for a compound which modulates the interaction of Dkk with LRP5, LRP6, HBM, or a Dkk-binding fragment of LRP5, LRP6, or HBM comprising:

(a) exposing Dkk and a LRP5, LRP6, and/or HBM binding fragment thereof to a compound; and

(b) determining whether said compound modulates Dkk interaction with the LRP5/LRP6/HBM binding fragment.

5

38. The method of Claim 37, wherein said modulation is determined by whether said compound binds to Dkk or the LRP5, LRP6, or HBM binding fragment thereof.

10

39. The method of Claim 37, wherein Dkk or a LRP-binding fragment thereof is attached to a substrate.

15

40. The method of Claim 37, wherein said compound comprises one or more compounds selected from the group consisting of Dkk interacting proteins, or a Dkk-binding fragment thereof.

41. The method of Claim 37 or 48, wherein said compound comprises a Dkk peptide aptamer.

20

42. The method of Claim 37 or 48, wherein said compound comprises a mimetic of a Dkk peptide aptamer.

43. The method of Claim 37 or 48, wherein said compound comprises a Dkk interacting protein peptide aptamer.

25

44. The method of Claim 37 or 48, wherein the compound comprises an LRP5 peptide aptamer.

30

45. The method of Claim 44, wherein the peptide aptamer is OST262 (SEQ ID NO:208).

46. The method of Claim 37 or 48, wherein the compound comprises an LRP5 antibody.

5 47. The method of Claim 37 or 48, wherein said compound is a mimetic of a Dkk interacting protein peptide aptamer.

48. A method of screening for a compound which modulates the interaction of Dkk with a Dkk interacting protein comprising:

- 10 (a) exposing a Dkk interacting protein or a Dkk-binding fragment thereof to a compound; and
- (b) determining whether said compound bound to a Dkk interacting protein or the Dkk-binding fragment thereof; and
- (c) further determining whether said compound modulates the interaction of Dkk interacting protein and Dkk.

15

49. The method of Claim 48, wherein the Dkk interacting protein or a Dkk-binding fragment thereof is attached to a substrate.

20 50. A composition comprising a LRP5, LRP6, or HBM activity-modulating compound and a pharmaceutically acceptable carrier therefor.

51. The composition of Claim 50, wherein said LRP5, LRP6, or HBM activity-modulating compound comprises a compound which binds to Dkk thereby modulating the interaction of Dkk with LRP5, LRP6, or HBM.

25

52. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises one or more Dkk interacting proteins and Dkk-binding fragments thereof.

53. The composition of Claim 50, wherein said LRP5, or LRP6, or HBM modulating compound is a monoclonal antibody or an immunologically active fragment thereof which binds to a Dkk interacting protein, or a Dkk-binding fragment thereof.

5

54. The composition of Claim 53, wherein the monoclonal antibody is human, chimeric, humanized, primatized®, or bispecific.

55. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises an antisense, a siRNA, or shRNA molecule which recognizes and binds to a nucleic acid encoding one or more Dkk interacting proteins.

10

56. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises a Dkk peptide aptamer.

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57. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises a mimetic of a Dkk peptide aptamer.

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58. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises a Dkk interacting protein peptide aptamer.

59. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises a mimetic of a Dkk interacting protein peptide aptamer.

25

60. The composition of Claim 50, wherein the compound comprises an LRP5 peptide aptamer.

61. The composition of Claim 60, wherein the peptide aptamer is OST262.

5 62. The composition of Claim 50, wherein the compound comprises an LRP5 antibody.

63. A pharmaceutical composition comprising a compound which modulates Dkk activity and a pharmaceutically acceptable carrier therefor.

10 64. A method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:

- (a) creating an LRP5, LRP6, or HBM fluorescent fusion protein using a first fluorescent tag; and
- (b) creating a Dkk fusion protein comprising a second fluorescent tag;
- 15 (c) adding a test compound; and
- (d) assessing changes in the ratio of fluorescent tag emissions using Fluorescence Resonance Energy Transfer (FRET) or Bioluminescence Resonance Energy Transfer (BRET) to determine whether the compound modulates Dkk and LRP5/LRP6/HBM interactions.

20

65. A method of identifying binding partners for a Dkk protein comprising the steps of:

- (a) exposing the Dkk protein(s) or a LRP5/LRP6 binding fragment thereof to a potential binding partner; and
- 25 (b) determining if the potential binding partner binds to a Dkk protein or the LRP5/LRP6 binding fragment thereof.

66. A nucleic acid encoding a Dkk interacting protein peptide aptamer comprising a nucleic acid encoding a scaffold protein in-frame with the activation

domain of Gal4 or LexA that is in-frame with a nucleic acid that encodes a Dkk interacting protein amino acid sequence.

- 5
67. A vector comprising the nucleic acid of Claim 66.
68. The nucleic acid of Claim 66, wherein the scaffold protein is trxA.
- 10
69. A method of detecting a modulatory activity of a compound on the binding interaction of a first peptide and a second peptide of a peptide binding pair that bind through extracellular interaction in their natural environment, comprising:
- 15
- (i) culturing at least one eukaryotic cell comprising:
- a) a nucleotide sequence encoding a first heterologous fusion protein comprising the first peptide or a segment thereof joined to a transcriptional activation protein DNA binding domain;
- 20
- b) a nucleotide sequence encoding a second heterologous fusion protein comprising the second peptide or a segment thereof joined to a transcriptional activation protein transcriptional activation domain;
- 25
- wherein binding of the first peptide or segment thereof and the second peptide or segment thereof reconstitutes a transcriptional activation protein; and
- c) a reporter element activated under positive transcriptional control of the reconstituted transcriptional activation protein, wherein expression of the reporter element produces a selected phenotype;
- (ii) incubating the eukaryotic cell in the presence of a compound under conditions suitable to detect the selected phenotype; and

- (iii) detecting the ability of the compound to affect the binding interaction of the peptide binding pair by determining whether the compound affects the expression of the reporter element which produces the selected phenotype;

5 wherein (1) said first peptide is a Dkk peptide and the second peptide is a peptide selected from LRP5, HBM, LRP6 and the Dkk-binding portion of LRP5/LRP6/HBM or (2) said first peptide is a Dkk interacting protein or the Dkk-binding fragment thereof and said second peptide is a Dkk peptide.

10 70. The method of Claim 69, wherein the eukaryotic cell is a yeast cell.

71. The method of Claim 70, wherein the yeast cell is *Saccharomyces*.

15 72. The method of Claim 71, wherein the *Saccharomyces* cell is *Saccharomyces cerevisiae*.

20 73. The method of Claim 69, wherein the Dkk is Dkk-1 and wherein the compound comprises one or more Dkk interacting proteins or a Dkk-binding fragment thereof.

74. The method of Claim 73, wherein the compound is directly added to assay.

25 75. The method of Claim 73, wherein the compound is recombinantly expressed by said eukaryotic cell in addition to said first and second peptides.

30 76. The method of Claim 69, wherein the compound comprises a Dkk peptide aptamer.

77. The method of Claim 69, wherein the compound comprises a mimetic of a Dkk peptide aptamer.

78. The method of Claim 69, wherein the compound comprises a Dkk interacting protein peptide aptamer.

79. The method of Claim 69, wherein the compound comprises a mimetic of a Dkk interacting protein peptide aptamer.

80. The method of Claim 69, wherein the eukaryotic cell further comprises at least one endogenous nucleotide sequence selected from the group consisting of a nucleotide sequence encoding the DNA binding domain of a transcriptional activation protein, a nucleotide sequence encoding the transcriptional activation domain of a transcriptional activation protein, and a nucleotide sequence encoding the reporter element, wherein at least one of the endogenous nucleotide sequences is inactivated by mutation or deletion.

81. The method of Claim 69, wherein the peptide binding pair comprises a ligand and a receptor to which the ligand binds.

82. The method of Claim 69, wherein the transcriptional activation protein is Gal4, Gcn4, Hap1, Adr1, Swi5, Ste12, Mcm1, Yap1, Ace1, Ppr1, Arg81, Lac9, Qa1F, VP16, or a mammalian nuclear receptor.

83. The method of Claim 69, wherein at least one of the heterologous fusion proteins is expressed from an autonomously-replicating plasmid.

84. The method of Claim 69, wherein the DNA binding domain is a heterologous DNA-binding domain of a transcriptional activation protein.

85. The method of Claim 84, wherein the DNA binding protein is selected from the group consisting of a mammalian steroid receptor and bacterial LexA protein.

5 86. The method of Claim 69, wherein the reporter element is selected from the group consisting of *lacZ*, a polynucleotide encoding luciferase, a polynucleotide encoding green fluorescent protein (GFP), and a polynucleotide encoding chloramphenicol acetyltransferase.

10 87. The method of Claim 86, wherein the reporter element is LacZ.

88. The method of Claim 69, wherein the test sample comprises an LRP5 peptide aptamer.

15 89. The method of Claim 88, wherein the peptide aptamer is OST262 (SEQ ID NO:208).

90. The method of Claim 69, wherein the test sample comprises an LRP5 antibody.

20 91. A transgenic animal wherein Dkk-1 is knocked out in a tissue-specific fashion.

92. The transgenic animal of Claim 91, wherein the tissue specificity is
25 bone tissue, cancer tissue, or liver tissue.

93. A method for identifying potential compounds which modulate Dkk activity comprising:

30 a) measuring the effect on binding of one or more Dkk interacting proteins, or a Dkk-binding fragment thereof, with Dkk or a

fragment thereof in the presence and absence of a compound;
and

b) identifying as a potential Dkk modulatory compound a
compound which modulates the binding between one or more Dkk
interacting proteins or Dkk-binding fragment thereof and Dkk or
fragment thereof.

94. A peptide aptamer of Figure 3 (SEQ ID NOs:171-188) or Figure 4
(SEQ ID NOs:189-192).

95. An antibody or antibody fragment which recognizes and binds to
one or more peptides of amino acid sequences GNKYQTIDNYQPYPYPC (SEQ ID
NO:118), LDGYSRRTLSSKMYHTKGQEG (SEQ ID NO:119),
RIQKDDHHQASNSSRLHTCQRH (SEQ ID NO:120), RGEIETITESFGND (SEQ
ID NO:121), EIFQRCYCGEGLSCRIQKD (SEQ ID NO:122),
MYWTDWVETPRIE (SEQ ID NO:123), MYWTDWGETPRIE (SEQ ID NO:124),
KRTGGKRKEILSA (SEQ ID NO:125), ERVEKTTGDKRTRIQGR (SEQ ID
NO:126), KQQCDSFPDCIDGSDE (SEQ ID NO:127), or a Dkk-1 amino acid
sequence selected from the group consisting Asn34-His266 (SEQ ID NO:110),
Asn34-Cys245 (SEQ ID NO:111), Asn34-Lys182 (SEQ ID NO:112), Cys97-
His266 (SEQ ID NO:113), Val139-His266 (SEQ ID NO:114), Gly183-His266
(SEQ ID NO:115), Cys97-Cys245 (SEQ ID NO:116), or Val139-Cys245 (SEQ ID
NO:117).

96. The antibody or antibody fragment of Claim 95, wherein the
antibody is a monoclonal antibody.

97. The antibody or antibody fragment of Claim 95, wherein the
antibody is a polyclonal antibody

98. A method of identifying Dkk interacting proteins which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

(a) injecting Dkk and potential Dkk interacting protein mRNA into a *Xenopus* blastomere; and

5 (b) assessing axis duplication or analyzing marker gene expression; and

(c) identifying compositions which elicit changes in axis duplication or marker gene expression as Dkk interacting proteins which modulate the interaction of Dkk with the Wnt signaling pathway.

10

99. The method of Claim 98, wherein the mRNA of HBM, LRP5/6, any Wnt, Wnt antagonist, Wnt pathway modulator, or combination of these is co-injected into the *Xenopus* blastomere.

15

100. The method of Claim 98, wherein the marker gene analyzed is Siamese, Xnr3, slug, Xbra, HNK-1, endodermin, Xlhx8, BMP2, BMP4, XLRP6, EF-1, or ODC.

20

101. A method for identifying Dkk interacting proteins which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

(a) transfecting cells with constructs containing Dkk and potential Dkk interacting proteins; and

(b) assessing changes in expression of a reporter gene linked to a Wnt-responsive promoter; and

25

(c) identifying as a Dkk interacting protein any protein which alters reporter gene expression compared with cells transfected with a Dkk construct alone.

30

102. The method of Claim 101, wherein the cells are HOB-03-CE6, HEK293, or U2OS cells.

103. The method of Claim 101, wherein the Wnt-responsive promoter is TCF or LEF.

5 104. The method of Claim 101, wherein the cells are co-transfected with CMV β -galactosidase.

105. A method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:

- 10 (a) immobilizing LRP5/LRP6/HBM to a solid surface; and
- (b) treating the solid surface with a secreted Dkk protein or a secreted epitope-tagged Dkk and a test compound; and
- (c) determining whether the compound regulates binding between Dkk and LRP5/LRP6/HBM using antibodies to Dkk or the epitope tag or by directly measuring activity of an epitope tag.

15

106. The method of Claim 105, wherein the epitope tag is alkaline phosphatase, histidine, or a V5 tag.

20 107. A method for identifying compounds which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

- (a) transfecting cells with constructs containing Dkk and Wnt proteins;
 - (b) assessing changes in expression of a reporter element linked to a Wnt- responsive promoter; and
 - (c) identifying as a Dkk/Wnt interaction modulating compound any
- 25 compound which alters reporter gene expression compared with cells transfected with a Dkk construct alone.

108. The method according to Claim 107, wherein Wnt3a and Wnt1 constructs are co-transfected into the cells.

30

109. The method according to Claim 107, wherein the cells are U2-OS, HOB-03-CE6, or HEK293 cells.

5 110. The method according to Claim 107, wherein the reporter element used is TCF-luciferase, tk-Renilla, or a combination thereof.

111. A method of testing compounds that modulate Dkk-mediated activity in a mammal comprising

- 10 (a) providing a group of transgenic animals having (1) a regulatable one or more Dkk genes, (2) a knock-out of Dkk genes, or (3) a knock-in of one or more Dkk genes;
- (b) providing a second group of control animals respectively for the group of transgenic animals in step (a); and
- 15 (c) exposing the transgenic animal group and control animal group to a potential Dkk-modulating compound which modulates bone mass or lipid levels; and
- (d) comparing the transgenic animals and the control group of animals and determining the effect of the compound on bone mass or lipid levels in the transgenic animals as compared to the control animals.

20

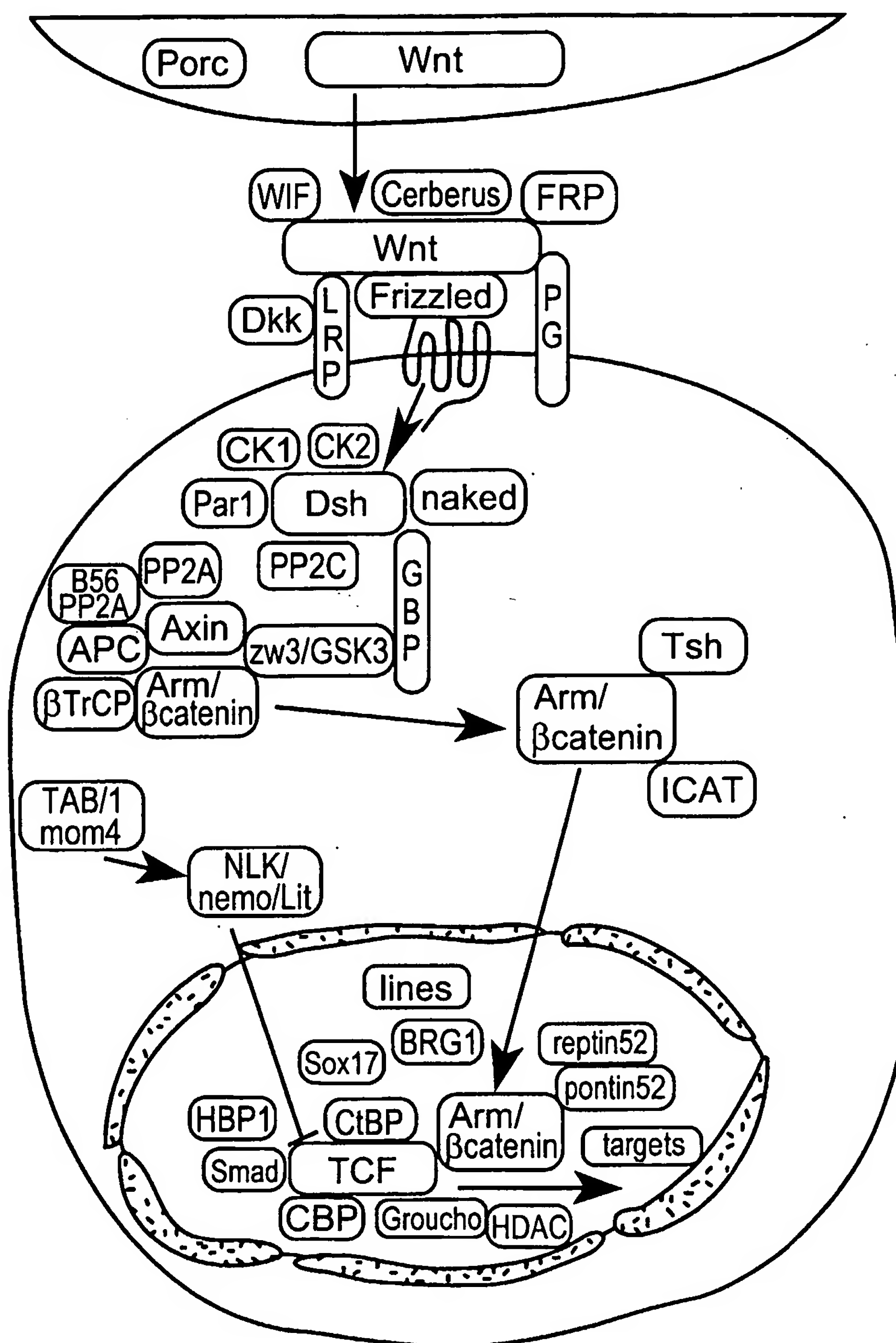
112. A method of screening for compounds or compositions which modulate the interaction of Dkk and a Dkk interacting protein comprising:

- (a) exposing a Dkk interacting proteins or a Dkk-binding fragment thereof to a compound; and
- 25 (b) determining whether said compound binds to a Dkk interacting proteins or the Dkk-binding fragment thereof.

113. The method of Claim 112, wherein said modulation is determined by whether said compound binds to the Dkk interacting protein or the Dkk-binding fragment thereof.

5 114. An antibody or antibody fragment which recognizes and binds to a sequence depicted in Figure 3 (SEQ ID NOs:171-188) or Figure 4 (SEQ ID NOs:189-192).

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Model of Wnt signaling

FIG. 1

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Sequence of baits used in Y2H screens

>DKK1 (SEQ ID NO: 168)

AATTCCAACGCTATCAAGAACCTGCCCCCACCCTGGGGCGGGCGCTG
CGGGGCACCCAGGCTCTGCAGTCAGCGCCGCGCCGGGAATCCTGTA
CCCGGGCGGGGAATAAGTACCAGACCATTGACAACCTACCAGCCGTAC
CCGTGCGCAGAGGACGAGGAGTGCGGGCACTGATGAGTACTGCGCT
AGTCCCACCCGCGGAGGGGACGCGGGGCGTGCAAATCTGTCTCGCCT
GCAGGAAGCGCCGAAAACGCTGCATGCGTCACGCTATGTGCTGCCC
CGGGAATTACTGCAAAAATGGAATATGTGTGTCTTCTGATCAAAAT
CATTTCCGAGGAGAAATTGAGGAAACCATCACTGAAAGCTTTGGTA
ATGATCATAGCACCTTGGATGGGTATTCCAGAAGAACCACCTTGTC
TTCAAAAATGTATCACACCAAAGGACAAGAAGGTTCTGTTTGTCTC
CGGTCATCAGACTGTGCCTCAGGATTGTGTTGTGCTAGACACTTCTG
GTCCAAGATCTGTAAACCTGTCCTGAAAGAAGGTCAAGTGTGTACC
AAGCATAGGAGAAAAGGCTCTCATGGACTAGAAATATTCCAGCGTT
GTTACTGTGGAGAAGGTCTGTCTTGCCGGATACAGAAAGATCACCA
TCAAGCCAGTAATTCTTCTAGGCTTCACACTTGTCAGAGACACTAA

FIG. 2A

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>zmax1 LBD1 (SEQ ID NO: 169)

CTCATCCTGCCCCTGCATGGACTGAGGAACGTCAAAGCCATCGACTAT
GACCCACTGGACAAGTTCATCTACTGGGTGGATGGGGCGCCAGAACATC
AAGCGAGCCAAGGACGACGGGACCCAGCCCTTTGTTTTGACCTCTCTG
AGCCAAGGCCAAAACCCAGACAGGCAGCCCCACGACCTCAGCATCGA
CATCTACAGCCGGACACTGTTCTGGACGTGCGAGGCCACCAATACCAT
CAACGTCCACAGGCTGAGCGGGGAAGCCATGGGGGTGGTGCTGCGTG
GGGACCGCGACAAGCCCAGGGCCATCGTCGTCAACGCGGAGCGAGGG
TACCTGTACTTCACCAACATGCAGGACCGGGCAGCCAAGATCGAACGC
GCAGCCCTGGACGGCACCGAGCGCGAGGTCCTCTTCACCACCGGCCTC
ATCCGCCCTGTGGCCCTGGTGGTAGACAACACACTGGGGCAAGCTGTTC
TGGGTGGACGCGGACCTGAAGCGCATTGAGAGCTGTGACCTGTCAGG
GGCCAACCGCCTGACCCTGGAGGACGCCAACATCGTGCAGCCTCTGGG
CCTGACCATCCTTGGCAAGCATCTCTACTGGATCGACCGCCAGCAGCA
GATGATCGAGCGTGTGGAGAAGACCACCGGGGACAAGCGGACTCGCA
TCCAGGGCCGTGTCGCCCACCTCACTGGCATCCATGCAGTGGAGGAAG
TCAGCCTGGAGGAGTTCTCAGCCCACCCATGTGCCCGTGACAATGGTG
GCTGCTCCACATCTGTATTGCCAAGGGTGATGGGACACCACGGTGCT
CATGCCCAGTCCACCTCGTGCTCCTGCAGAACCTGCTGACCTGTGGAG
AGCCGCCCACCTGCTCCCCGGACCAAGTTTGCATGTGCCACAGGGGAGA
TCGACTGTATCCCCGGGGCCTGGCGCTGTGACGGCTTTCCCGAGTGCG
ATGACCAGAGCGACGAGGAGGGCTGCCCCGTGTGCTCCGCCGCCAGT
TCCCCTGCGCGCGGGGTCAAGTGTGTGGACCTGCGCCTGCGCTGCGACG
GCGAGGCAGACTGTCAGGACCGCTCAGACGAGGCGGACTGTGACGCC
ATCTGCCTGCCCAACCAGTTCCGGTGTGCGAGCGGCCAGTGTGTCCTC
ATCAAACAGCAGTGCGACTCCTTCCCCGACTGTATCGACGGCTCCGAC
GAGCTCATGTGTGAAATCACCAAGCCGCCC

FIG. 2B

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>zmax1 LBD4 (SEQ ID NO: 170)

AGGGCCATCGTCGTCAACGCGGAGCGAGGGTACCTGTACTTCACCAA
CATGCAGGACCGGGCAGCCAAGATCGAACGCGCAGCCCTGGACGGCA
CCGAGCGCGAGGTCCTCTTCACCACCGGCCTCATCCGCCCTGTGGCCC
TGGTGGTAGACAACACACTGGGCAAGCTGTTCTGGGTGGACGCGGAC
CTGAAGCGCATTGAGAGCTGTGACCTGTCAGGGGGCCAACCGCCTGAC
CCTGGAGGACGCCAACATCGTGCAGCCTCTGGGGCCTGACCATCCTTGG
CAAGCATCTCTACTGGATCGACCGCCAGCAGCAGATGATCGAGCGTG
TGGAGAAGACCACCGGGGACAAGCGGACTCGCATCCAGGGGCCGTGTC
GCCCACCTCACTGGCATCCATGCAGTGGAGGAAGTCAGCCTGGAGGA
GTTCTCAGCCCACCCATGTGCCCCGTGACAATGGTGGCTGCTCCCACAT
CTGTATTGCCAAGGGTGATGGGACACCACGGTGCTCATGCCCAGTCCA
CCTCGTGCTCCTGCAGAACCTGCTGACCTGTGGAGAGCCGCCACCTG
CTCCCCGGACCAGTTTGCATGTGCCACAGGGGAGATCGACTGTATCCC
CGGGGCCTGGCGCTGTGACGGCTTTCCCGAGTGCGATGACCAGAGCG
ACGAGGAGGGCTGCCCCGTGTGCTCCGCCGCCAGTTCCCCTGCGCGC
GGGGTCAGTGTGTGGACCTGCGCCTGCGCTGCGACGGCGAGGCAGAC
TGTCAGGACCGCTCAGACGAGGCGGACTGTGACGCCATCTGCCTGCC
CAACCAGTTCCGGTGTGCGAGCGGCCAGTGTGTCCTCATCAACAGC
AGTGCGACTCCTTCCCCGACTGTATCGACGGCTCCGACGAGCTCATGT
GTGAAATCACCAAGCCGCCCTAAGCGGCCGC

FIG. 2C

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Screen of DKK1 X
Peptide Library

name	motif	# hits	SEQ ID NO:
252-1	SVGCLLCAGLGWWSLS	3	171
252-2	WCCCGLFRGVCVWSCGAD D	2	172
252-3	GWRRCDWCGCVSWCWV	1	173
252-4	MPGSVSHCWGGICEAL	8	174
252-15	SCCAVDVCLRCGGWFR	1	175
252-16	SVLGTCCCCGGWILCE	2	176
252-17	VLSVCEVCGGVFVRRRC	1	177
252-18	GMWYWWSGRDCALCWL	1	178
252-19	CTAVMWGVGSAVAYLGE	1	179
252-20	WCWWCGCRGVVWR	1	180
252-21	CVCASFCCCVCGLRLL	1	181
252-23	TYEVCEECEGGRVRMWV	6	182
252-25	VVVCASCGQVWHGSGA	2	183
252-26	CCRCCHCWDCEWHMCV	1	184
252-27	FCASCCWCGCDCFGWV	2	185
252-32	CDYCWSCGVWCPSSWL	3	186
252-47	VYLCVWCGAARFGCYG	1	187
252-48	FCVCGCCWCWCAACWC	1	188

FIG. 3

peptide #	peptide seq	# hits	SEQ ID NO:
9	VVLCSRCGRLWRWSCG	1	189
12	EVRQVTCIRCRRGFL	1	190
13	GGGGMWEAWSCYACG	1	191
14	GWRWCGRCGALWRRV	3	192

FIG. 4

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Gene	Genbank Accession #	Protein Accession #
granulin	M75161	AAA58617
similar to cys/His rich protein	BC004544	AAH04544
IGF-BINDING PROTEIN 6	M69054	AAA88070
latent TGFb binding protein 4	AF051344	AAC39879
NOTCH 2	AF315356	AAG37073
fibulin 1	X53743	CAA37772
MDC15 (ADAM15)	U46005	AAC51112
DKFZp761G02121(notch1 Ca++ binding like)	AL137311	CAB70690
chordin	AF076612	AAC69835
fibronectin 1	U42594	AAD00019
MG50(melanoma associated antigen)	AF200348	AAF06354
unknown (notch 4-like)	AX068260	CAC27245
Slit 1	AB017167	BAA35184
tomoregulin (agarin repeat homology)	AB004064	BAA90820
sprouty 1	AF041037	AAC39566
sprouty 2	AF039843	AAC04258
NOV1	X96584	CAA65403
agrin	AF016903	AAC39776
fibrillin 1	L13923	AAB02036
thrombospondin1	X04665	CAA28370
ADAM19	AF134707	AAF22162
Nafl alpha	AJ011895	CAA09855
laminin alpha 5	Z95636	CAB09137
CRIM1	AF167706	AAF34409
nidogen	M30269	AAA59932
fibulin-2	X82494	CAA57876
thrombospondin 2	L12350	AAA03703
KIAA1323	AB037744	BAA92561
fibrillin-2	U03272	AAA18950
MEGF9	AB011542	BAA32470
integrin beta 1	X07979	CAA30790
matrilin-2 precursor	U69263	AAC51260
tenascin	X56160	A32160

FIG. 5

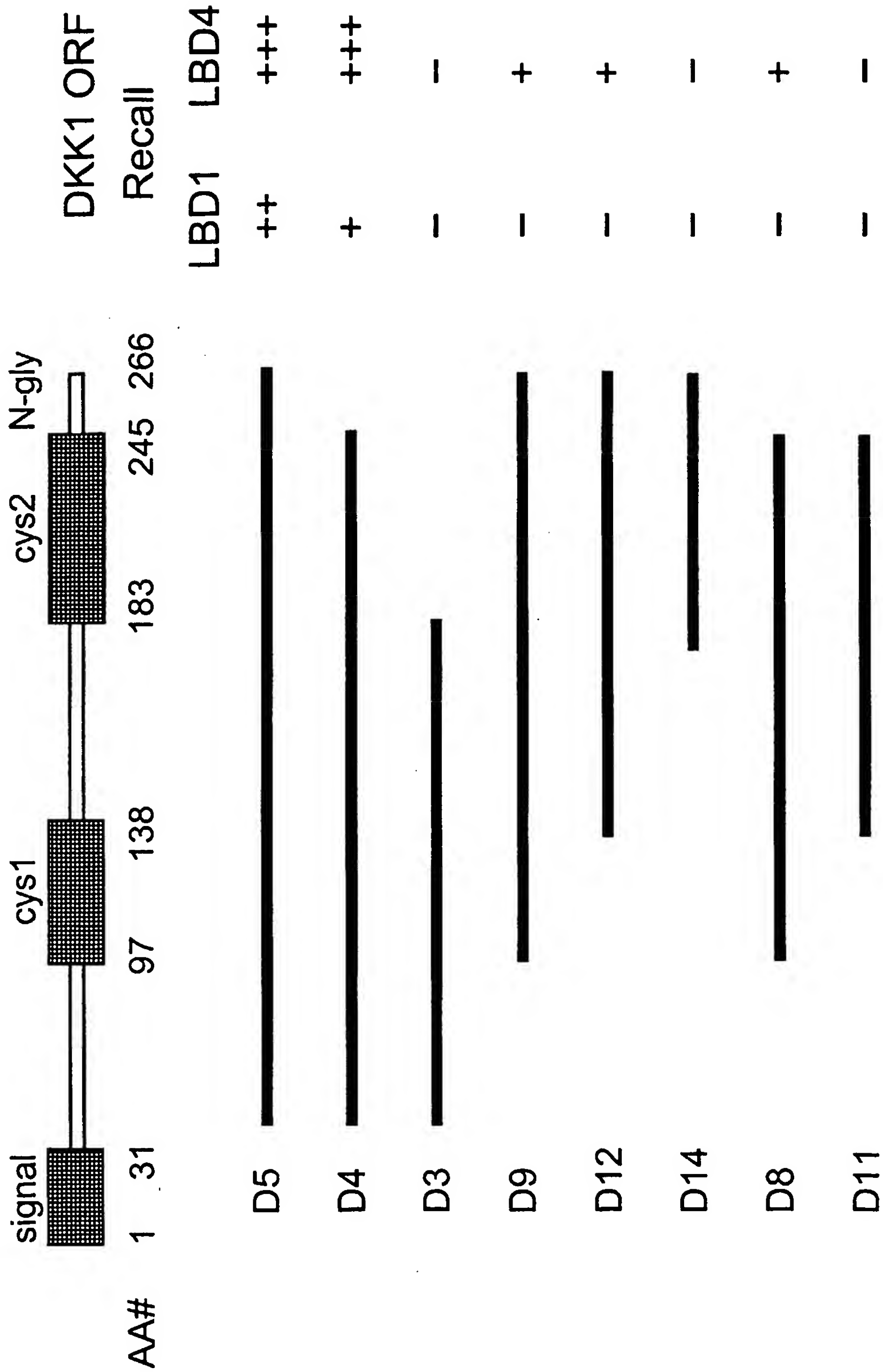
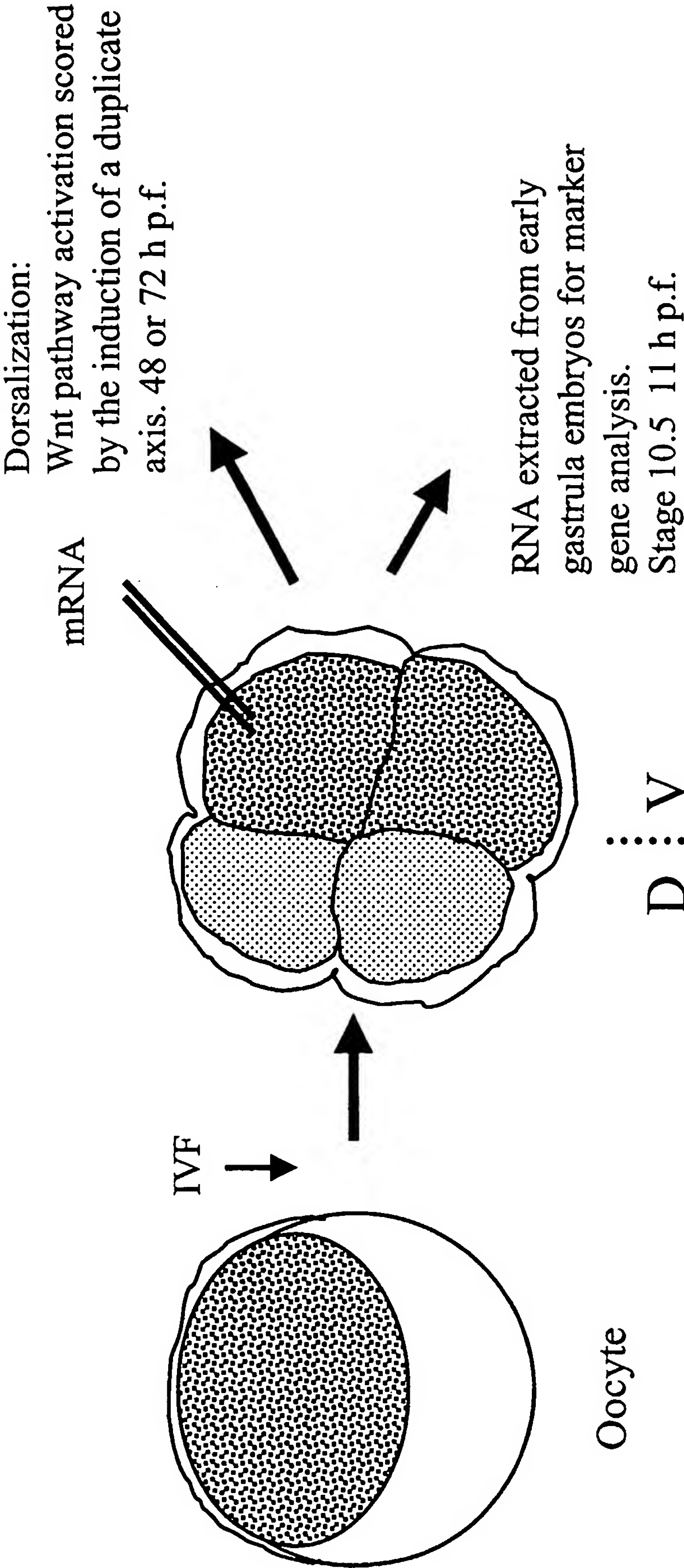


FIG. 6

Xenopus Embryo Assay for Wnt Activity



RNA Injection into
ventral blastomere
Stage 4 2 h p.f.

FIG. 7

In the Presence of Wnt5a, HBM1 is More Potent than Zmax as a Stimulator of Wnt Signaling

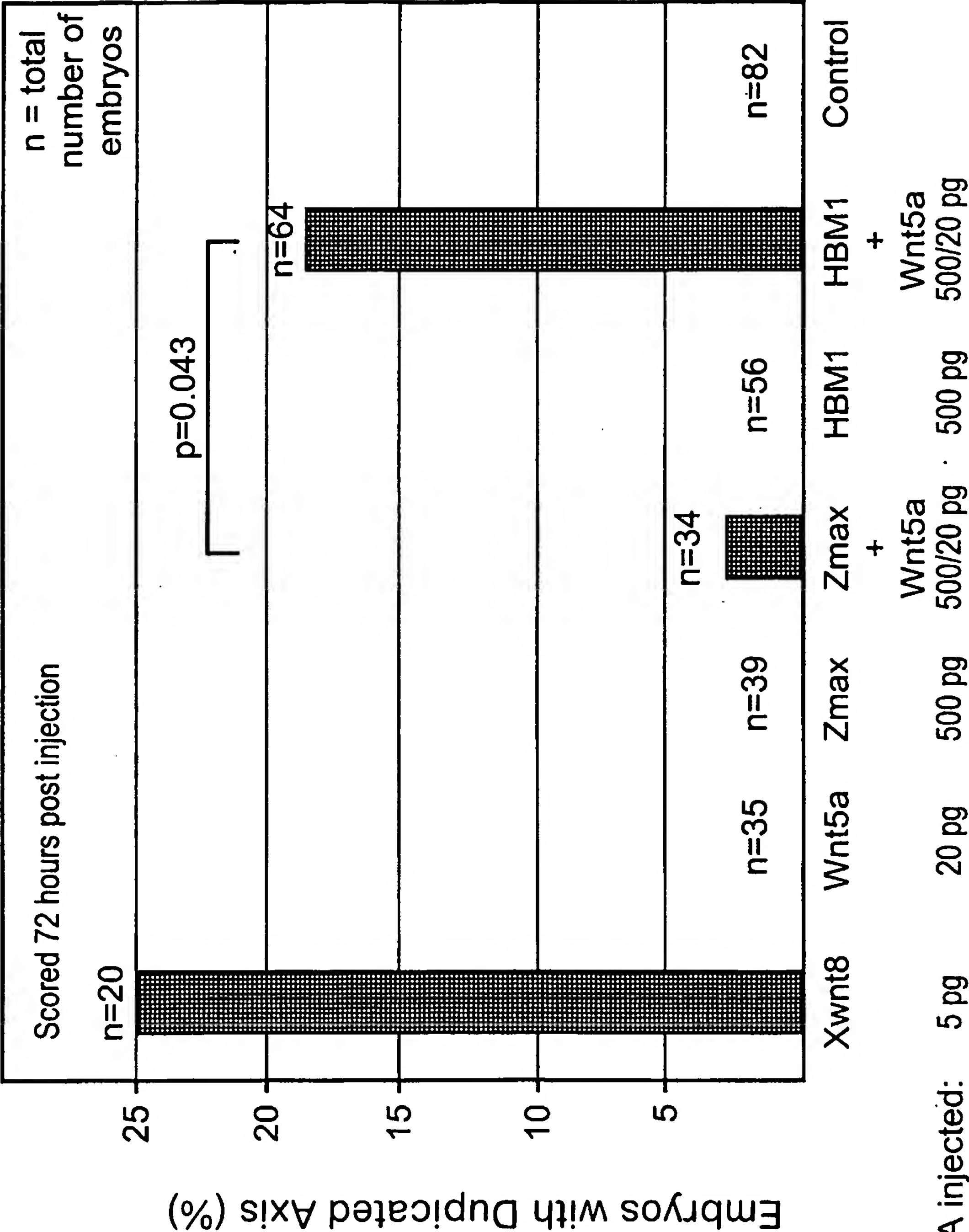


FIG. 8

Both Zmax and HBM1, in the presence of Wnt5a, induce secondary axis formation in Xenopus (photos at 48 hrs post-injection)

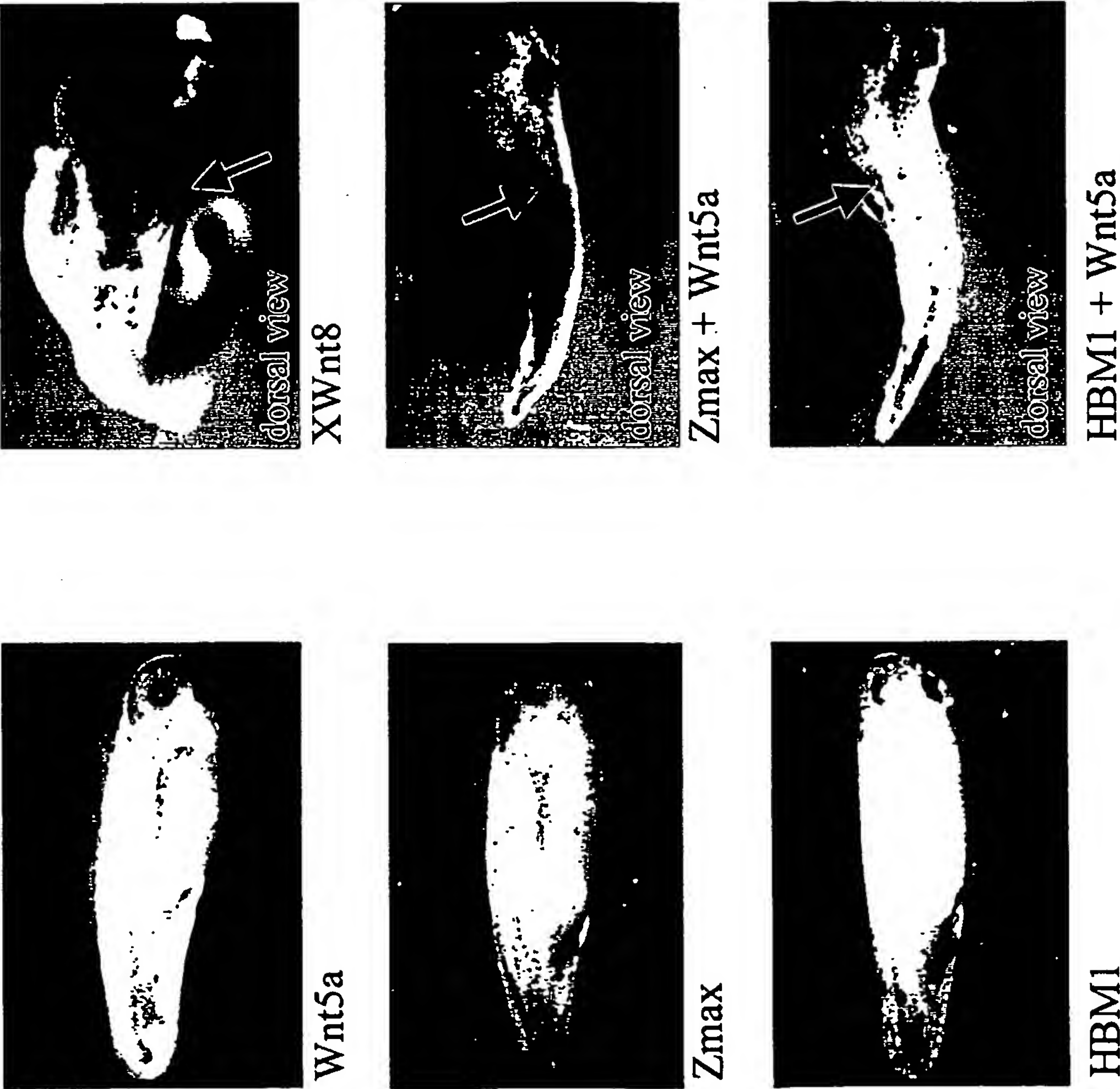


FIG. 9

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Human Dkk-1 Represses the Canonical Wnt Pathway

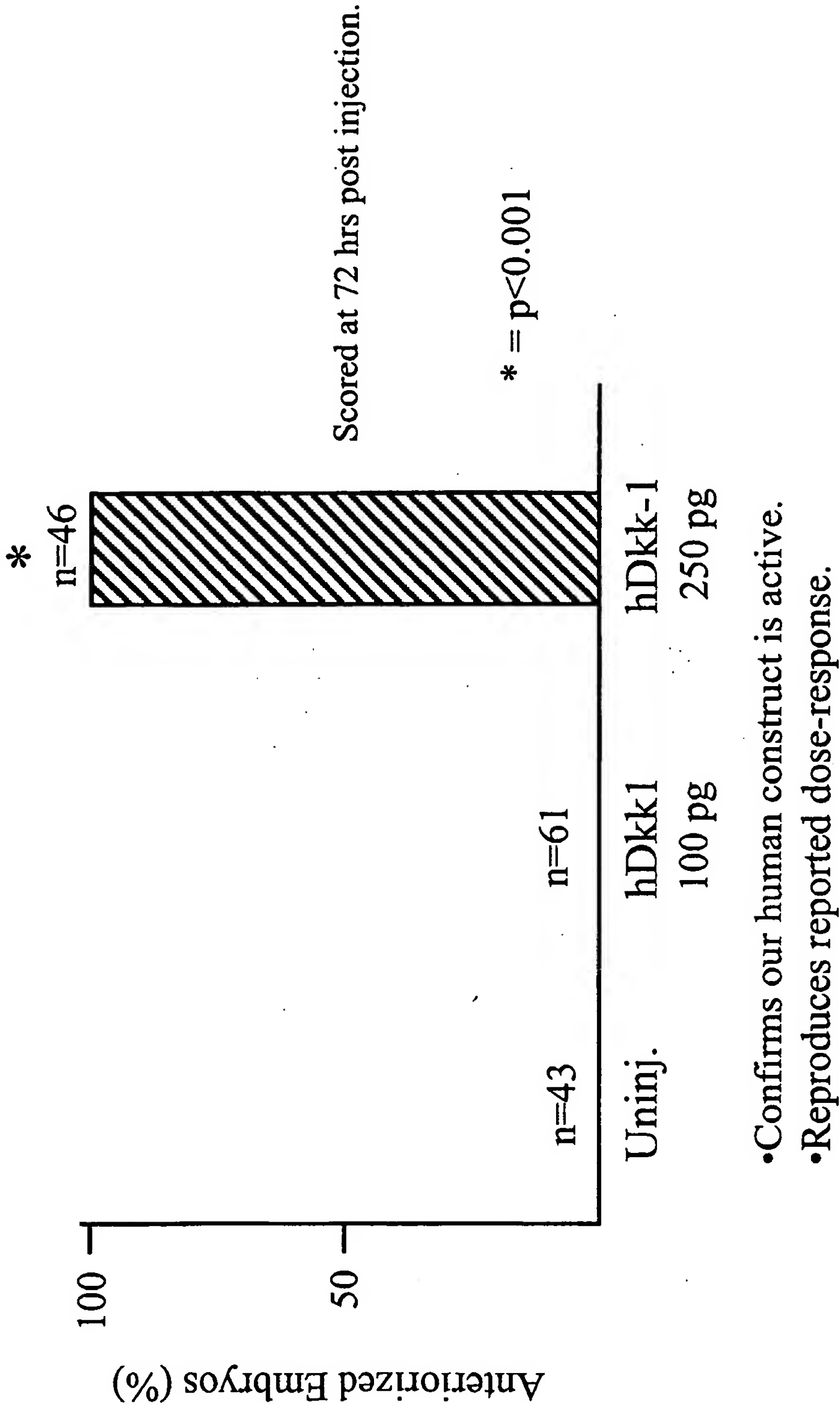


FIG. 10

hDkk1 Represses Zmax- but Not HBM1-Mediated Wnt Signaling

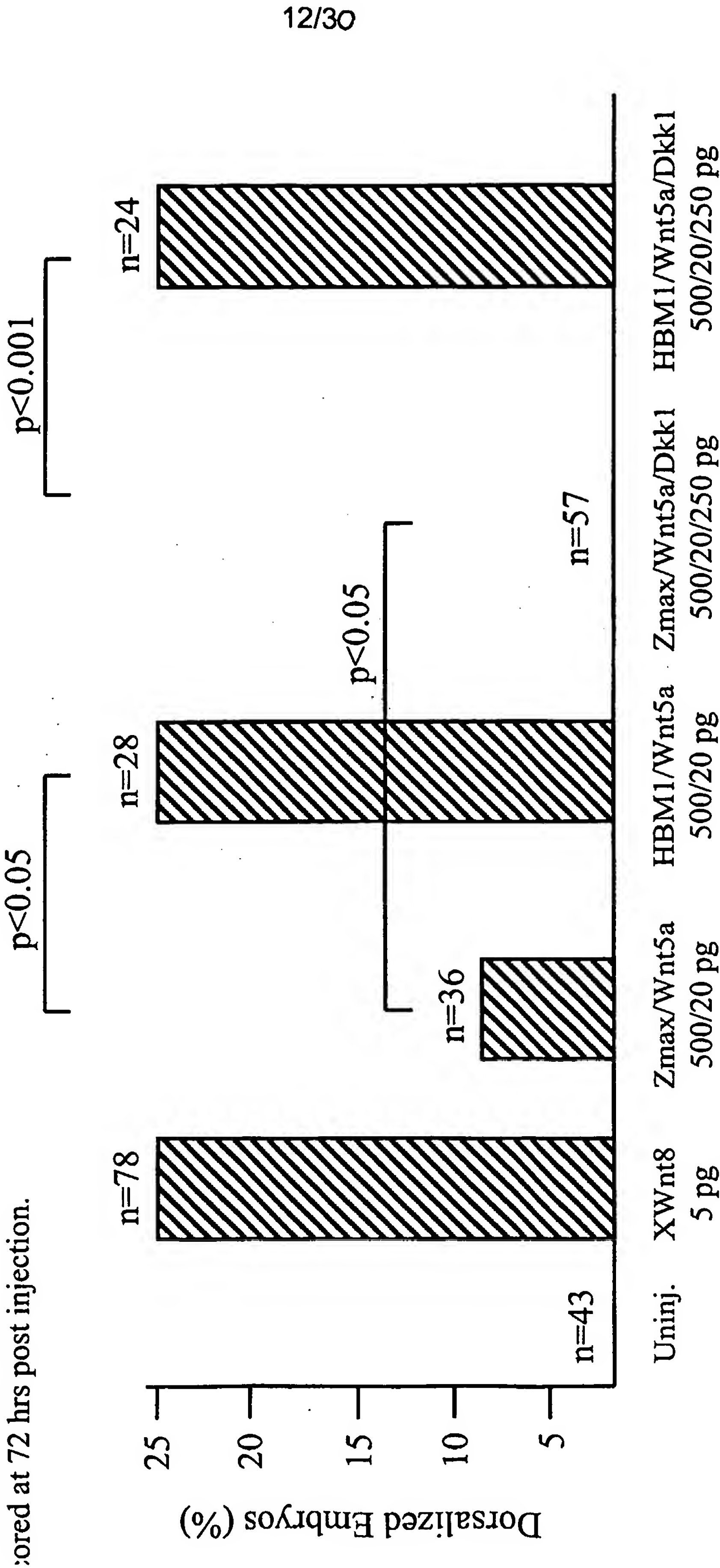


FIG. 11

Listed are the pCDNA3.1 construct names followed by the DNA sequence
OST258 (control for OST 259-OST262 and OST264, OST265)

AAGCTTGCCACCATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCCAGGTTCCACTGGTGACGG
ATCC

OST259. (SEQ ID NO: 193)

AAGCTTGCCACCATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCCAGGTTCCACTGGTGACGG
ATCCATGAGCGATAAAATTATTCACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGGGGCGATCC
TCGTGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCCGTGGTTCTGTGTTCGCGTTGTGGGCGTTTGTGGCGGTGG
TCGTGTGGGACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAGGGCAAAC
GACCGTTGCAAACTGAACATCGATCAAAACCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCCGACTCTGC
TGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGTTCCTCGAC
GCTAACCTGGCGTAAGCGGCCGC

OST260 (SEQ ID NO: 194)

AAGCTTGCCACCATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCAGGTTCCACTGGTGACGG
ATCCATGAGCGATAAAATTATTACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGGGGCGATCC
TCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCGCGGTGGCGGTGGTGTGGTTCGGTGTGGGGCTTTGTGGTGG
CGGCGFTTACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAGGGCAAAC
GACCGTTGCAAACTGAACATCGATCAAAACCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCCGACTCTGC
TGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGTTCCTCGAC
GCTAACCTGGCGTAAGCGGCCGC

OST261 (SEQ ID NO: 195)

[illegible]

OST262 (SEQ ID NO: 196)

AAGCTTGCCACCATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCAGGTTCCACTGGTGACGG
ATCCATGAGCGATAAAATTATTACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGGGGCGATCC
TCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCGGGTGGTGGGGGGATGATTGAGGCTTGGAGTTGTTAT
GCGTGTGGGACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAGGGCAAAC
GACCGTTGCAAACTGAACATCGATCAAAACCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCCGACTCTGC
TGCTGTTCAAAAACGGTGAAGTGGGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAAGTTGAAAGAGTTCCTCGAC
GCTAACCTGGCGTAAGCGGCCGC

OST263 (SEQ ID NO: 197)

AAGCTTGCCACCATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCAGGTTCCACTGGTGACGG
ATCCATGAGCGATAAAATTATTACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGGGGCGATCC
TCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCCTTGTGGATTGGGCCGGGTGATCAGGGTCTGTTTCGGCGT
TTTGTTTTACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAGGGCAAAC
GACCGTTGCAAACTGAACATCGATCAAAACCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCCGACTCTGC
TGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAGAGTTCCTCGAC
GCTAACCTGGCGTAAGCGGCGCGC

FIGURE 12A

OST264 (SEQ ID NO: 198)

AAGCTTGGCACCACATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCAGGTTCCACTGGTGACGG
ATCCGTGTCTTCTGATCAAAATCATTTCCGAGGAGAAATTGAGGAAACCATCACTGAAAGCTTTGGTAATGATCATA
GCACCTTGGATGGGTATTCCAGAAGAACCACCTTGTCTTCAAAAATGTATCACACCAAAGGACAAGAAGGTTCTGTT
TGTCTCCGGTCATCAGACTGTGCCTCAGGATTGTGTTGTGCTAGACACTTCTGGTCCAAGATCTGTAAACCTGTCCT
GAAAGAAGGTCAAGTGTGTACCAAGCATAGGAGAAAAGGCTCTCATGGACTAGAAATATTCCAGCGTTGTTACTGTG
GAGAAGGTCTGTCTTGCCGGATACAGAAAGATCACCATCAAGCCAGTAATTCTTAGGCTTCACACTTGTCAGAGA
CACTAAGCGGCEGC

OST265 (SEQ ID NO: 199)

AAGCTTGGCACCACATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCAGGTTCCACTGGTGACGG
ATCCTGCGCTAGTCCCACCCGCGGAGGGGACGCGGGCGTGCAATCTGTCTCGCCTGCAGGAAGCGCCGAAAACGCT
GCATGCGTCACGCTATGTGCTGCCCGGGAATTACTGCAAAAATGGAATATGTGTCTTCTGATCAAAATCATTTT
CGAGGAGAAATTGAGGAAACCATCACTGAAAGCTTTGGTAATGATCATAGCACCTTGGATGGGTATTCCAGAAGAAC
CACCTTGTCTTCAAAAATGTATCACACCAAAGGACAAGAAGGTTCTGTTTGTCTCCGGTCATCAGACTGTGCCTCAG
GATTGTGTTGTGCTAGACACTTCTGGTCCAAGATCTGTAAACCTGTCTGAAAGAAGGTCAAGTGTGTACCAAGCAT
AGGAGAAAAGGCTCTCATGGACTAGAAATATTCCAGCGTTGTTACTGTGGAGAAGGTCTGTCTTGCTAAGCGGCCGC

OST266 (SEQ ID NO: 200)

AAGCTTGGCACCACATGGGCGATAAAATTATTCACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGG
GGCGATCCTCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCCTATGCGTGTTGTTTTCTTGTAGTAGGTGTA
GGTGGTGGTTGCCTTGGACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAG
GGCAAACCTGACCGTTGCAAACTGAACATCGATCAAACCCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCC
GACTCTGCTGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGT
TCCTCGACGCTAACCTGGCGTAAGCGGCCGC

OST267 (SEQ ID NO: 201)

AAGCTTGGCACCACATGGGCGATAAAATTATTCACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGG
GGCGATCCTCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCCTATGCGTGTTGTTTTCTTGTAGTAGGTGTA
ATCCTTGGTCTTGGGTGACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAG
GGCAAACCTGACCGTTGCAAACTGAACATCGATCAAACCCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCC
GACTCTGCTGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGT
TCCTCGACGCTAACCTGGCGTAAGCGGCCGC

OST268 (SEQ ID NO: 202)

AAGCTTGGCACCACATGGGCGATAAAATTATTCACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGG
GGCGATCCTCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCGGTTGTACTAGTGCGGTGTGTGGTGCTTGGG
CTGAGGCGGGTAGGTTTTATTGTACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAA
TATCAGGGCAAACCTGACCGTTGCAAACTGAACATCGATCAAACCCCTGGCACTGCGCCGAAATATGGCATCCGTGG
TATCCCGACTCTGCTGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGA
AAGAGTTCCTCGACGCTAACCTGGCGTAAGCGGCCGC

OST269 (SEQ ID NO: 203)

(irrelevant control peptide for OST266-OST268)
AAGCTTGGCACCACATGGGCGATAAAATTATTCACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGG
GGCGATCCTCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCCTTGTGGATTGGGCCGGGTGATCAGGGTCTGT
TTCGGCGTTTTGTTTTTACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAG
GGCAAACCTGACCGTTGCAAACTGAACATCGATCAAACCCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCC
GACTCTGCTGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGT
TCCTCGACGCTAACCTGGCGTAAGCGGCCGC

FIGURE 12B

Listed below are the amino acid sequences corresponding to the pcDNA3.1 constructs in Appendix 1A

OST258

METDTLLLWVLLLWVPGSTGDGS

OST259 (SEQ ID NO: 204)

METDTLLLWVLLLWVPGSTGDGSM SDKI IHLTDDSFDTDLKADGAILVDFWAEWCGPNSVVLCSRCGRLWRWSCGT
SGPCKMIAPILDEIADEYQGKLTVAKLNI DQNP GTAPKYGIRGIPTLLL FKNGEVAATKVGALSKGQLKEFLDANLA

OST260 (SEQ ID NO: 205)

METDTLLLWVLLLWVPGSTGDGSM SDKI IHLTDDSFDTDLKADGAILVDFWAEWCGPNSGWRWCGRCGALWWRRT
SGPCKMIAPILDEIADEYQGKLTVAKLNI DQNP GTAPKYGIRGIPTLLL FKNGEVAATKVGALSKGQLKEFLDANLA

OST261 (SEQ ID NO: 206)

METDTLLLWVLLLWVPGSTGDGSM SDKI IHLTDDSFDTDLKADGAILVDFWAEWCGPNSEVRQVTCIRCRGFLLT
SGPCKMIAPILDEIADEYQGKLTVAKLNI DQNP GTAPKYGIRGIPTLLL FKNGEVAATKVGALSKGQLKEFLDANLA

OST262 (SEQ ID NO: 207)

METDTLLLWVLLLWVPGSTGDGSM SDKI IHLTDDSFDTDLKADGAILVDFWAEWCGPNSGGGGMIWEAWSCYACGT
SGPCKMIAPILDEIADEYQGKLTVAKLNI DQNP GTAPKYGIRGIPTLLL FKNGEVAATKVGALSKGQLKEFLDANLA

OST263 (SEQ ID NO: 208)

METDTLLLWVLLLWVPGSTGDGSM SDKI IHLTDDSFDTDLKADGAILVDFWAEWCGPNSLWIGPGDQGLFRRFVFT
SGPCKMIAPILDEIADEYQGKLTVAKLNI DQNP GTAPKYGIRGIPTLLL FKNGEVAATKVGALSKGQLKEFLDANLA

OST264 (SEQ ID NO: 209)

METDTLLLWVLLLWVPGSTGDGSSVSDQNHFRGEIEETITESFGNDHSTLDGYSRRTTLSSKMYHTKGQEGSVCLRS
SDCASGLCCARHFWSKICKPVLKEGQVCTKHRRKGS HGLEIFQRCYCGEGLS CRIQKDHQASNSSRLHTCQRH

OST265 (SEQ ID NO: 210)

METDTLLLWVLLLWVPGSTGDGSCASPTRGGDAGVQICLACRKRKRRCMRHAMCCPGNYCKNGICVSSDQNHFRGEI
EETITESFGNDHSTLDGYSRRTTLSSKMYHTKGQEGSVCLRSSDCASGLCCARHFWSKICKPVLKEGQVCTKHRRKGS
HGLEI
FQRCYCGEGLSC.

OST266 (SEQ ID NO: 211)

MGDKI IHLTDDSFDTDLKADGAILVDFWAEWCGPNSYAWLFSCSRCRWWLPWTS GPCCKMIAPILDEIADEYQGKLT
VAKLNI DQNP GTAPKYGIRGIPTLLL FKNGEVAATKVGALSKGQLKEFLDANLA

OST267 (SEQ ID NO: 212)

MGDKI IHLTDDSFDTDLKADGAILVDFWAEWCGPNSICEVVRLWSRYFWSWVTSGPCCKMIAPILDEIADEYQGKLT
VAKLNI DQNP GTAPKYGIRGIPTLLL FKNGEVAATKVGALSKGQLKEFLDANLA

OST268 (SEQ ID NO: 213)

MGDKI IHLTDDSFDTDLKADGAILVDFWAEWCGPNSGCTSAVCGAWAEAGREYCTSGPCCKMIAPILDEIADEYQKG
LTVAKLNI DQNP GTAPKYGIRGIPTLLL FKNGEVAATKVGALSKGQLKEFLDANLA

OST269 (SEQ ID NO: 214)

MGDKI IHLTDDSFDTDLKADGAILVDFWAEWCGPNSLWIGPGDQGLFRRFVFTSGPCCKMIAPILDEIADEYQGKLT
VAKLNI DQNP GTAPKYGIRGIPTLLL FKNGEVAATKVGALSKGQLKEFLDANLA

FIGURE 13

Effect of Dkks on Wnt1 signaling
with Coreceptors LRP5, HBM or LRP6
HOB03CE6 Cells

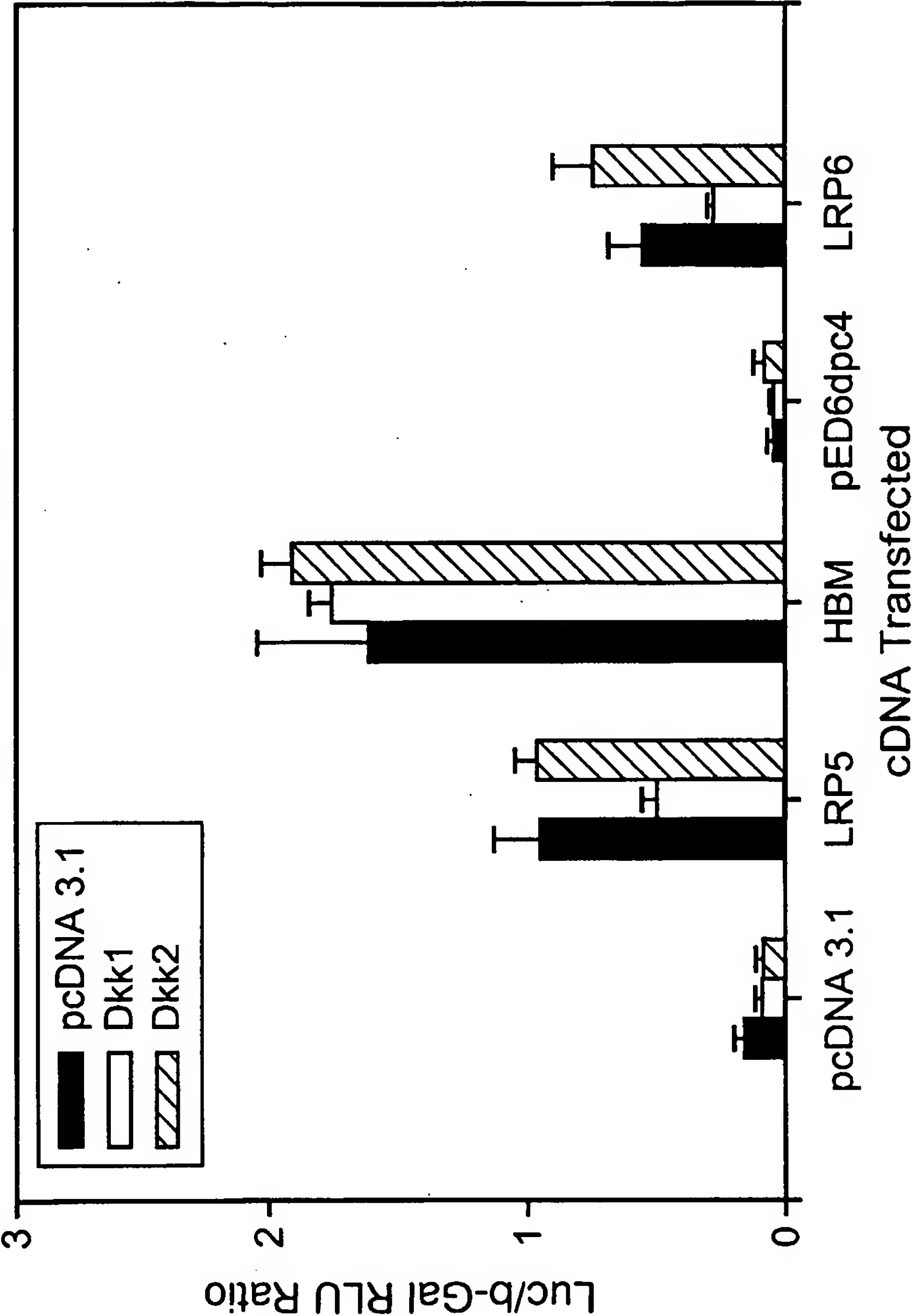


FIG. 14

Effect of Dkks on Wnt3a Signaling
with Coreceptors LRP5, HBM or LRP6
HOB03CE6 Cells

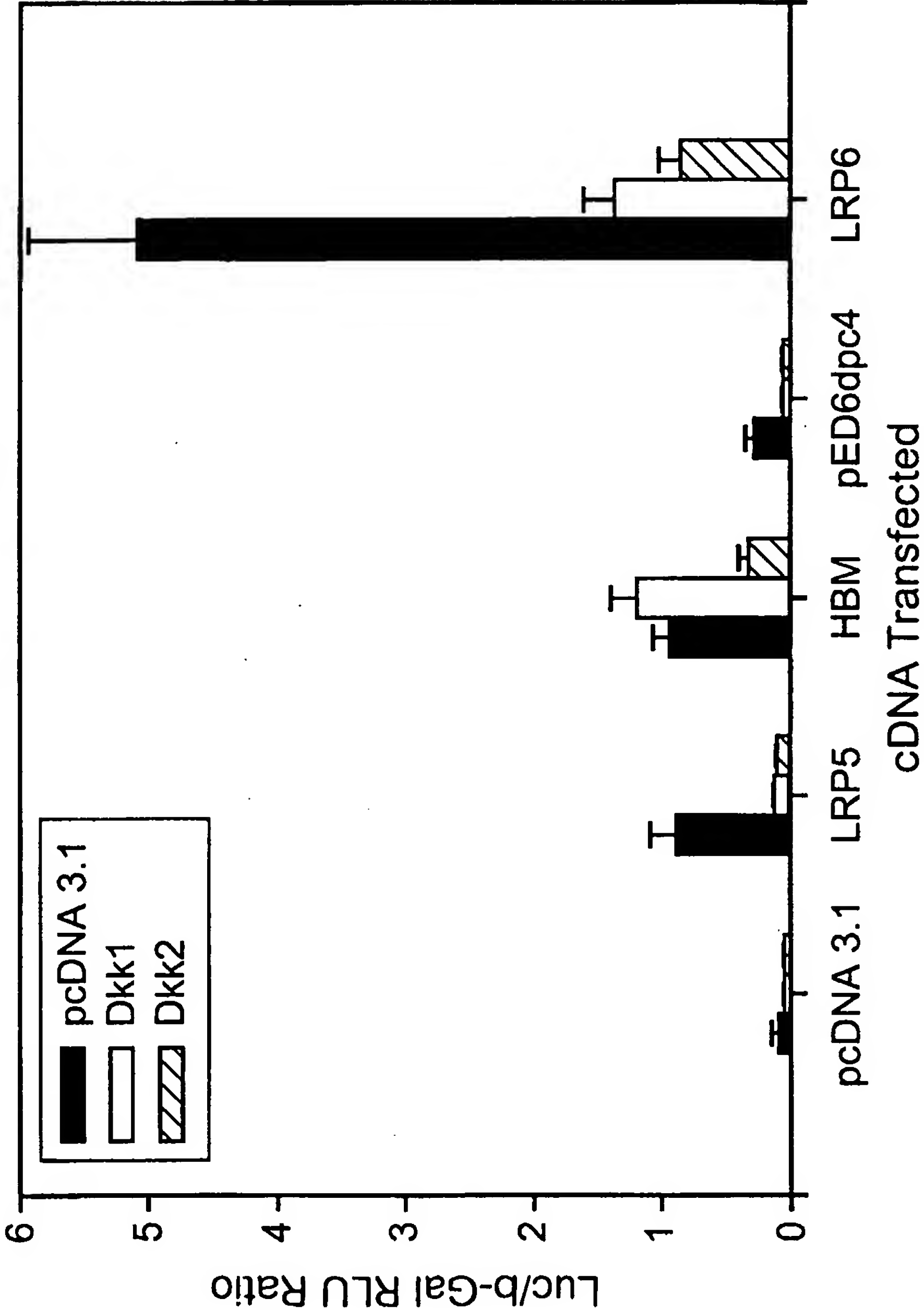


FIG. 15

LRP5-LBD Peptide Aptamer 262 Activates Wnt
Signaling in the presence of Wnt3a

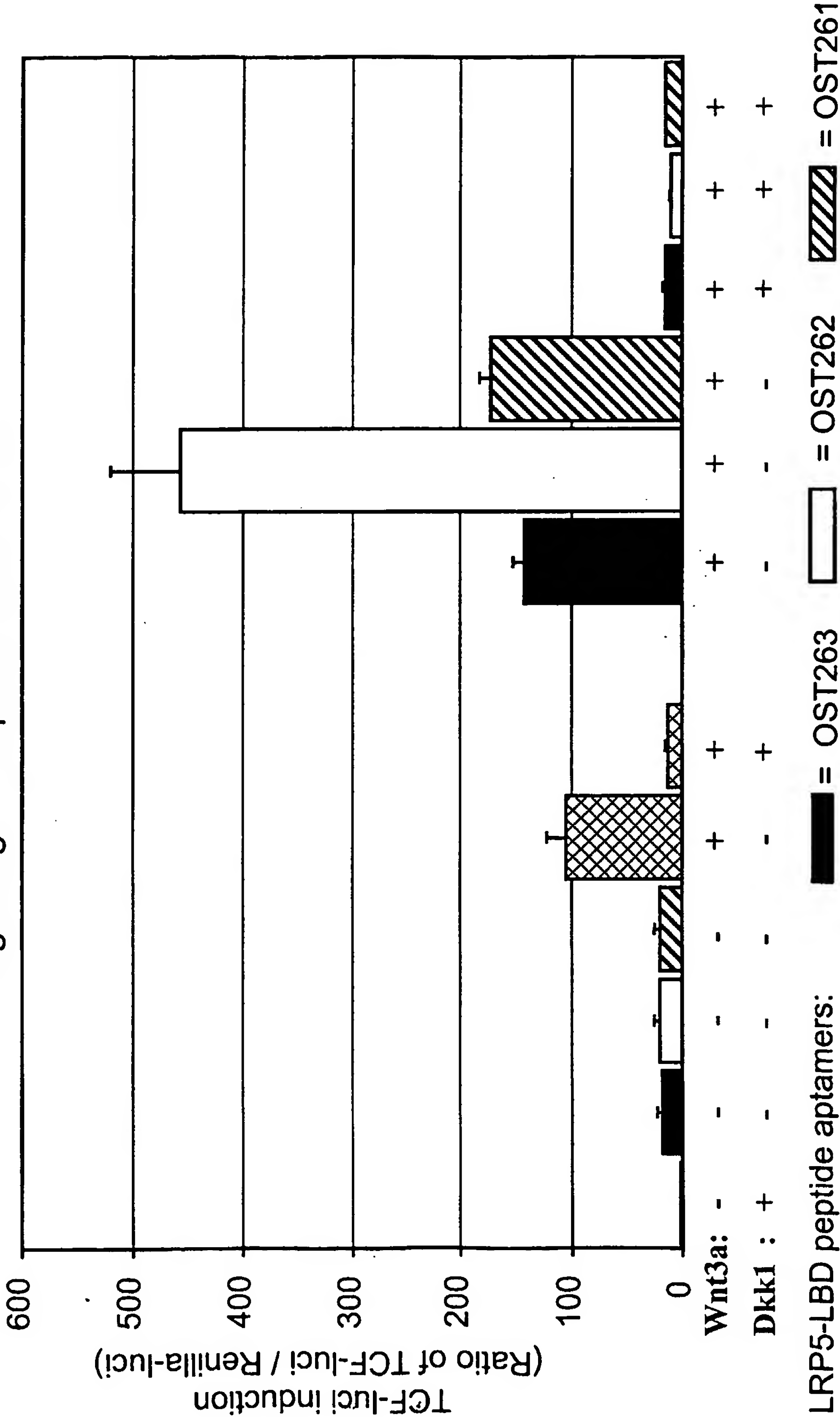
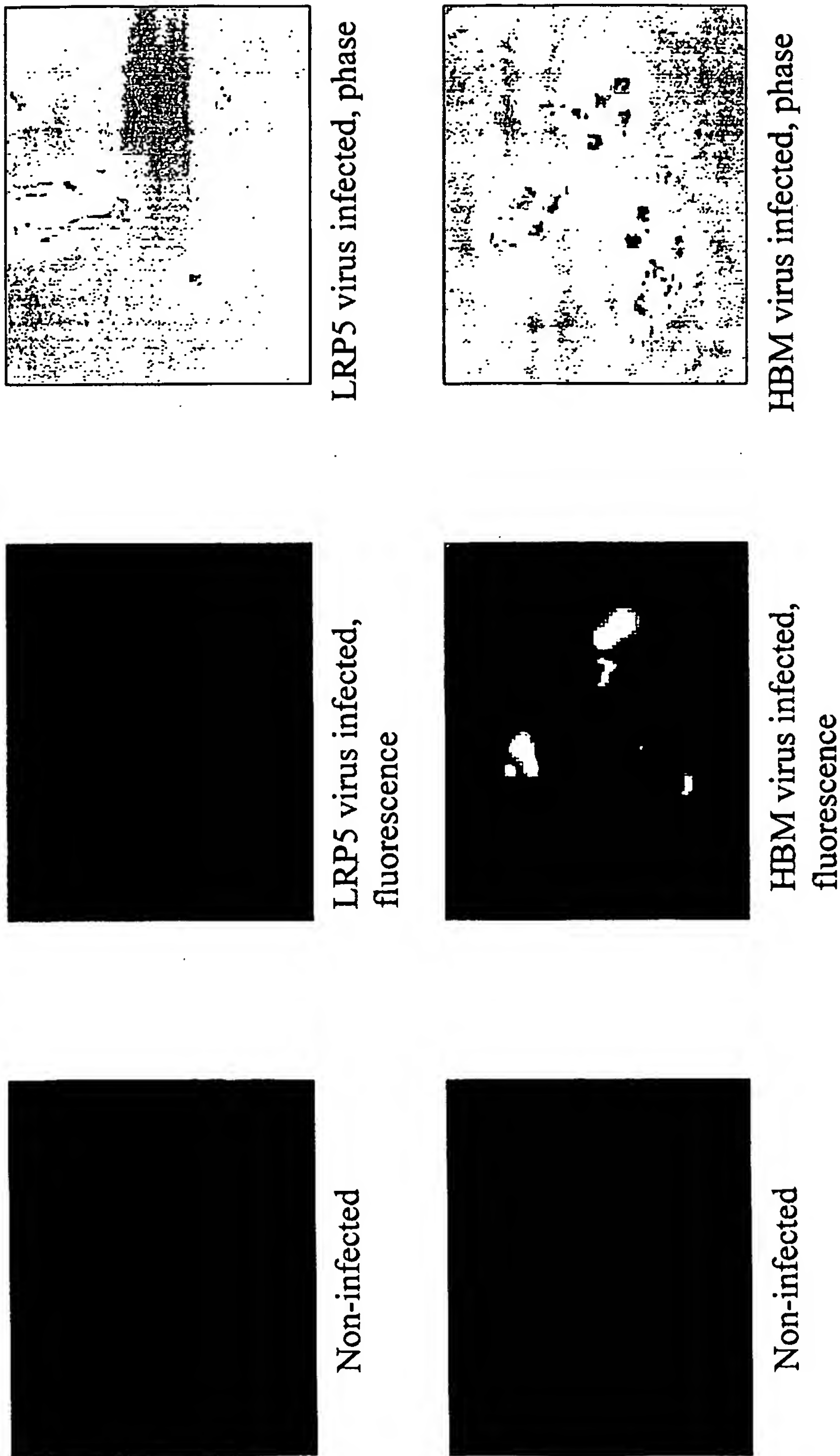


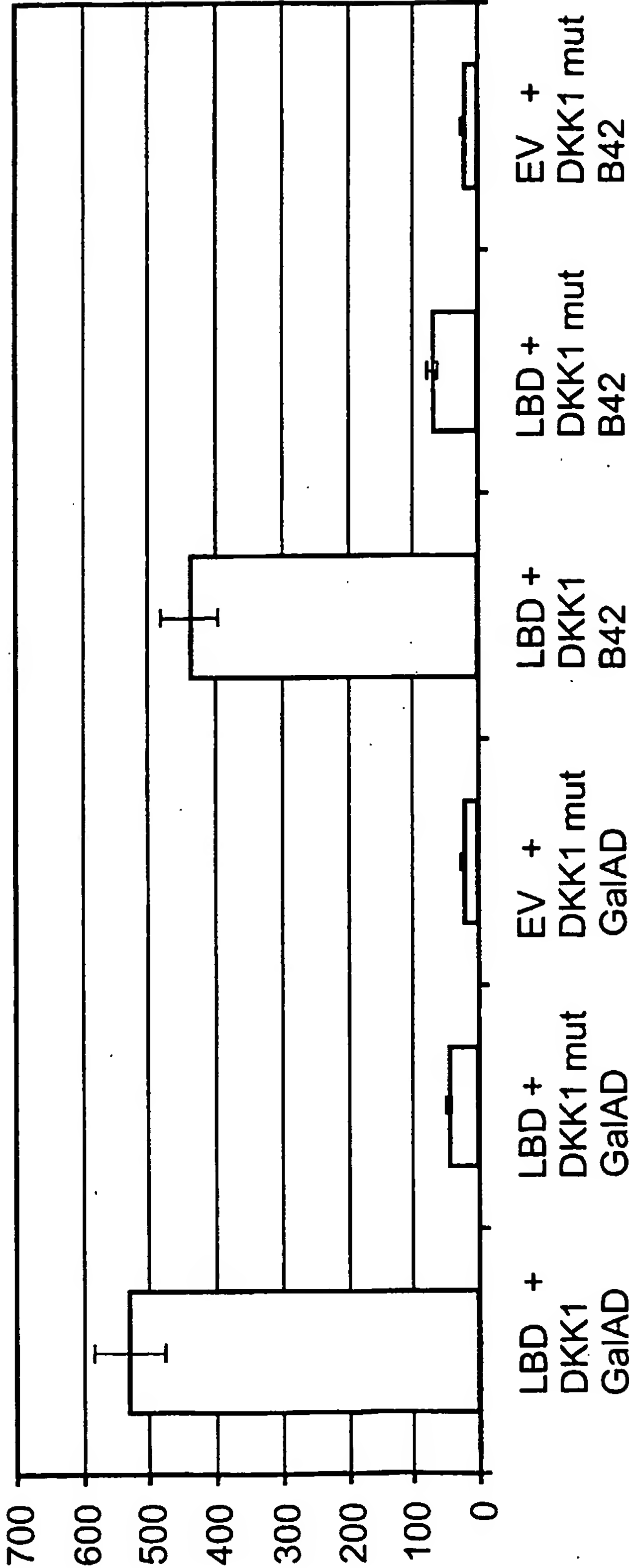
FIG. 16

FIG. 17
Affinity Purified 69546/47



Antibody to: aa 165-177 (Mutation)

EGY188 strain, 1 LexAop:LacZ reporter

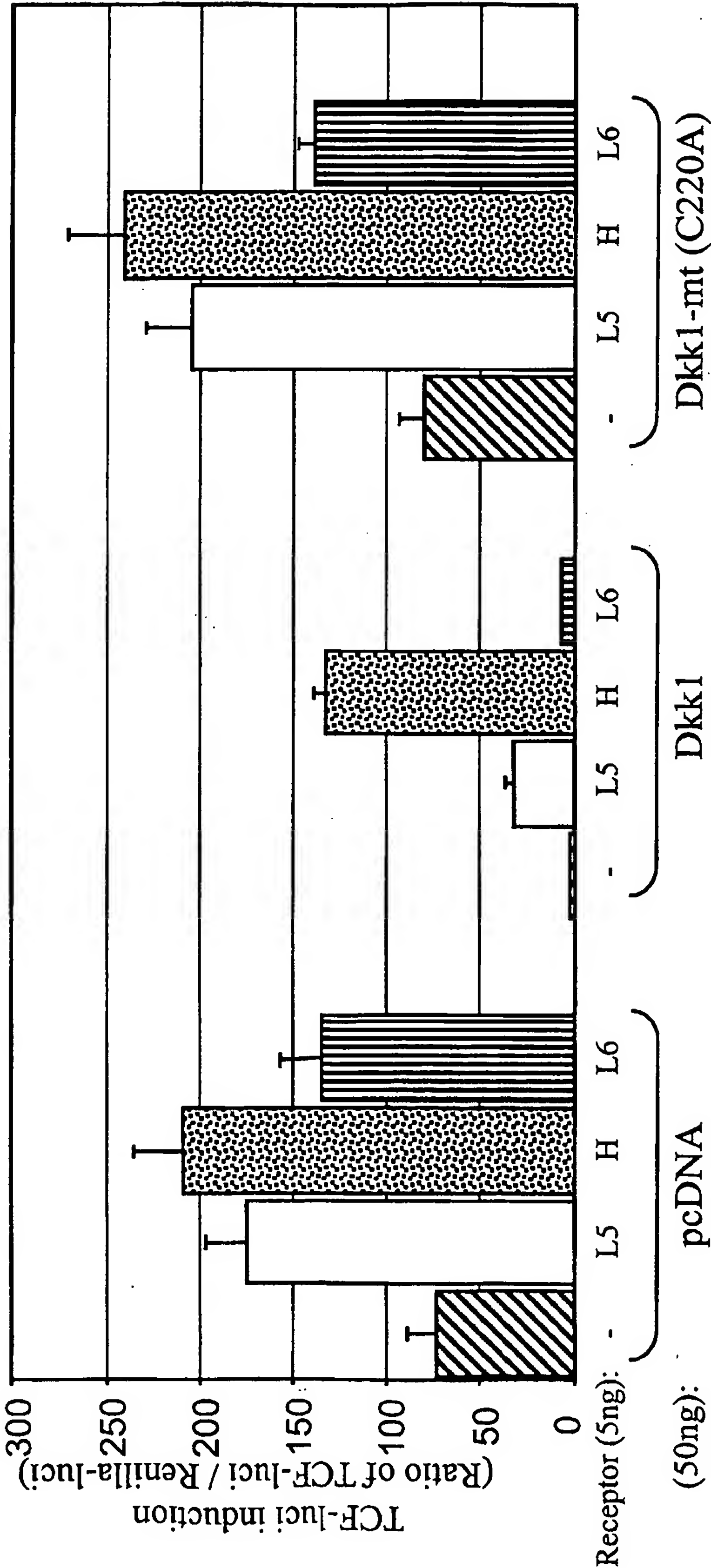


- A mutant DKK1, C220A, unable to bind to LRP5, was cloned in GalAD and B42 and tested for its ability to bind to LBD in Y2H
- Interaction LBD-DKK1 20 fold above background
- Interaction LBD-DKK1 C220A 2 to 3 fold above background
- Interaction LBD-DKK1 10 fold above LBD-DKK1 C220A mutant

FIG. 18

FIG. 20

Wnt1 - HBM generated TCF-luci is not efficiently inhibited
by Dkk1 in U2-OS bone cells.



- With Wnt1 the TCF-signal generated by LRP5 is greater than that of LRP6.
- LRP5/6 - Wnt1 induced TCF- is efficiently blocked by Dkk1

In U2-OS cells TCF-signal can be modulated by Dkk1, Dkk1-AP, without Wnt DNA transfection.

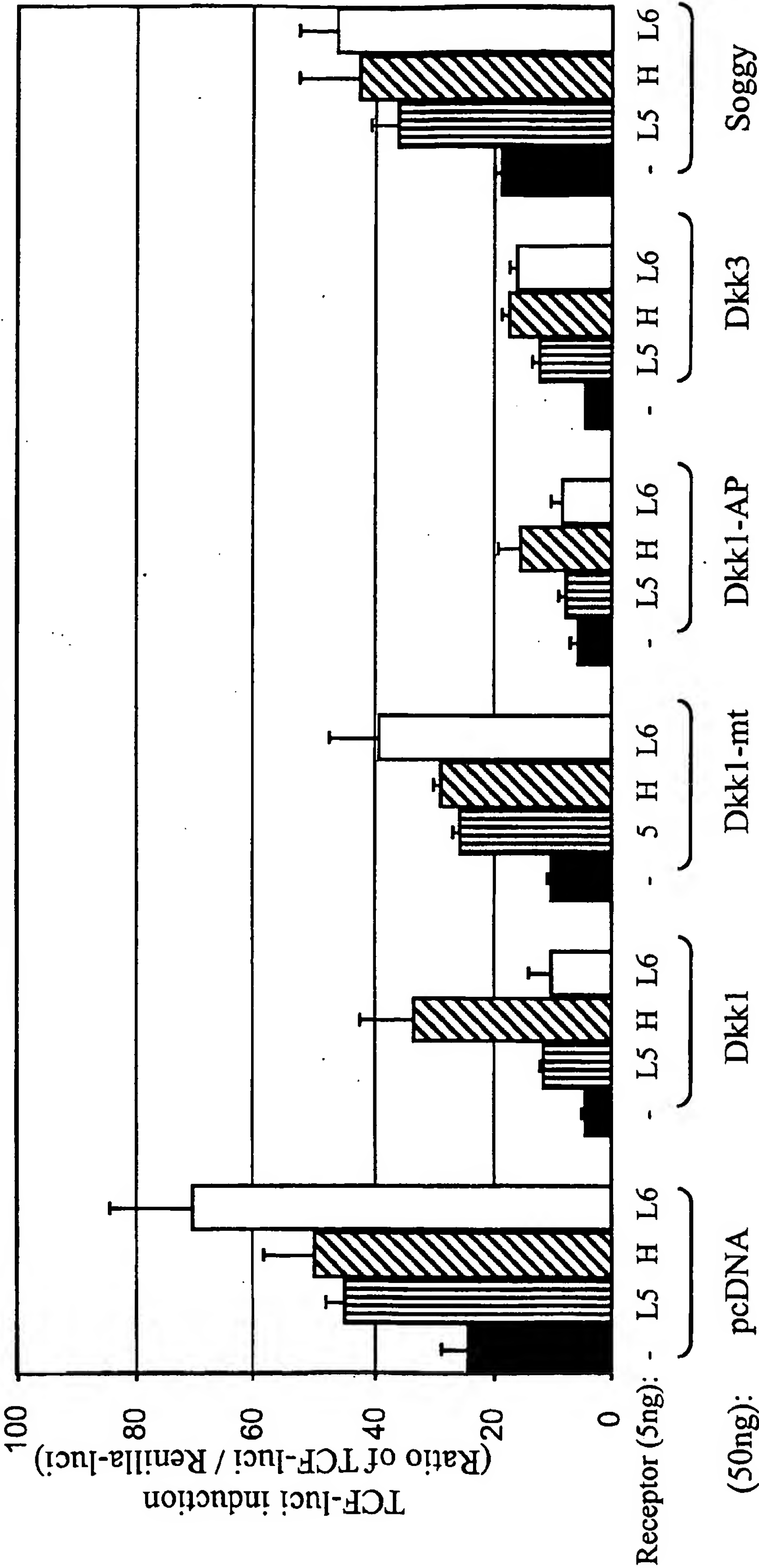


FIG. 21

FIG. 22

Aptamers 261 and 262 from the LRP5-LBD Activate Wnt Signaling in *Xenopus*



261 - LBD-Binding Peptide



262 - LBD-Binding Peptide



263 - Negative Control



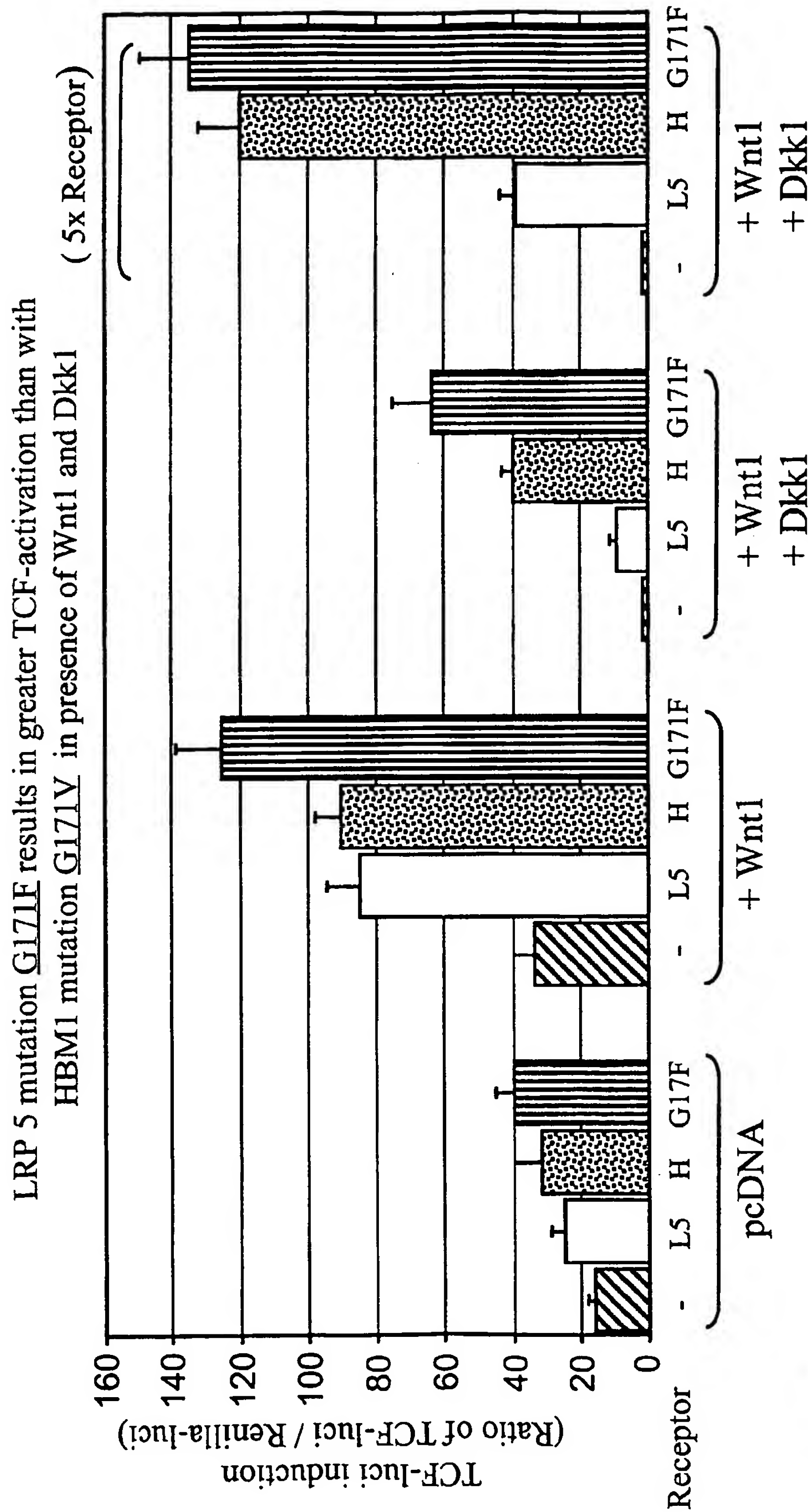
262 - LBD-Binding Peptide

FIG. 23

LRP5 Peptide Aptamers 261 and 262

Induce Wnt Signaling

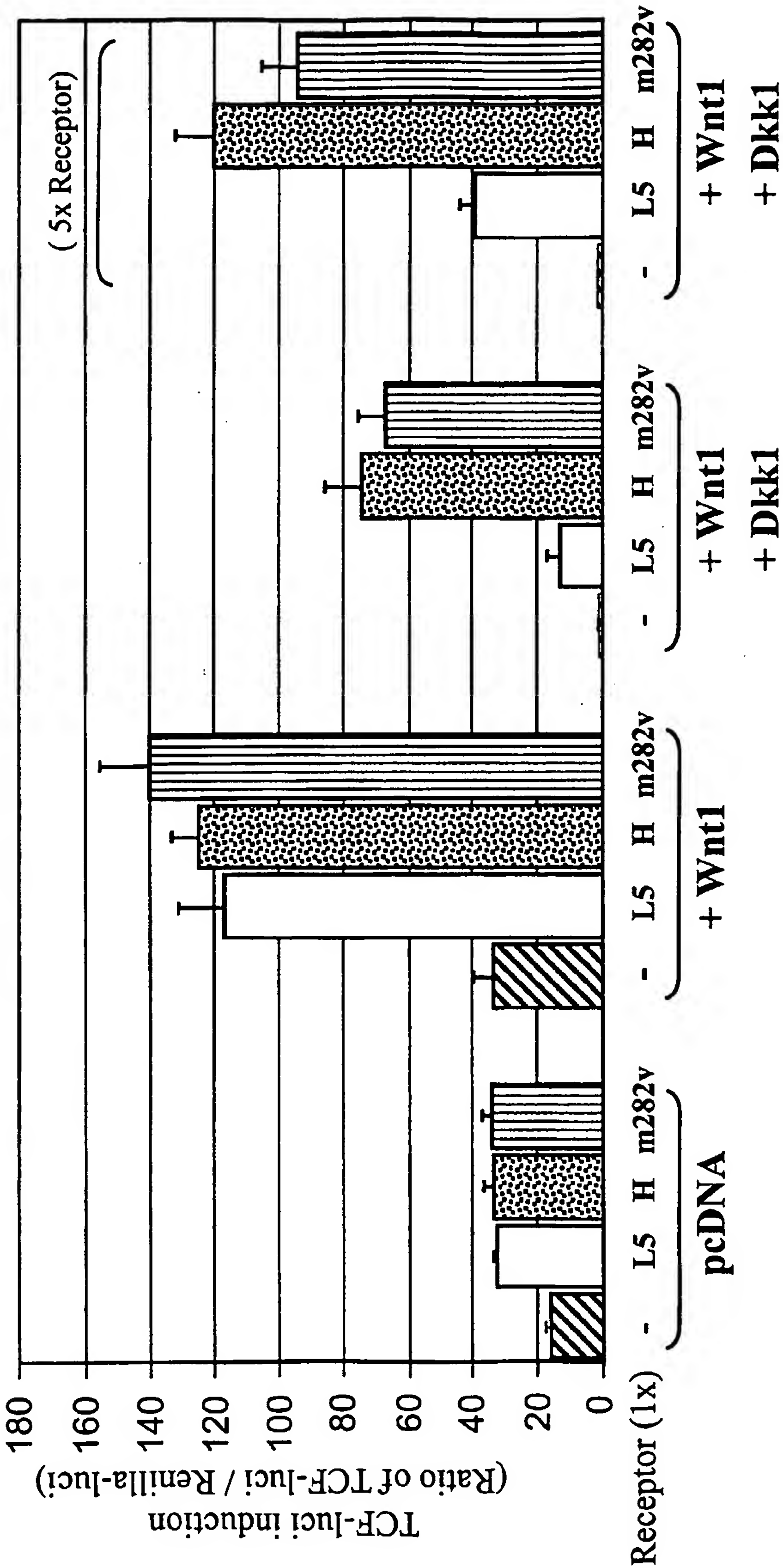




• G171F mutation involves the ringed R group (F) alteration and leads to marginally greater TCF-luciferase activation than that with HBM1 mutation G171V.

FIG. 24

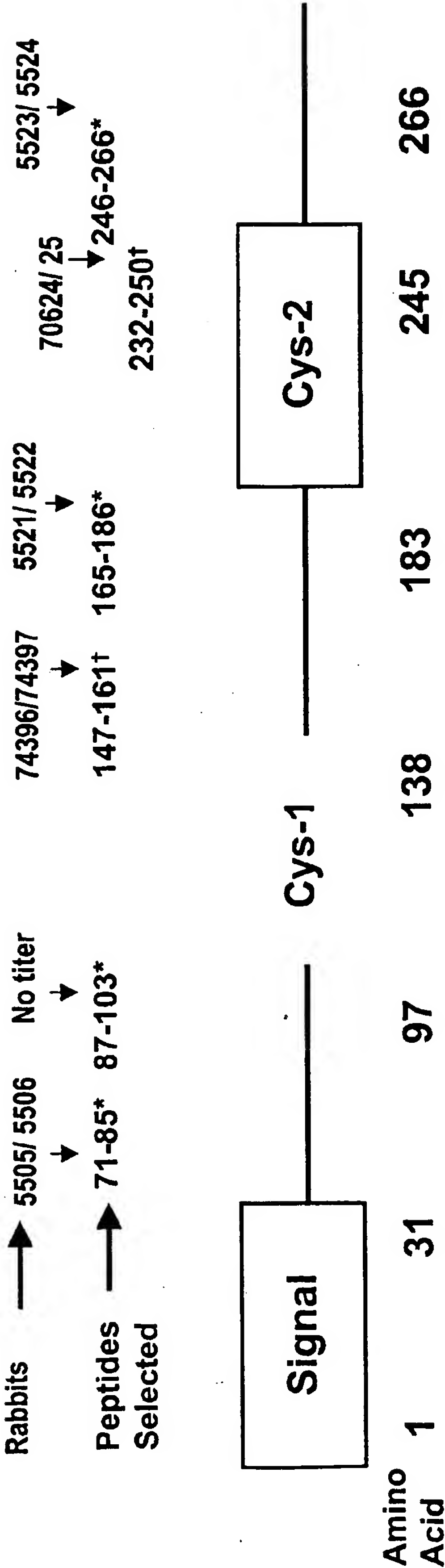
LRP5-blade1 Mutant M282V leads to HBM1 type TCF-signal
with Wnt1 and Dkk1 in U2-OS cells



- In blade 1, propeller 1, M282 is at the accessible interior position.
- It is conserved in propellers 1-3

FIG. 25

DKK1 Protein
Polyclonal Antibodies



* Sigma/Genosys

† ResGen

FIG. 26

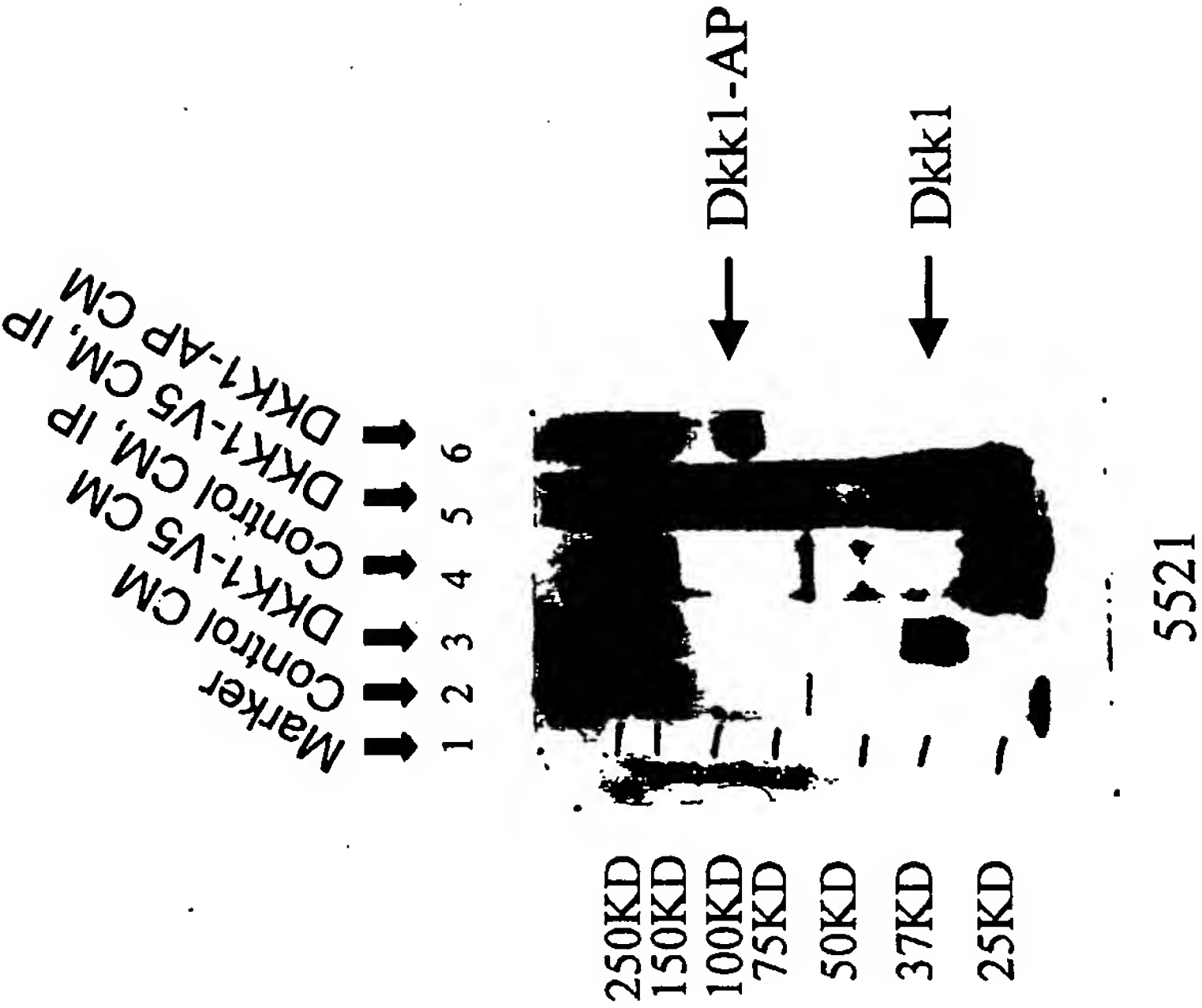


FIG. 27B

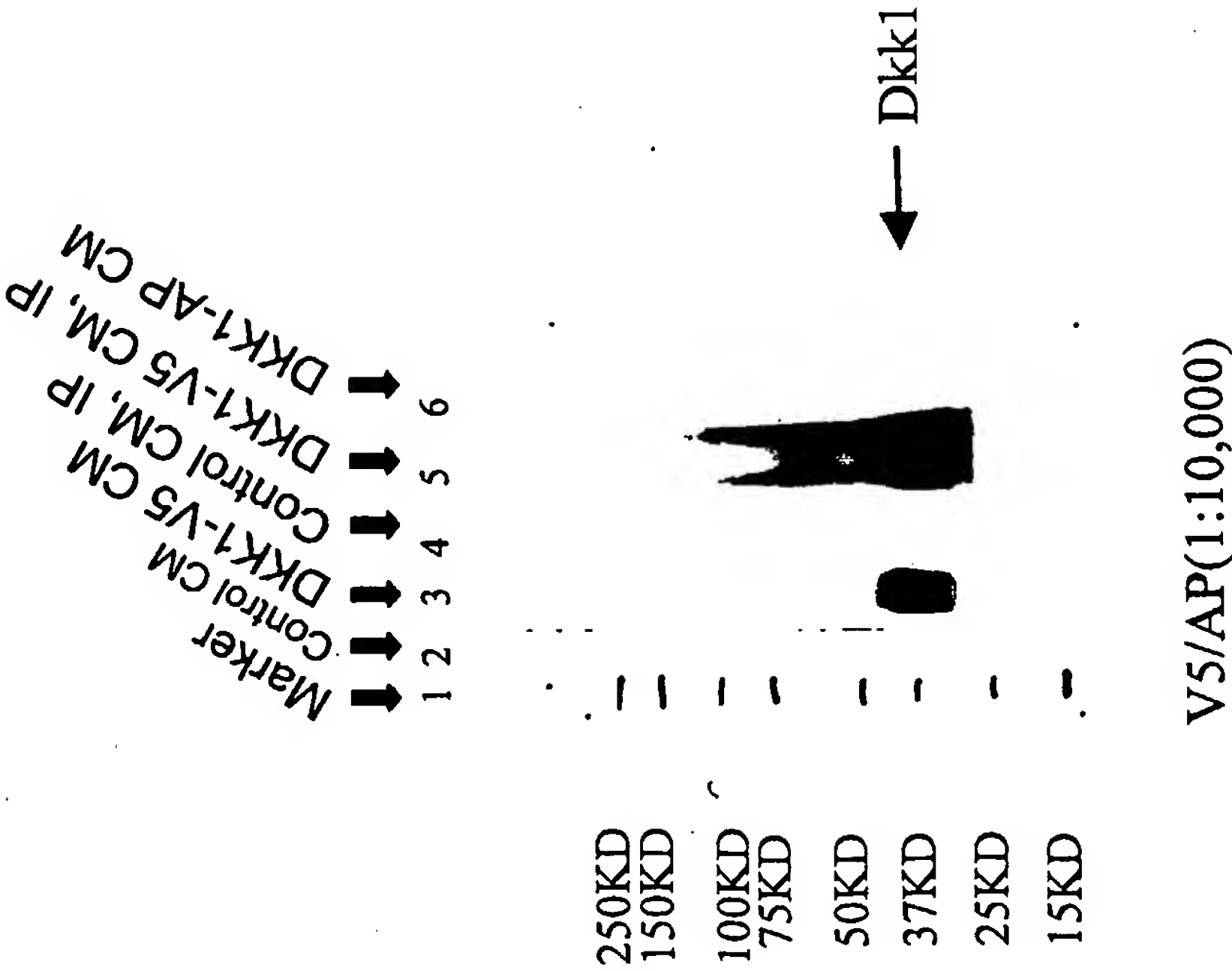


FIG. 27A

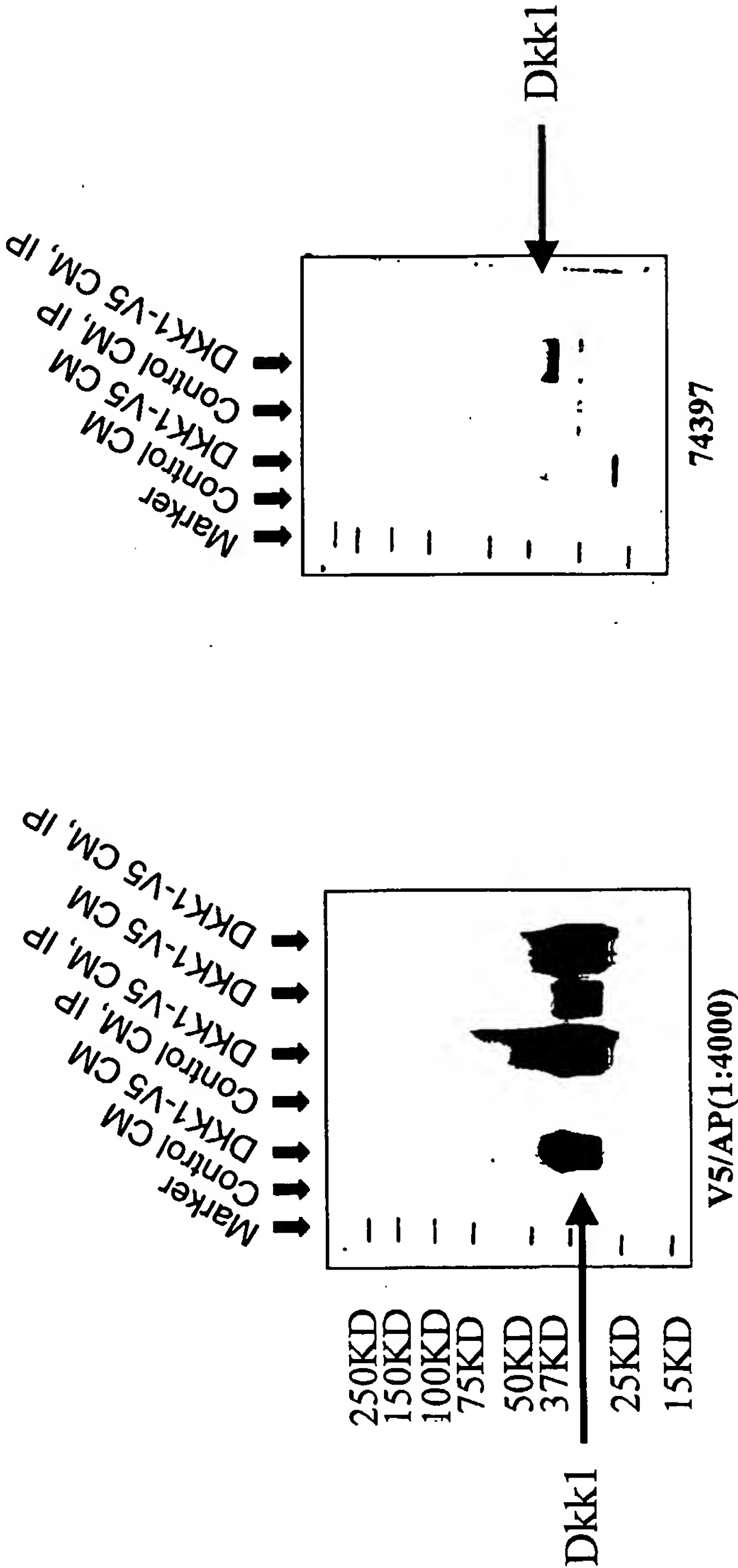


FIG. 28

032796-132.ST25

SEQUENCE LISTING

<110> Allen, Kristina M.
 Anisowicz, Anthony
 Bhat, Bheem
 Damagnez, Veronique
 Robinson, John
 Yaworsky, Paul

<120> Reagents and Method for Modulating DKK-Mediated Interactions

<130> 032796-132

<150> US 60/291,311

<151> 2001-05-17

<150> US 60/353,058

<151> 2002-02-01

<150> US 60/361,293

<151> 2002-03-04

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<170> FastSEQ for Windows Version 4.0

<210> 1

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<212> DNA

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Met Glu Ala Ala Pro Pro Gly Pro Pro Trp Pro Leu Leu	
1 5 10	
ctg ctg ctg ctg ctg ctg ctg gcg ctg tgc ggc tgc ccg gcc ccc gcc	157
Leu Leu Leu Leu Leu Leu Leu Ala Leu Cys Gly Cys Pro Ala Pro Ala	
15 20 25	
gcg gcc tcg ccg ctc ctg cta ttt gcc aac cgc cgg gac gta cgg ctg	205
Ala Ala Ser Pro Leu Leu Leu Phe Ala Asn Arg Arg Asp Val Arg Leu	
30 35 40 45	
gtg gac gcc ggc gga gtc aag ctg gag tcc acc atc gtg gtc agc ggc	253
Val Asp Ala Gly Gly Val Lys Leu Glu Ser Thr Ile Val Val Ser Gly	
50 55 60	
ctg gag gat gcg gcc gca gtg gac ttc cag ttt tcc aag gga gcc gtg	301
Leu Glu Asp Ala Ala Ala Val Asp Phe Gln Phe Ser Lys Gly Ala Val	
65 70 75	
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Tyr Trp Thr Asp Val Ser Glu Glu Ala Ile Lys Gln Thr Tyr Leu Asn	
80 85 90	
cag acg ggg gcc gcc gtg cag aac gtg gtc atc tcc ggc ctg gtc tct	397

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Gln	Thr	Gly	Ala	Ala	Val	Gln	Asn	Val	Val	Ile	Ser	Gly	Leu	Val	Ser		
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ccc	gac	ggc	ctc	gcc	tgc	gac	tgg	gtg	ggc	aag	aag	ctg	tac	tgg	acg		445
Pro	Asp	Gly	Leu	Ala	Cys	Asp	Trp	Val	Gly	Lys	Lys	Leu	Tyr	Trp	Thr		
110					115					120					125		
gac	tca	gag	acc	aac	cgc	atc	gag	gtg	gcc	aac	ctc	aat	ggc	aca	tcc		493
Asp	Ser	Glu	Thr	Asn	Arg	Ile	Glu	Val	Ala	Asn	Leu	Asn	Gly	Thr	Ser		
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cgg	aag	gtg	ctc	ttc	tgg	cag	gac	ctt	gac	cag	ccg	agg	gcc	atc	gcc		541
Arg	Lys	Val	Leu	Phe	Trp	Gln	Asp	Leu	Asp	Gln	Pro	Arg	Ala	Ile	Ala		
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Leu	Asp	Pro	Ala	His	Gly	Tyr	Met	Tyr	Trp	Thr	Asp	Trp	Gly	Glu	Thr		
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Val	Asp	Ser	Asp	Ile	Tyr	Trp	Pro	Asn	Gly	Leu	Thr	Ile	Asp	Leu	Glu		
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Glu	Gln	Lys	Leu	Tyr	Trp	Ala	Asp	Ala	Lys	Leu	Ser	Phe	Ile	His	Arg		
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gcc	aac	ctg	gac	ggc	tcg	ttc	cgg	cag	aag	gtg	gtg	gag	ggc	agc	ctg		781
Ala	Asn	Leu	Asp	Gly	Ser	Phe	Arg	Gln	Lys	Val	Val	Glu	Gly	Ser	Leu		
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Thr	His	Pro	Phe	Ala	Leu	Thr	Leu	Ser	Gly	Asp	Thr	Leu	Tyr	Trp	Thr		
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Asp	Trp	Gln	Thr	Arg	Ser	Ile	His	Ala	Cys	Asn	Lys	Arg	Thr	Gly	Gly		
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Lys	Arg	Lys	Glu	Ile	Leu	Ser	Ala	Leu	Tyr	Ser	Pro	Met	Asp	Ile	Gln		
270					275					280					285		
gtg	ctg	agc	cag	gag	cgg	cag	cct	ttc	ttc	cac	act	cgc	tgt	gag	gag		973
Val	Leu	Ser	Gln	Glu	Arg	Gln	Pro	Phe	Phe	His	Thr	Arg	Cys	Glu	Glu		
			290						295					300			
gac	aat	ggc	ggc	tgc	tcc	cac	ctg	tgc	ctg	ctg	tcc	cca	agc	gag	cct		1021
Asp	Asn	Gly	Gly	Cys	Ser	His	Leu	Cys	Leu	Leu	Ser	Pro	Ser	Glu	Pro		
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ttc	tac	aca	tgc	gcc	tgc	ccc	acg	ggg	gtg	cag	ctg	cag	gac	aac	ggc		1069
Phe	Tyr	Thr	Cys	Ala	Cys	Pro	Thr	Gly	Val	Gln	Leu	Gln	Asp	Asn	Gly		
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Thr	Asp	Leu	Arg	Arg	Ile	Ser	Leu	Asp	Thr	Pro	Asp	Phe	Thr	Asp	Ile		
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Val	Leu	Gln	Val	Asp	Asp	Ile	Arg	His	Ala	Ile	Ala	Ile	Asp	Tyr	Asp		
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Pro	Leu	Glu	Gly	Tyr	Val	Tyr	Trp	Thr	Asp	Asp	Glu	Val	Arg	Ala	Ile		
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Ala	Thr	Arg	Cys	Gly	Cys	Pro	Ile	Gly	Leu	Glu	Leu	Leu	Ser	Asp	Met		
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Lys	Thr	Cys	Ile	Val	Pro	Glu	Ala	Phe	Leu	Val	Phe	Thr	Ser	Arg	Ala		
		640					645					650					
gcc	atc	cac	agg	atc	tcc	ctc	gag	acc	aat	aac	aac	gac	gtg	gcc	atc		2077
Ala	Ile	His	Arg	Ile	Ser	Leu	Glu	Thr	Asn	Asn	Asn	Asp	Val	Ala	Ile		
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Pro	Leu	Thr	Gly	Val	Lys	Glu	Ala	Ser	Ala	Leu	Asp	Phe	Asp	Val	Ser		
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aac	aac	cac	atc	tac	tgg	aca	gac	gtc	agc	ctg	aag	acc	atc	agc	cgc		2173
Asn	Asn	His	Ile	Tyr	Trp	Thr	Asp	Val	Ser	Leu	Lys	Thr	Ile	Ser	Arg		
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ctg	gcc	ctg	gat	ccc	acc	aag	ggc	tac	atc	tac	tgg	acc	gag	tgg	ggc	2413
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Cys	Gly	Cys	Ala	Ser	His	Tyr	Thr	Leu	Asp	Pro	Ser	Ser	Arg	Asn	Cys	
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Ser	Pro	Pro	Thr	Thr	Phe	Leu	Leu	Phe	Ser	Gln	Lys	Ser	Ala	Ile	Ser	
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Arg	Met	Ile	Pro	Asp	Asp	Gln	His	Ser	Pro	Asp	Leu	Ile	Leu	Pro	Leu	
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His	Gly	Leu	Arg	Asn	Val	Lys	Ala	Ile	Asp	Tyr	Asp	Pro	Leu	Asp	Lys	
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Pro Asp Arg Gln Pro His Asp Leu Ser Ile Asp Ile Tyr Ser Arg Thr	
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Ala Lys Gly Asp Gly Thr Pro Arg Cys Ser Cys Pro Val His Leu Val	
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Trp Arg Cys Asp Gly Phe Pro Glu Cys Asp Asp Gln Ser Asp Glu Glu	
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Gly Cys Pro Val Cys Ser Ala Ala Gln Phe Pro Cys Ala Arg Gly Gln	
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Leu Asn Ser Asp Ser Asp Pro Tyr Pro Pro Pro Pro Thr Pro His Ser	
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Gln Tyr Leu Ser Ala Glu Asp Ser Cys Pro Pro Ser Pro Ala Thr Glu	
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Arg Ser Tyr Phe His Leu Phe Pro Pro Pro Pro Ser Pro Cys Thr Asp	
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Ser Ser
1615

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<212> DNA

<213> Homo sapiens

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Met.Glu Ala Ala Pro Pro Gly Pro Pro Trp Pro Leu Leu
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Leu Leu Leu Leu Leu Leu Leu Ala Leu Cys Gly Cys Pro Ala Pro Ala
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Ala Ala Ser Pro Leu Leu Leu Phe Ala Asn Arg Arg Asp Val Arg Leu
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Val Asp Ala Gly Gly Val Lys Leu Glu Ser Thr Ile Val Val Ser Gly
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Tyr Trp Thr Asp Val Ser Glu Glu Ala Ile Lys Gln Thr Tyr Leu Asn
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Gln Thr Gly Ala Ala Val Gln Asn Val Val Ile Ser Gly Leu Val Ser
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Pro Arg Ile Glu Arg Ala Gly Met Asp Gly Ser Thr Arg Lys Ile Ile
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Glu Gln Lys Leu Tyr Trp Ala Asp Ala Lys Leu Ser Phe Ile His Arg

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Lys	Arg	Lys	Glu	Ile	Leu	Ser	Ala	Leu	Tyr	Ser	Pro	Met	Asp	Ile	Gln				
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Val	Leu	Gln	Val	Asp	Asp	Ile	Arg	His	Ala	Ile	Ala	Ile	Asp	Tyr	Asp				
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Ile	Tyr	Trp	Thr	Asp	Trp	Gln	Arg	Arg	Ser	Ile	Glu	Arg	Val	His	Lys	
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gcc	ttc	atg	aac	ggg	agc	tcg	gtg	gag	cac	gtg	gtg	gag	ttt	ggc	ctt	2221
Ala	Phe	Met	Asn	Gly	Ser	Ser	Val	Glu	His	Val	Val	Glu	Phe	Gly	Leu	
			705					710					715			
gac	tac	ccc	gag	ggc	atg	gcc	gtt	gac	tgg	atg	ggc	aag	aac	ctc	tac	2269
Asp	Tyr	Pro	Glu	Gly	Met	Ala	Val	Asp	Trp	Met	Gly	Lys	Asn	Leu	Tyr	
		720					725					730				
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Trp	Ala	Asp	Thr	Gly	Thr	Asn	Arg	Ile	Glu	Val	Ala	Arg	Leu	Asp	Gly	
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cag	ttc	cgg	caa	gtc	ctc	gtg	tgg	agg	gac	ttg	gac	aac	ccg	agg	tcg	2365
Gln	Phe	Arg	Gln	Val	Leu	Val	Trp	Arg	Asp	Leu	Asp	Asn	Pro	Arg	Ser	
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ctg	gcc	ctg	gat	ccc	acc	aag	ggc	tac	atc	tac	tgg	acc	gag	tgg	ggc	2413
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Gly	Lys	Pro	Arg	Ile	Val	Arg	Ala	Phe	Met	Asp	Gly	Thr	Asn	Cys	Met	
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Thr	Leu	Val	Asp	Lys	Val	Gly	Arg	Ala	Asn	Asp	Leu	Thr	Ile	Asp	Tyr	
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815						820					825					
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Ser	Ser	Asn	Met	Leu	Gly	Gln	Glu	Arg	Val	Val	Ile	Ala	Asp	Asp	Leu	
830					835					840					845	
ccg	cac	ccg	ttc	ggc	ctg	acg	cag	tac	agc	gat	tat	atc	tac	tgg	aca	2653
Pro	His	Pro	Phe	Gly	Leu	Thr	Gln	Tyr	Ser	Asp	Tyr	Ile	Tyr	Trp	Thr	
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gac	tgg	aac	ctg	cac	agc	att	gag	cgg	gcc	gac	aag	act	agc	ggc	cgg	2701
Asp	Trp	Asn	Leu	His	Ser	Ile	Glu	Arg	Ala	Asp	Lys	Thr	Ser	Gly	Arg	
			865					870					875			
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Asn	Arg	Thr	Leu	Ile	Gln	Gly	His	Leu	Asp	Phe	Val	Met	Asp	Ile	Leu	
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gtg	ttc	cac	tcc	tcc	cgc	cag	gat	ggc	ctc	aac	gac	tgt	atg	cac	aac	2797
Val	Phe	His	Ser	Ser	Arg	Gln	Asp	Gly	Leu	Asn	Asp	Cys	Met	His	Asn	
	895					900					905					
aac	ggg	cag	tgt	ggg	cag	ctg	tgc	ctt	gcc	atc	ccc	ggc	ggc	cac	cgc	2845
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tgc	ggc	tgc	gcc	tca	cac	tac	acc	ctg	gac	ccc	agc	agc	cgc	aac	tgc	2893
Cys	Gly	Cys	Ala	Ser	His	Tyr	Thr	Leu	Asp	Pro	Ser	Ser	Arg	Asn	Cys	
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agc	ccg	ccc	acc	acc	ttc	ttg	ctg	ttc	agc	cag	aaa	tct	gcc	atc	agt	2941
Ser	Pro	Pro	Thr	Thr	Phe	Leu	Leu	Phe	Ser	Gln	Lys	Ser	Ala	Ile	Ser	
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Arg	Met	Ile	Pro	Asp	Asp	Gln	His	Ser	Pro	Asp	Leu	Ile	Leu	Pro	Leu	
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Leu	Phe	Trp	Thr	Cys	Glu	Ala	Thr	Asn	Thr	Ile	Asn	Val	His	Arg	Leu	
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Ser	Gly	Glu	Ala	Met	Gly	Val	Val	Leu	Arg	Gly	Asp	Arg	Asp	Lys	Pro	
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Arg	Ala	Ile	Val	Val	Asn	Ala	Glu	Arg	Gly	Tyr	Leu	Tyr	Phe	Thr	Asn	
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Lys	Arg	Ile	Glu	Ser	Cys	Asp	Leu	Ser	Gly	Ala	Asn	Arg	Leu	Thr	Leu	
		1135					1140					1145				
gag	gac	gcc	aac	atc	gtg	cag	cct	ctg	ggc	ctg	acc	atc	ctt	ggc	aag	3565
Glu	Asp	Ala	Asn	Ile	Val	Gln	Pro	Leu	Gly	Leu	Thr	Ile	Leu	Gly	Lys	
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His	Leu	Tyr	Trp	Ile	Asp	Arg	Gln	Gln	Gln	Met	Ile	Glu	Arg	Val	Glu	
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Lys	Thr	Thr	Gly	Asp	Lys	Arg	Thr	Arg	Ile	Gln	Gly	Arg	Val	Ala	His	
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Leu	Thr	Gly	Ile	His	Ala	Val	Glu	Glu	Val	Ser	Leu	Glu	Glu	Phe	Ser	
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Trp	Arg	Cys	Asp	Gly	Phe	Pro	Glu	Cys	Asp	Asp	Gln	Ser	Asp	Glu	Glu	
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Val	Cys	Gln	Arg	Val	Val	Cys	Gln	Arg	Tyr	Ala	Gly	Ala	Asn	Gly	Pro	
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Pro	Ser	Pro	Ala	Thr	Asp	Pro	Ser	Leu	Tyr	Asn	Met	Asp	Met	Phe	Tyr		
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Ser	Ser	Asn	Ile	Pro	Ala	Thr	Ala	Arg	Pro	Tyr	Arg	Pro	Tyr	Ile	Ile		
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Arg	Gly	Met	Ala	Pro	Pro	Thr	Thr	Pro	Cys	Ser	Thr	Asp	Val	Cys	Asp		
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Ser	Asp	Tyr	Ser	Ala	Ser	Arg	Trp	Lys	Ala	Ser	Lys	Tyr	Tyr	Leu	Asp		
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Leu	Asn	Ser	Asp	Ser	Asp	Pro	Tyr	Pro	Pro	Pro	Pro	Thr	Pro	His	Ser		
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Gln	Tyr	Leu	Ser	Ala	Glu	Asp	Ser	Cys	Pro	Pro	Ser	Pro	Ala	Thr	Glu		
		1585					1590					1595					
agg	agc	tac	ttc	cat	ctc	ttc	ccg	ccc	cct	ccg	tcc	ccc	tgc	acg	gac		4909
Arg	Ser	Tyr	Phe	His	Leu	Phe	Pro	Pro	Pro	Pro	Ser	Pro	Cys	Thr	Asp		
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Ser	Ser																
		1615															
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<211> 1615

<212> PRT

<213> Homo sapiens

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			20					25					30				
Pro	Leu	Leu	Leu	Phe	Ala	Asn	Arg	Arg	Asp	Val	Arg	Leu	Val	Asp	Ala		
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Ala Ala Ala Val Asp Phe Gln Phe Ser Lys Gly Ala Val Tyr Trp Thr		
65	70	75
Asp Val Ser Glu Glu Ala Ile Lys Gln Thr Tyr Leu Asn Gln Thr Gly		80
	85	90
Ala Ala Val Gln Asn Val Val Ile Ser Gly Leu Val Ser Pro Asp Gly		95
	100	105
Leu Ala Cys Asp Trp Val Gly Lys Lys Leu Tyr Trp Thr Asp Ser Glu		110
	115	120
Thr Asn Arg Ile Glu Val Ala Asn Leu Asn Gly Thr Ser Arg Lys Val		125
	130	135
Leu Phe Trp Gln Asp Leu Asp Gln Pro Lys Ala Ile Ala Leu Asp Pro		140
145	150	155
Ala His Gly Tyr Met Tyr Trp Thr Asp Trp Gly Glu Thr Pro Arg Ile		160
	165	170
Glu Arg Ala Gly Met Asp Gly Ser Thr Arg Lys Ile Ile Val Asp Ser		175
	180	185
Asp Ile Tyr Trp Pro Asn Gly Leu Thr Ile Asp Leu Glu Glu Gln Lys		190
	195	200
Leu Tyr Trp Ala Asp Ala Lys Leu Ser Phe Ile His Arg Ala Asn Leu		205
	210	215
Asp Gly Ser Phe Arg Gln Lys Val Val Glu Gly Ser Leu Thr His Pro		220
225	230	235
Phe Ala Leu Thr Leu Ser Gly Asp Thr Leu Tyr Trp Thr Asp Trp Gln		240
	245	250
Thr Arg Ser Ile His Ala Cys Asn Lys Arg Thr Gly Gly Lys Arg Lys		255
	260	265
Glu Ile Leu Ser Ala Leu Tyr Ser Pro Met Asp Ile Gln Val Leu Ser		270
	275	280
Gln Glu Arg Gln Pro Phe Phe His Thr Arg Cys Glu Glu Asp Asn Gly		285
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Gly Trp Ser His Leu Cys Leu Leu Ser Pro Ser Glu Pro Phe Tyr Thr		300
305	310	315
Cys Ala Cys Pro Thr Gly Val Gln Met Gln Asp Asn Gly Arg Thr Cys		320
	325	330
Lys Ala Gly Ala Glu Glu Val Leu Leu Leu Ala Arg Arg Thr Asp Leu		335
	340	345
Arg Arg Ile Ser Leu Asp Thr Pro Asp Phe Thr Asp Ile Val Leu Gln		350
	355	360
Val Asp Asp Ile Arg His Ala Ile Ala Ile Asp Tyr Asp Pro Leu Glu		365
	370	375
Gly Tyr Val Tyr Trp Thr Asp Asp Glu Val Arg Ala Ile Arg Arg Ala		380
385	390	395
Tyr Leu Asp Gly Ser Gly Ala Gln Thr Leu Val Asn Thr Glu Ile Asn		400
	405	410
Asp Pro Asp Gly Ile Ala Val Asp Trp Val Ala Arg Asn Leu Tyr Trp		415
	420	425
Thr Asp Thr Gly Thr Asp Arg Ile Glu Val Thr Arg Leu Asn Gly Thr		430
	435	440
Ser Arg Lys Ile Leu Val Ser Glu Asp Leu Asp Glu Pro Arg Ala Ile		445
	450	455
Ala Leu His Pro Val Met Gly Leu Met Tyr Trp Thr Asp Trp Gly Glu		460
465	470	475
Asn Pro Lys Ile Glu Cys Ala Asn Leu Asp Gly Gln Glu Arg Arg Val		480
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Leu Val Asn Ala Ser Leu Gly Trp Pro Asn Gly Leu Ala Leu Asp Leu		495
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Leu	Pro	His	Ile	Phe	Gly	Phe	Thr	Leu	Leu	Gly	Asp	Phe	Ile	Tyr	Trp
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Thr	Asp	Trp	Gln	Arg	Arg	Ser	Ile	Glu	Arg	Val	His	Lys	Val	Lys	Ala
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Ser	Arg	Asp	Val	Ile	Ile	Asp	Gln	Leu	Pro	Asp	Leu	Met	Gly	Leu	Lys
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Ala	Val	Asn	Val	Ala	Lys	Val	Val	Gly	Thr	Asn	Pro	Cys	Ala	Asp	Arg
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Cys	Gly	Cys	Pro	Ile	Gly	Leu	Glu	Leu	Leu	Ser	Asp	Met	Lys	Thr	Cys
625					630					635					640
Ile	Val	Pro	Glu	Ala	Phe	Leu	Val	Phe	Thr	Ser	Arg	Ala	Ala	Ile	His
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Arg	Ile	Ser	Leu	Glu	Thr	Asn	Asn	Asn	Asp	Val	Ala	Ile	Pro	Leu	Thr
			660					665					670		
Gly	Val	Lys	Glu	Ala	Ser	Ala	Leu	Asp	Phe	Asp	Val	Ser	Asn	Asn	His
		675					680					685			
Ile	Tyr	Trp	Thr	Asp	Val	Ser	Leu	Lys	Asn	Ile	Ser	Arg	Ala	Phe	Met
		690				695					700				
Asn	Gly	Ser	Ser	Val	Glu	His	Val	Val	Glu	Phe	Gly	Leu	Asp	Tyr	Pro
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 Asp Val Ser Glu Glu Ala Ile Lys Gln Thr Tyr Leu Asn Gln Thr Gly
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 Ala Ala Val Gln Asn Val Val Ile Ser Gly Leu Val Ser Pro Asp Gly
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 Leu Ala Cys Asp Trp Val Gly Lys Lys Leu Tyr Trp Thr Asp Ser Glu
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 Thr Asn Arg Ile Glu Val Ala Asn Leu Asn Gly Thr Ser Arg Lys Val
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 Leu Phe Trp Gln Asp Leu Asp Gln Pro Lys Ala Ile Ala Leu Asp Pro
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 Ala His Gly Tyr Met Tyr Trp Thr Asp Trp Val Glu Thr Pro Arg Ile
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 Glu Arg Ala Gly Met Asp Gly Ser Thr Arg Lys Ile Ile Val Asp Ser
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Phe	Ala	Leu	Thr	Leu	Ser	Gly	Asp	Thr	Leu	Tyr	Trp	Thr	Asp	Trp	Gln
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Cys	Ala	Cys	Pro	Thr	Gly	Val	Gln	Met	Gln	Asp	Asn	Gly	Arg	Thr	Cys
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Lys	Ala	Gly	Ala	Glu	Glu	Val	Leu	Leu	Leu	Ala	Arg	Arg	Thr	Asp	Leu
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Tyr	Leu	Asp	Gly	Ser	Gly	Ala	Gln	Thr	Leu	Val	Asn	Thr	Glu	Ile	Asn
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Phe	Gly	Leu	Thr	Gln	Tyr	Ser	Asp	Tyr	Ile	Tyr	Trp	Thr	Asp	Trp	Asn	
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<212> DNA

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<212> DNA

<213> Homo sapiens

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<211> 221

<212> DNA

<213> Homo sapiens

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<400> 48

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<212> DNA

<213> Homo sapiens

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<212> DNA

<213> Homo sapiens

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<211> 352

<212> DNA

<213> Homo sapiens

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<212> DNA

<213> Homo sapiens

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<210> 62
<211> 496
<212> DNA
<213> Homo sapiens

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<400> 62

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<210> 63

<211> 3081

<212> DNA

<213> Homo sapiens

<400> 63

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<211> 1324

<212> DNA

<213> Homo sapiens

<220>

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<222> (1)...(1324)

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<400> 64

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<210> 65

<211> 2377

<212> DNA

<213> Homo sapiens

<400> 65

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<210> 66

<211> 1295

<212> DNA

<213> Homo sapiens

<400> 66

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<211> 3411

<212> DNA

<213> Homo sapiens

<400> 67

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<212> DNA

<213> Homo sapiens

<400> 68

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<211> 3513

<212> DNA

<213> Homo sapiens

<400> 69

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<211> 3597

<212> DNA

<213> Homo sapiens

<400> 70

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<211> 855

<212> DNA

<213> Homo sapiens

<400> 71

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<211> 3791

<212> DNA

<213> Homo sapiens

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<211> 1683

<212> DNA

<213> Homo sapiens

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<211> 1696
 <212> DNA
 <213> Homo sapiens

<400> 74

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 <211> 7680
 <212> DNA
 <213> Homo sapiens

<400> 75

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 <212> DNA
 <213> Homo sapiens

<400> 76

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<400> 77

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<212> DNA

<213> Homo sapiens

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<211> 2346

<212> DNA

<213> Homo sapiens

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<212> DNA

<213> Homo sapiens

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<213> Homo sapiens

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<212> DNA

<213> Homo sapiens

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			805						810					815			
Ser	Arg	Glu	Thr	Ala	Asp	Thr	Asp	Thr	Ala	Asp	Gln	Val	Met	Ala	Ser		
		820						825					830				
Phe	Lys	Ile	Leu	Ala	Gly	Asp	Lys	Asn	Tyr	Ile	Thr	Met	Asp	Glu	Leu		
	835						840					845					
Arg	Arg	Glu	Leu	Pro	Pro	Asp	Gln	Ala	Glu	Tyr	Cys	Ile	Ala	Arg	Met		
850						855					860						
Ala	Pro	Tyr	Thr	Gly	Pro	Asp	Ser	Val	Pro	Gly	Ala	Leu	Asp	Tyr	Met		
865					870					875					880		
Ser	Phe	Ser	Thr	Ala	Leu	Tyr	Gly	Glu	Ser	Asp	Leu						
			885					890									

<210> 88

<211> 197

<212> PRT

<213> Homo sapiens

<400> 88

Met	Met	Phe	Pro	Gln	Ser	Arg	His	Ser	Gly	Ser	Ser	His	Leu	Pro	Gln		
1				5					10					15			
Gln	Leu	Lys	Phe	Thr	Thr	Ser	Asp	Ser	Cys	Asp	Arg	Ile	Lys	Asp	Glu		
		20						25					30				
Phe	Gln	Leu	Leu	Gln	Ala	Gln	Tyr	His	Ser	Leu	Lys	Leu	Glu	Cys	Asp		
	35						40					45					
Lys	Leu	Ala	Ser	Glu	Lys	Ser	Glu	Met	Gln	Arg	His	Tyr	Val	Met	Tyr		
50						55				60							
Tyr	Glu	Met	Ser	Tyr	Gly	Leu	Asn	Ile	Glu	Met	His	Lys	Gln	Ala	Glu		
65					70					75					80		
Ile	Val	Lys	Arg	Leu	Asn	Gly	Ile	Cys	Ala	Gln	Val	Leu	Pro	Tyr	Leu		
			85					90					95				
Ser	Gln	Glu	His	Gln	Gln	Gln	Val	Leu	Gly	Ala	Ile	Glu	Arg	Ala	Lys		
			100					105					110				
Gln	Val	Thr	Ala	Pro	Glu	Leu	Asn	Ser	Ile	Ile	Arg	Gln	Gln	Leu	Gln		
	115						120					125					
Ala	His	Gln	Leu	Ser	Gln	Leu	Gln	Ala	Leu	Ala	Leu	Pro	Leu	Thr	Pro		
130						135					140						
Leu	Pro	Val	Gly	Leu	Gln	Pro	Pro	Ser	Leu	Pro	Ala	Val	Ser	Ala	Gly		
145					150					155					160		
Thr	Gly	Leu	Leu	Ser	Leu	Ser	Ala	Leu	Gly	Ser	Gln	Ala	His	Leu	Ser		
			165					170					175				
Lys	Glu	Asp	Lys	Asn	Gly	His	Asp	Gly	Asp	Thr	His	Gln	Glu	Asp	Asp		
		180						185					190				
Gly	Glu	Lys	Ser	Asp													
	195																

<210> 89

<211> 739

<212> PRT

<213> Homo sapiens

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<400> 89

Gly	Asp	Lys	Glu	Pro	Thr	Glu	Thr	Ile	Gly	Asp	Leu	Ser	Ile	Cys	Leu
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Asp	Gly	Leu	Gln	Leu	Glu	Ser	Glu	Val	Val	Thr	Asn	Gly	Glu	Thr	Thr
		20						25					30		
Cys	Ser	Glu	Ser	Ala	Ser	Gln	Asn	Asp	Asp	Gly	Ser	Arg	Ser	Lys	Asp
		35					40					45			
Glu	Thr	Arg	Val	Ser	Thr	Asn	Gly	Ser	Asp	Asp	Pro	Glu	Asp	Ala	Gly
	50					55					60				
Ala	Gly	Glu	Asn	Arg	Arg	Val	Ser	Gly	Asn	Asn	Ser	Pro	Ser	Leu	Ser
65					70				75						80
Asn	Gly	Gly	Phe	Lys	Pro	Ser	Arg	Pro	Pro	Arg	Pro	Ser	Arg	Pro	Pro
				85					90					95	
Pro	Pro	Thr	Pro	Arg	Arg	Pro	Ala	Ser	Val	Asn	Gly	Ser	Pro	Ser	Ala
			100					105						110	
Thr	Ser	Glu	Ser	Asp	Gly	Ser	Ser	Thr	Gly	Ser	Leu	Pro	Pro	Thr	Asn
		115					120						125		
Thr	Asn	Thr	Asn	Thr	Ser	Glu	Gly	Ala	Thr	Ser	Gly	Leu	Ile	Ile	Pro
	130					135					140				
Leu	Thr	Ile	Ser	Gly	Gly	Ser	Gly	Pro	Arg	Pro	Leu	Asn	Pro	Val	Thr
145					150					155					160
Gln	Ala	Pro	Leu	Pro	Pro	Gly	Trp	Glu	Gln	Arg	Val	Asp	Gln	His	Gly
				165					170					175	
Arg	Val	Tyr	Tyr	Val	Asp	His	Val	Glu	Lys	Arg	Thr	Thr	Trp	Asp	Arg
			180					185					190		
Pro	Glu	Pro	Leu	Pro	Pro	Gly	Trp	Glu	Arg	Arg	Val	Asp	Asn	Met	Gly
		195				200						205			
Arg	Ile	Tyr	Tyr	Val	Asp	His	Phe	Thr	Arg	Thr	Thr	Thr	Trp	Gln	Arg
	210				215						220				
Pro	Thr	Leu	Glu	Ser	Val	Arg	Asn	Tyr	Glu	Gln	Trp	Gln	Leu	Gln	Arg
225					230					235					240
Ser	Gln	Leu	Gln	Gly	Ala	Met	Gln	Gln	Phe	Asn	Gln	Arg	Phe	Ile	Tyr
			245						250					255	
Gly	Asn	Gln	Asp	Leu	Phe	Ala	Thr	Ser	Gln	Ser	Lys	Glu	Phe	Asp	Pro
			260					265					270		
Leu	Gly	Pro	Leu	Pro	Pro	Gly	Trp	Glu	Lys	Arg	Thr	Asp	Ser	Asn	Gly
		275				280						285			
Arg	Val	Tyr	Phe	Val	Asn	His	Asn	Thr	Arg	Ile	Thr	Gln	Trp	Glu	Asp
	290				295						300				
Pro	Arg	Ser	Gln	Gly	Gln	Leu	Asn	Glu	Lys	Pro	Leu	Pro	Glu	Gly	Trp
305					310					315					320
Glu	Met	Arg	Phe	Thr	Val	Asp	Gly	Ile	Pro	Tyr	Phe	Val	Asp	His	Asn
				325					330					335	
Arg	Arg	Thr	Thr	Thr	Tyr	Ile	Asp	Pro	Arg	Thr	Gly	Lys	Ser	Ala	Leu
			340					345						350	
Asp	Asn	Gly	Pro	Gln	Ile	Ala	Tyr	Val	Arg	Asp	Phe	Lys	Ala	Lys	Val
		355				360						365			
Gln	Tyr	Phe	Arg	Phe	Trp	Cys	Gln	Gln	Leu	Ala	Met	Pro	Gln	His	Ile
	370					375					380				
Lys	Ile	Thr	Val	Thr	Arg	Lys	Thr	Leu	Phe	Glu	Asp	Ser	Phe	Gln	Gln
385					390					395					400
Ile	Met	Ser	Phe	Ser	Pro	Gln	Asp	Leu	Arg	Arg	Arg	Leu	Trp	Val	Ile
				405					410					415	
Phe	Pro	Gly	Glu	Glu	Gly	Leu	Asp	Tyr	Gly	Gly	Val	Ala	Arg	Glu	Trp
		420						425					430		
Phe	Phe	Leu	Leu	Ser	His	Glu	Val	Leu	Asn	Pro	Met	Tyr	Cys	Leu	Phe

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435					440					445					
Glu	Tyr	Ala	Gly	Lys	Asp	Asn	Tyr	Cys	Leu	Gln	Ile	Asn	Pro	Ala	Ser
450					455					460					
Tyr	Ile	Asn	Pro	Asp	His	Leu	Lys	Tyr	Phe	Arg	Phe	Ile	Gly	Arg	Phe
465					470					475					480
Ile	Ala	Met	Ala	Leu	Phe	His	Gly	Lys	Phe	Ile	Asp	Thr	Gly	Phe	Ser
485					490					495					
Leu	Pro	Phe	Tyr	Lys	Arg	Ile	Leu	Asn	Lys	Pro	Val	Gly	Leu	Lys	Asp
500					505					510					
Leu	Glu	Ser	Ile	Asp	Pro	Glu	Phe	Tyr	Asn	Ser	Leu	Ile	Trp	Val	Lys
515					520					525					
Glu	Asn	Asn	Ile	Glu	Glu	Cys	Asp	Leu	Glu	Met	Tyr	Phe	Ser	Val	Asp
530					535					540					
Lys	Glu	Ile	Leu	Gly	Glu	Ile	Lys	Ser	His	Asp	Leu	Lys	Pro	Asn	Gly
545					550					555					560
Gly	Asn	Ile	Leu	Val	Thr	Glu	Glu	Asn	Lys	Glu	Glu	Tyr	Ile	Arg	Met
565					570					575					
Val	Ala	Glu	Trp	Arg	Leu	Ser	Arg	Gly	Val	Glu	Glu	Gln	Thr	Gln	Ala
580					585					590					
Phe	Phe	Glu	Gly	Phe	Asn	Glu	Ile	Leu	Pro	Gln	Gln	Tyr	Leu	Gln	Tyr
595					600					605					
Phe	Asp	Ala	Lys	Glu	Leu	Glu	Val	Leu	Leu	Cys	Gly	Met	Gln	Glu	Ile
610					615					620					
Asp	Leu	Asn	Asp	Trp	Gln	Arg	His	Ala	Ile	Tyr	Arg	His	Tyr	Ala	Arg
625					630					635					640
Thr	Ser	Lys	Gln	Ile	Met	Trp	Phe	Trp	Gln	Phe	Val	Lys	Glu	Ile	Asp
645					650					655					
Asn	Glu	Lys	Arg	Met	Arg	Leu	Leu	Gln	Phe	Val	Thr	Gly	Thr	Cys	Arg
660					665					670					
Leu	Pro	Val	Gly	Gly	Phe	Ala	Asp	Leu	Met	Gly	Ser	Asn	Gly	Pro	Gln
675					680					685					
Lys	Phe	Cys	Ile	Glu	Lys	Val	Gly	Lys	Glu	Asn	Trp	Leu	Pro	Arg	Ser
690					695					700					
His	Thr	Cys	Phe	Asn	Arg	Leu	Asp	Leu	Pro	Pro	Tyr	Lys	Ser	Tyr	Glu
705					710					715					720
Gln	Leu	Lys	Glu	Lys	Leu	Leu	Phe	Ala	Ile	Glu	Glu	Thr	Glu	Gly	Phe
725					730					735					

Gly Gln Glu

<210> 90
 <211> 431
 <212> PRT
 <213> Homo sapiens

<400> 90

Gly	Pro	Pro	Pro	Thr	Arg	Ala	Leu	Pro	Leu	Pro	Gln	Ser	Leu	Pro	Pro
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Asp	Phe	Arg	Leu	Glu	Pro	Thr	Ala	Pro	Ala	Leu	Ser	Pro	Arg	Ser	Ser
			20					25					30		
Phe	Ala	Ser	Ser	Ser	Ala	Ser	Asp	Ala	Ser	Lys	Pro	Ser	Ser	Pro	Arg
			35				40					45			
Gly	Ser	Leu	Leu	Leu	Asp	Gly	Ala	Gly	Ala	Gly	Gly	Ala	Gly	Gly	Ser
			50			55				60					
Arg	Pro	Cys	Ser	Asn	Arg	Thr	Ser	Gly	Ile	Ser	Met	Gly	Tyr	Asp	Gln
65				70					75					80	
Arg	His	Gly	Ser	Pro	Leu	Pro	Ala	Gly	Pro	Cys	Leu	Phe	Gly	Pro	Pro

85																90				95			
Leu	Ala	Gly	Ala	Pro	Ala	Gly	Tyr	Ser	Pro	Gly	Gly	Val	Pro	Ser	Ala								
100								105				110											
Tyr	Pro	Glu	Leu	His	Ala	Ala	Leu	Asp	Arg	Leu	Tyr	Ala	Gln	Arg	Pro								
115								120				125											
Ala	Gly	Phe	Gly	Cys	Gln	Glu	Ser	Arg	His	Ser	Tyr	Pro	Pro	Ala	Leu								
130								135				140											
Gly	Ser	Pro	Gly	Ala	Leu	Ala	Gly	Ala	Arg	Val	Gly	Ala	Ala	Gly	Pro								
145								150				155											
Leu	Glu	Arg	Arg	Gly	Ala	Gln	Pro	Gly	Arg	His	Ser	Val	Thr	Gly	Tyr								
				165								170											
Gly	Asp	Cys	Ala	Val	Gly	Ala	Arg	Tyr	Gln	Asp	Glu	Leu	Thr	Ala	Leu								
				180								185											
Leu	Arg	Leu	Thr	Val	Gly	Thr	Gly	Gly	Arg	Glu	Ala	Gly	Ala	Arg	Gly								
				195								200											
Glu	Pro	Ser	Gly	Ile	Glu	Pro	Ser	Gly	Leu	Glu	Glu	Pro	Pro	Gly	Pro								
				210								215											
Phe	Val	Pro	Glu	Ala	Ala	Arg	Ala	Arg	Met	Arg	Glu	Pro	Glu	Ala	Arg								
225												230											
Glu	Asp	Tyr	Phe	Gly	Thr	Cys	Ile	Lys	Cys	Asn	Lys	Gly	Ile	Tyr	Gly								
				245								250											
Gln	Ser	Asn	Ala	Cys	Gln	Ala	Leu	Asp	Ser	Leu	Tyr	His	Thr	Gln	Cys								
				260								265											
Phe	Val	Cys	Cys	Ser	Cys	Gly	Arg	Thr	Leu	Arg	Cys	Lys	Ala	Phe	Tyr								
				275								280											
Ser	Val	Asn	Gly	Ser	Val	Tyr	Cys	Glu	Glu	Asp	Tyr	Leu	Phe	Ser	Gly								
				290								295											
Phe	Gln	Glu	Ala	Ala	Glu	Lys	Cys	Cys	Val	Cys	Gly	His	Leu	Ile	Leu								
305												310											
Glu	Lys	Ile	Leu	Gln	Ala	Met	Gly	Lys	Ser	Tyr	His	Pro	Gly	Cys	Phe								
				325								330											
Arg	Cys	Ile	Val	Cys	Asn	Lys	Cys	Leu	Asp	Gly	Ile	Pro	Phe	Thr	Val								
				340								345											
Asp	Phe	Ser	Asn	Gln	Val	Tyr	Cys	Val	Thr	Asp	Tyr	His	Lys	Asn	Tyr								
				355								360											
Ala	Pro	Lys	Cys	Ala	Ala	Cys	Gly	Gln	Pro	Ile	Leu	Pro	Ser	Glu	Gly								
				370								375											
Cys	Glu	Asp	Ile	Val	Arg	Val	Ile	Ser	Met	Asp	Arg	Asp	Tyr	His	Phe								
385												390											
Glu	Cys	Tyr	His	Cys	Glu	Asp	Cys	Arg	Met	Gln	Leu	Ser	Asp	Glu	Glu								
				405								410											
Gly	Cys	Cys	Cys	Phe	Pro	Leu	Asp	Gly	His	Leu	Leu	Cys	His	Gly									
				420								425											
																430							

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<400> 91
Gly Pro Gly Ser Arg His His Arg Ala Arg Asp Arg Leu Ile His Phe
 1          5          10          15
Gly Ala Val Ser Thr Asp Val Leu Gly Cys Ser Ala His Cys Ser Leu
          20          25          30
Thr Gln Ser Pro Lys Met Asn Ile Gln Glu Gln Gly Phe Pro Leu Asp
          35          40          45
Leu Gly Ala Ser Phe Thr Glu Asp Ala Pro Arg Pro Pro Val Pro Gly

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50	55	60
Glu Glu Gly Glu Leu	Val Ser Thr Asp Pro Arg	Pro Ala Ser Tyr Ser
65	70	75
Phe Cys Ser Gly Lys	Gly Val Gly Ile Lys	Gly Glu Thr Ser Thr Ala
85	90	95
Thr Pro Arg Arg Ser	Asp Leu Asp Leu Gly Tyr Glu	Pro Glu Gly Ser
100	105	110
Ala Ser Pro Thr Pro	Pro Tyr Leu Lys Trp Ala	Glu Ser Leu His Ser
115	120	125
Leu Leu Asp Asp Gln	Asp Gly Ile Ser Leu Phe	Arg Thr Phe Leu Lys
130	135	140
Gln Glu Gly Cys Ala	Asp Leu Leu Asp Phe Trp	Phe Ala Cys Thr Gly
145	150	155
Phe Arg Lys Leu Glu	Pro Cys Asp Ser Asn	Glu Glu Lys Arg Leu Lys
165	170	175
Leu Ala Arg Ala Ile	Tyr Arg Lys Tyr Ile	Leu Asp Asn Asn Gly Ile
180	185	190
Val Ser Arg Gln Thr	Lys Pro Ala Thr Lys Ser	Phe Ile Lys Gly Cys
195	200	205
Ile Met Lys Gln Leu	Ile Asp Pro Ala Met Phe	Asp Gln Ala Gln Thr
210	215	220
Glu Ile Gln Ala Thr	Met Glu Glu Asn Thr Tyr	Pro Ser Phe Leu Lys
225	230	235
Ser Asp Ile Tyr Leu	Glu Tyr Thr Arg Thr	Gly Ser Glu Ser Pro Lys
245	250	255
Val Cys Ser Asp Gln	Ser Ser Gly Ser Gly Thr	Gly Lys Gly Ile Ser
260	265	270
Gly Tyr Leu Pro Thr	Leu Asn Glu Asp Glu Glu	Trp Lys Cys Asp Gln
275	280	285
Asp Met Asp Glu Asp	Asp Gly Arg Asp Ala Ala	Pro Pro Gly Arg Leu
290	295	300
Pro Gln Lys Leu Leu	Leu Glu Thr Ala Ala	Pro Arg Val Ser Ser Ser
305	310	315
Arg Arg Tyr Ser Glu	Gly Arg Glu Phe Arg	Tyr Gly Ser Trp Arg Glu
325	330	335
Pro Val Asn Pro Tyr	Tyr Val Asn Ala Gly Tyr	Ala Leu Ala Pro Ala
340	345	350
Thr Ser Ala Asn Asp	Ser Glu Gln Gln Ser Leu	Ser Ser Asp Ala Asp
355	360	365
Thr Leu Ser Leu Thr	Asp Ser Ser Val Asp	Gly Ile Pro Pro Tyr Arg
370	375	380
Ile Arg Lys Gln His	Arg Arg Glu Met Gln	Glu Ser Ala Gln Val Asn
385	390	395
Gly Arg Val Pro Leu	Pro His Ile Pro Arg	Thr Tyr Arg Val Pro Lys
405	410	415
Glu Val Arg Val Glu	Pro Gln Lys Phe Ala	Glu Glu Leu Ile His Arg
420	425	430
Leu Glu Ala Val Gln	Arg Thr Arg Glu Ala	Glu Glu Lys Leu Glu Glu
435	440	445
Arg Leu Lys Arg Val	Arg Met Glu Glu Glu	Gly Glu Asp Gly Asp Pro
450	455	460
Ser Ser Gly Pro Pro	Gly Pro Cys His Lys	Leu Pro Pro Ala Pro Ala
465	470	475
Trp His His Phe Pro	Pro Arg Leu Cys Trp	Thr Trp Ala Cys Ala Gly
485	490	495
Leu Arg Asp Ala His	Glu Glu Asn Pro Glu	Ser Ile Leu Asp Glu His
500	505	510

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Val	Gln	Arg	Val	Leu	Arg	Thr	Thr	Gly	Arg	Gln	Ser	Pro	Gly	Pro	Gly		
	515						520					525					
His	Arg	Ser	Pro	Asp	Ser	Gly	His	Val	Ala	Lys	Met	Pro	Val	Ala	Leu		
	530					535					540						
Gly	Gly	Ala	Ala	Ser	Gly	His	Gly	Lys	His	Val	Pro	Lys	Ser	Gly	Ala		
545					550					555					560		
Lys	Leu	Asp	Ala	Ala	Gly	Leu	His	His	His	Arg	His	Val	His	His	His		
				565					570					575			
Val	His	His	Ser	Thr	Ala	Arg	Pro	Lys	Glu	Gln	Val	Glu	Ala	Glu	Ala		
			580					585					590				
Thr	Arg	Arg	Ala	Gln	Ser	Ser	Phe	Ala	Trp	Gly	Leu	Glu	Pro	His	Ser		
	595					600						605					
His	Gly	Ala	Arg	Ser	Arg	Gly	Tyr	Ser	Glu	Ser	Val	Gly	Ala	Ala	Pro		
610					615						620						
Asn	Ala	Ser	Asp	Gly	Leu	Ala	His	Ser	Gly	Lys	Val	Gly	Val	Ala	Cys		
625				630						635					640		
Lys	Arg	Asn	Ala	Lys	Lys	Ala	Glu	Ser	Gly	Lys	Ser	Ala	Ser	Thr	Glu		
			645						650					655			
Val	Pro	Gly	Ala	Ser	Glu	Asp	Ala	Glu	Lys	Asn	Gln	Lys	Ile	Met	Gln		
		660						665					670				
Trp	Ile	Ile	Glu	Gly	Glu	Lys	Glu	Ile	Ser	Arg	His	Arg	Arg	Thr	Gly		
	675						680					685					
His	Gly	Ser	Ser	Gly	Thr	Arg	Lys	Pro	Gln	Pro	His	Glu	Asn	Ser	Arg		
690					695						700						
Pro	Leu	Ser	Leu	Glu	His	Pro	Trp	Ala	Gly	Pro	Gln	Leu	Arg	Thr	Ser		
705				710					715						720		
Val	Gln	Pro	Ser	His	Leu	Phe	Ile	Gln	Asp	Pro	Thr	Met	Pro	Pro	His		
			725					730						735			
Pro	Ala	Pro	Asn	Pro	Leu	Thr	Gln	Leu	Glu	Glu	Ala	Arg	Arg	Arg	Leu		
			740				745						750				
Glu	Glu	Glu	Glu	Lys	Arg	Ala	Ser	Arg	Ala	Pro	Ser	Lys	Gln	Arg	Tyr		
	755						760					765					
Val	Gln	Glu	Val	Met	Arg	Arg	Gly	Arg	Ala	Cys	Val	Arg	Pro	Ala	Cys		
770					775						780						
Ala	Pro	Val	Leu	His	Val	Val	Pro	Ala	Val	Ser	Asp	Met	Glu	Leu	Ser		
785				790						795					800		
Glu	Thr	Glu	Thr	Arg	Ser	Gln	Arg	Lys	Val	Gly	Gly	Gly	Ser	Ala	Gln		
			805						810					815			
Pro	Cys	Asp	Ser	Ile	Val	Val	Ala	Tyr	Tyr	Phe	Cys	Gly	Glu	Pro	Ile		
		820						825					830				
Pro	Tyr	Arg	Thr	Leu	Val	Arg	Gly	Arg	Ala	Val	Thr	Leu	Gly	Gln	Phe		
	835						840					845					
Lys	Glu	Leu	Leu	Thr	Lys	Lys	Gly	Ser	Tyr	Arg	Tyr	Tyr	Phe	Lys	Lys		
	850				855						860						
Val	Ser	Asp	Glu	Phe	Asp	Cys	Gly	Val	Val	Phe	Glu	Glu	Val	Arg	Glu		
865				870						875					880		
Asp	Glu	Ala	Val	Leu	Pro	Val	Phe	Glu	Glu	Lys	Ile	Ile	Gly	Lys	Val		
			885					890						895			
Glu	Lys	Val	Asp														
			900														

<210> 92

<211> 591

<212> PRT

<213> Homo sapiens

<400> 92

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Met	Val	Pro	Val	Ala	Val	Thr	Ala	Ala	Val	Ala	Pro	Val	Leu	Ser	Ile
1				5					10					15	
Asn	Ser	Asp	Phe	Ser	Asp	Leu	Arg	Glu	Ile	Lys	Lys	Gln	Leu	Leu	Leu
			20					25					30		
Ile	Ala	Gly	Leu	Thr	Arg	Glu	Arg	Gly	Leu	Leu	His	Ser	Ser	Lys	Trp
		35					40					45			
Ser	Ala	Glu	Leu	Ala	Phe	Ser	Leu	Pro	Ala	Leu	Pro	Leu	Ala	Glu	Leu
	50					55					60				
Gln	Pro	Pro	Pro	Pro	Ile	Thr	Glu	Glu	Asp	Ala	Gln	Asp	Met	Asp	Ala
65					70				75						80
Tyr	Thr	Leu	Ala	Lys	Ala	Tyr	Phe	Asp	Val	Lys	Glu	Tyr	Asp	Arg	Ala
				85					90					95	
Ala	His	Phe	Leu	His	Gly	Cys	Asn	Ser	Lys	Lys	Ala	Tyr	Phe	Leu	Tyr
			100					105					110		
Met	Tyr	Ser	Arg	Tyr	Leu	Ser	Gly	Glu	Lys	Lys	Lys	Asp	Asp	Glu	Thr
		115					120					125			
Val	Asp	Ser	Leu	Gly	Pro	Leu	Glu	Lys	Gly	Gln	Val	Lys	Asn	Glu	Ala
	130					135					140				
Leu	Arg	Glu	Leu	Arg	Val	Glu	Leu	Ser	Lys	Lys	His	Gln	Ala	Arg	Glu
145					150						155				160
Leu	Asp	Gly	Phe	Gly	Leu	Tyr	Leu	Tyr	Gly	Val	Val	Leu	Arg	Lys	Leu
				165					170					175	
Asp	Leu	Val	Lys	Glu	Ala	Ile	Asp	Val	Phe	Val	Glu	Ala	Thr	His	Val
			180					185						190	
Leu	Pro	Leu	His	Trp	Gly	Ala	Trp	Leu	Glu	Leu	Cys	Asn	Leu	Ile	Thr
		195					200					205			
Asp	Lys	Glu	Met	Leu	Lys	Phe	Leu	Ser	Leu	Pro	Asp	Thr	Trp	Met	Lys
	210					215					220				
Glu	Phe	Phe	Leu	Ala	His	Ile	Tyr	Thr	Glu	Leu	Gln	Leu	Ile	Glu	Glu
225					230						235				240
Ala	Leu	Gln	Lys	Tyr	Gln	Asn	Leu	Ile	Asp	Val	Gly	Phe	Ser	Lys	Ser
				245					250					255	
Ser	Tyr	Ile	Val	Ser	Gln	Ile	Ala	Val	Ala	Tyr	His	Asn	Ile	Arg	Asp
			260					265						270	
Ile	Asp	Lys	Ala	Leu	Ser	Ile	Phe	Asn	Glu	Leu	Arg	Lys	Gln	Asp	Pro
		275					280						285		
Tyr	Arg	Ile	Glu	Asn	Met	Asp	Thr	Phe	Ser	Asn	Leu	Leu	Tyr	Val	Arg
	290					295					300				
Ser	Met	Lys	Ser	Glu	Leu	Ser	Tyr	Leu	Ala	His	Asn	Leu	Cys	Glu	Ile
305					310					315					320
Asp	Lys	Tyr	Arg	Val	Glu	Thr	Cys	Cys	Val	Ile	Gly	Asn	Tyr	Tyr	Ser
				325					330					335	
Leu	Arg	Ser	Gln	His	Glu	Lys	Ala	Ala	Leu	Tyr	Phe	Gln	Arg	Ala	Leu
				340				345						350	
Lys	Leu	Asn	Pro	Arg	Tyr	Leu	Gly	Ala	Trp	Thr	Leu	Met	Gly	His	Glu
		355					360					365			
Tyr	Met	Glu	Met	Lys	Asn	Thr	Ser	Ala	Ala	Ile	Gln	Ala	Tyr	Arg	His
	370					375						380			
Ala	Ile	Glu	Val	Asn	Lys	Arg	Asp	Tyr	Arg	Ala	Trp	Tyr	Gly	Leu	Gly
385					390					395					400
Gln	Thr	Tyr	Glu	Ile	Leu	Lys	Met	Pro	Phe	Tyr	Cys	Leu	Tyr	Tyr	Tyr
				405					410					415	
Arg	Arg	Ala	His	Gln	Leu	Arg	Pro	Asn	Asp	Ser	Arg	Met	Leu	Val	Ala
			420					425					430		
Leu	Gly	Glu	Cys	Tyr	Glu	Lys	Leu	Asn	Gln	Leu	Val	Glu	Ala	Lys	Lys
		435					440					445			
Cys	Tyr	Trp	Arg	Ala	Tyr	Ala	Val	Gly	Asp	Val	Glu	Lys	Met	Ala	Leu

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450	455	460
Val Lys Leu Ala Lys Leu	His Glu Gln Leu Thr	Glu Ser Glu Gln Ala
465	470	475
Ala Gln Cys Tyr Ile Lys	Tyr Ile Gln Asp Ile	Tyr Ser Cys Gly Glu
485	490	495
Ile Val Glu His Leu Glu	Glu Ser Thr Ala Phe	Arg Tyr Leu Ala Gln
500	505	510
Tyr Tyr Phe Lys Cys Lys	Leu Trp Asp Glu Ala	Ser Thr Cys Ala Gln
515	520	525
Lys Cys Cys Ala Phe Asn	Asp Thr Arg Glu Glu	Gly Lys Ala Leu Leu
530	535	540
Arg Gln Ile Leu Gln Leu	Arg Asn Gln Gly Glu	Thr Pro Thr Thr Glu
545	550	555
Val Pro Ala Pro Phe Phe	Leu Pro Ala Ser Leu	Ser Ala Asn Asn Thr
565	570	575
Pro Thr Arg Arg Val Ser	Pro Leu Asn Leu Ser	Ser Val Thr Pro
580	585	590

<210> 93

<211> 914

<212> PRT

<213> Homo sapiens

<400> 93

Val Tyr Gln Val Leu Leu	Val Gly Ser Thr Leu Leu	Lys Glu Val Pro
1	5	10
Ser Gly Leu Gln Leu Glu	Gln Leu Pro Ser Gln Ser	Leu Leu Thr His
20	25	30
Ile Pro Thr Ala Gly Leu	Pro Thr Ser Leu Gly Gly	Gly Leu Pro Tyr
35	40	45
Cys His Gln Ala Trp Leu	Asp Phe Arg Arg Arg	Leu Glu Ala Leu Leu
50	55	60
Gln Asn Cys Gln Ala Ala	Cys Ala Leu Leu Gln	Gly Ala Ile Glu Ser
65	70	75
Val Lys Ala Val Pro Gln	Pro Met Glu Pro Gly	Glu Val Gly Gln Leu
85	90	95
Leu Gln Gln Thr Glu Val	Leu Met Gln Gln Val	Leu Asp Ser Pro Trp
100	105	110
Leu Ala Trp Leu Gln Cys	Gln Gly Gly Arg Glu	Leu Thr Trp Leu Lys
115	120	125
Gln Glu Val Pro Glu Val	Thr Leu Ser Pro Asp	Tyr Arg Thr Ala Met
130	135	140
Asp Lys Ala Asp Glu Leu	Tyr Asp Arg Val Asp	Gly Leu Leu His Gln
145	150	155
Leu Thr Leu Gln Ser Asn	Gln Arg Ile Gln Ala	Leu Glu Leu Val Gln
165	170	175
Thr Leu Glu Ala Arg Glu	Ser Gly Leu His Gln	Ile Glu Val Trp Leu
180	185	190
Gln Gln Val Gly Trp Pro	Ala Leu Glu Glu Ala	Gly Glu Pro Ser Leu
195	200	205
Asp Met Leu Leu Gln Ala	Gln Gly Ser Phe Gln	Glu Leu Tyr Gln Val
210	215	220
Ala Gln Glu Gln Val Arg	Gln Gly Glu Lys Phe	Leu Gln Pro Leu Thr
225	230	235
Gly Trp Glu Ala Ala Glu	Leu Asp Pro Pro Gly	Ala Arg Phe Leu Ala
245	250	255
Leu Arg Ala Gln Leu Thr	Glu Phe Ser Arg Ala	Leu Ala Gln Arg Cys

			260				265				270				
Gln	Arg	Leu	Ala	Asp	Ala	Glu	Arg	Leu	Phe	Gln	Leu	Phe	Arg	Glu	Ala
			275				280				285				
Leu	Thr	Trp	Ala	Glu	Glu	Gly	Gln	Arg	Val	Leu	Ala	Glu	Leu	Glu	Gln
			290				295				300				
Glu	Arg	Pro	Gly	Val	Val	Leu	Gln	Gln	Leu	Gln	Leu	His	Trp	Thr	Arg
305				310						315			320		
His	Pro	Asp	Leu	Pro	Pro	Ala	His	Phe	Arg	Lys	Met	Trp	Ala	Leu	Ala
			325						330			335			
Thr	Gly	Leu	Gly	Ser	Glu	Ala	Ile	Arg	Gln	Glu	Cys	Arg	Trp	Ala	Trp
			340						345			350			
Ala	Arg	Cys	Gln	Asp	Thr	Trp	Leu	Ala	Leu	Asp	Gln	Lys	Leu	Glu	Ala
			355			360			365						
Ser	Leu	Lys	Leu	Pro	Pro	Val	Gly	Ser	Thr	Ala	Ser	Leu	Cys	Val	Ser
370						375			380						
Gln	Val	Pro	Ala	Ala	Pro	Ala	His	Pro	Pro	Leu	Arg	Lys	Ala	Tyr	Ser
385				390						395			400		
Phe	Asp	Arg	Asn	Leu	Gly	Gln	Ser	Leu	Ser	Glu	Pro	Ala	Cys	His	Cys
			405						410			415			
His	His	Ala	Ala	Thr	Ile	Ala	Ala	Cys	Arg	Arg	Pro	Glu	Ala	Gly	Gly
			420			425			430						
Gly	Ala	Leu	Pro	Gln	Ala	Ser	Pro	Thr	Val	Pro	Pro	Pro	Gly	Ser	Ser
			435			440			445						
Asp	Pro	Arg	Ser	Leu	Asn	Arg	Leu	Gln	Leu	Val	Leu	Ala	Glu	Met	Val
450						455			460						
Ala	Thr	Glu	Arg	Glu	Tyr	Val	Arg	Ala	Leu	Glu	Tyr	Thr	Met	Glu	Asn
465				470						475			480		
Tyr	Phe	Pro	Glu	Leu	Asp	Arg	Pro	Asp	Val	Pro	Gln	Gly	Leu	Arg	Gly
			485						490			495			
Gln	Arg	Ala	His	Leu	Phe	Gly	Asn	Leu	Glu	Lys	Leu	Arg	Asp	Phe	His
			500			505			510						
Cys	His	Phe	Phe	Leu	Arg	Glu	Leu	Glu	Ala	Cys	Thr	Arg	His	Pro	Pro
			515			520			525						
Arg	Val	Ala	Tyr	Ala	Phe	Leu	Arg	His	Arg	Val	Gln	Phe	Gly	Met	Tyr
530						535			540						
Ala	Leu	Tyr	Ser	Lys	Asn	Lys	Pro	Arg	Ser	Asp	Ala	Leu	Met	Ser	Ser
545				550			555			560					
Tyr	Gly	His	Thr	Phe	Phe	Lys	Asp	Lys	Gln	Gln	Ala	Leu	Gly	Asp	His
			565						570			575			
Leu	Asp	Leu	Ala	Ser	Tyr	Leu	Leu	Lys	Pro	Ile	Gln	Arg	Met	Gly	Lys
			580			585			590						
Tyr	Ala	Leu	Leu	Leu	Gln	Glu	Leu	Ala	Arg	Ala	Cys	Gly	Gly	Pro	Thr
595						600			605						
Gln	Glu	Leu	Ser	Ala	Leu	Arg	Glu	Ala	Gln	Ser	Leu	Val	His	Phe	Gln
610						615			620						
Leu	Arg	His	Gly	Asn	Asp	Leu	Leu	Ala	Met	Asp	Ala	Ile	Gln	Gly	Cys
625				630			635			640					
Asp	Val	Asn	Leu	Lys	Glu	Gln	Gly	Gln	Leu	Val	Arg	Gln	Asp	Glu	Phe
			645			650			655						
Val	Val	Arg	Thr	Gly	Arg	His	Lys	Ser	Val	Arg	Arg	Ile	Phe	Leu	Phe
			660			665			670						
Glu	Glu	Leu	Leu	Leu	Phe	Ser	Lys	Pro	Arg	His	Gly	Pro			

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Arg Arg Lys Ala Arg Asp Thr Phe Val Leu Gln Ala Ser Ser Leu Ala
 725 730 735
 Ile Lys Gln Ala Trp Thr Ala Asp Ile Ser His Leu Leu Trp Arg Gln
 740 745 750
 Ala Val His Asn Lys Glu Val Arg Met Ala Glu Met Val Ser Met Gly
 755 760 765
 Val Gly Asn Lys Ala Phe Arg Asp Ile Ala Pro Ser Glu Glu Ala Ile
 770 775 780
 Asn Asp Arg Thr Val Asn Tyr Val Leu Lys Cys Arg Glu Val Arg Ser
 785 790 795 800
 Arg Ala Ser Ile Ala Val Ala Pro Phe Asp His Asp Ser Leu Tyr Leu
 805 810 815
 Gly Ala Ser Asn Ser Leu Pro Gly Asp Pro Ala Ser Cys Ser Val Leu
 820 825 830
 Gly Ser Leu Asn Leu His Leu Tyr Arg Asp Pro Ala Leu Leu Gly Leu
 835 840 845
 Arg Cys Pro Leu Tyr Pro Ser Phe Leu Glu Glu Ala Ala Leu Glu Ala
 850 855 860
 Glu Ala Glu Leu Gly Gly Gln Pro Ser Leu Thr Ala Glu Asp Ser Glu
 865 870 875 880
 Ile Ser Ser Gln Cys Pro Ser Ala Ser Gly Ser Ser Gly Ser Asp Ser
 885 890 895
 Ser Cys Val Ser Gly Gln Ala Leu Gly Arg Gly Leu Glu Asp Leu Pro
 900 905 910
 Cys Val

<210> 94
 <211> 277
 <212> PRT
 <213> Homo sapiens

<400> 94
 Leu Asn Tyr Leu Leu Glu Ser Arg Leu Glu Ala Ala Ala His Cys Ala
 1 5 10 15
 Leu Lys Gln Gly Ile Ala Thr Ala Ser Leu Leu Pro Ala Gln Leu Gln
 20 25 30
 Pro Ala Val Leu Thr Val Val Thr Cys His Val Val Val Ser Val His
 35 40 45
 Gly His His Thr Asp Gly Cys Leu Ala Ala Leu Cys Arg Glu Asp Arg
 50 55 60
 Thr Gly Thr Gly Gly Ala Phe Trp Cys Lys Asn Arg Val Ile Val Ser
 65 70 75 80
 His Ala Val Asp Val Val Leu His Val His Gly Glu Gly Asn Pro Val
 85 90 95
 Gln Ala Leu Ile Ala His Gly Ala Pro Glu Ala Ala Trp Val Val Gly
 100 105 110
 Leu Ala Gln Gly Leu Gln Asp His Phe His Asp Glu Met Ser Thr His
 115 120 125
 Ala Ala Phe Val Gly Arg Leu Leu Glu Pro Gly Val Gln Glu Val Leu
 130 135 140
 Leu Ala Val His Phe Leu Thr His Val Val Glu Arg Leu Pro Thr Glu
 145 150 155 160
 Ser Ser Pro Thr Arg Val Ala Gly Glu Ala Val Ser Val Ile Lys Thr
 165 170 175
 Pro His Cys Leu Ala Arg Leu Leu Gly Ser Val Asp Ala Lys Pro Thr
 180 185 190

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Leu Asp Ala Asn Ala Glu Val Val Pro Arg Arg Ala Arg Leu Glu Arg
 195 200 205
 Pro Leu Gln Leu Pro Gly Glu Arg Leu Gln Pro Pro Leu Gly Arg Ala
 210 215 220
 Trp Ala Ala Leu Pro Ala Arg Gly Gln Arg Glu Cys Arg Gln Arg Glu
 225 230 235 240
 Gly Gly Arg Pro Arg Arg Leu Arg Gly Ala Ser Gly Arg Gly Ala Gly
 245 250 255
 Ala Gly Arg Glu Glu Val Ser Val Gly Phe Ser Ala Gln Trp Glu Phe
 260 265 270
 Gly Ser Gly Arg His
 275

<210> 95

<211> 1120

<212> PRT

<213> Homo sapiens

<400> 95

Met Trp Arg Val Lys Lys Leu Ser Leu Ser Leu Ser Pro Ser Pro Gln
 1 5 10 15
 Thr Gly Lys Pro Ser Met Arg Thr Pro Leu Arg Glu Leu Thr Leu Gln
 20 25 30
 Pro Gly Ala Leu Thr Thr Ser Gly Lys Arg Ser Pro Ala Cys Ser Ser
 35 40 45
 Leu Thr Pro Ser Leu Cys Lys Leu Gly Leu Gln Glu Gly Ser Asn Asn
 50 55 60
 Ser Ser Pro Val Asp Phe Val Asn Asn Lys Arg Thr Asp Leu Ser Ser
 65 70 75 80
 Glu His Phe Ser His Ser Ser Lys Trp Leu Glu Thr Cys Gln His Glu
 85 90 95
 Ser Asp Glu Gln Pro Leu Asp Pro Ile Pro Gln Ile Ser Ser Thr Pro
 100 105 110
 Lys Thr Ser Glu Glu Ala Val Asp Pro Leu Gly Asn Tyr Met Val Lys
 115 120 125
 Thr Ile Val Leu Val Pro Ser Pro Leu Gly Gln Gln Gln Asp Met Ile
 130 135 140
 Phe Glu Ala Arg Leu Asp Thr Met Ala Glu Thr Asn Ser Ile Ser Leu
 145 150 155 160
 Asn Gly Pro Leu Arg Thr Asp Asp Leu Val Arg Glu Glu Val Ala Pro
 165 170 175
 Cys Met Gly Asp Arg Phe Ser Glu Val Ala Ala Val Ser Glu Lys Pro
 180 185 190
 Ile Phe Gln Glu Ser Pro Ser His Leu Leu Glu Glu Ser Pro Pro Asn
 195 200 205
 Pro Cys Ser Glu Gln Leu His Cys Ser Lys Glu Ser Leu Ser Ser Arg
 210 215 220
 Thr Glu Ala Val Arg Glu Asp Leu Val Pro Ser Glu Ser Asn Ala Phe
 225 230 235 240
 Leu Pro Ser Ser Val Leu Trp Leu Ser Pro Ser Thr Ala Leu Ala Ala
 245 250 255
 Asp Phe Arg Val Asn His Val Asp Pro Glu Glu Glu Ile Val Glu His
 260 265 270
 Gly Ala Met Glu Glu Arg Glu Met Arg Phe Pro Thr His Pro Lys Glu
 275 280 285
 Ser Glu Thr Glu Asp Gln Ala Leu Val Ser Ser Val Glu Asp Ile Leu
 290 295 300

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Ser	Thr	Cys	Leu	Thr	Pro	Asn	Leu	Val	Glu	Met	Glu	Ser	Gln	Glu	Ala
305					310				315						320
Pro	Gly	Pro	Ala	Val	Glu	Asp	Val	Gly	Arg	Ile	Leu	Gly	Ser	Asp	Thr
				325					330					335	
Glu	Ser	Trp	Met	Ser	Pro	Leu	Ala	Trp	Leu	Glu	Lys	Gly	Val	Asn	Thr
			340					345					350		
Ser	Val	Met	Leu	Glu	Asn	Leu	Arg	Gln	Ser	Leu	Ser	Leu	Pro	Ser	Met
		355					360					365			
Leu	Arg	Asp	Ala	Ala	Ile	Gly	Thr	Thr	Pro	Phe	Ser	Thr	Cys	Ser	Val
	370					375				380					
Gly	Thr	Trp	Phe	Thr	Pro	Ser	Ala	Pro	Gln	Glu	Lys	Ser	Thr	Asn	Thr
385					390					395					400
Ser	Gln	Thr	Gly	Leu	Val	Gly	Thr	Lys	His	Ser	Thr	Ser	Glu	Thr	Glu
			405						410					415	
Gln	Leu	Leu	Cys	Gly	Arg	Pro	Pro	Asp	Leu	Thr	Ala	Leu	Ser	Arg	His
			420					425					430		
Asp	Leu	Glu	Asp	Asn	Leu	Leu	Ser	Ser	Leu	Val	Ile	Val	Glu	Phe	Leu
		435					440					445			
Ser	Arg	Gln	Leu	Arg	Asp	Trp	Lys	Ser	Gln	Leu	Ala	Val	Pro	His	Pro
	450					455					460				
Glu	Thr	Gln	Asp	Ser	Ser	Thr	Gln	Thr	Asp	Thr	Ser	His	Ser	Gly	Ile
465					470				475						480
Thr	Asn	Lys	Leu	Gln	His	Leu	Lys	Glu	Ser	His	Glu	Met	Gly	Gln	Ala
			485						490					495	
Leu	Gln	Gln	Ala	Arg	Asn	Val	Met	Gln	Ser	Trp	Val	Leu	Ile	Ser	Lys
			500					505					510		
Glu	Leu	Ile	Ser	Leu	Leu	His	Leu	Ser	Leu	Leu	His	Leu	Glu	Glu	Asp
		515					520					525			
Lys	Thr	Thr	Val	Asn	Gln	Glu	Ser	Arg	Arg	Ala	Glu	Thr	Leu	Val	Cys
	530					535				540					
Cys	Cys	Phe	Asp	Leu	Leu	Lys	Lys	Leu	Arg	Ala	Lys	Leu	Gln	Ser	Leu
545					550					555					560
Lys	Ala	Glu	Arg	Glu	Glu	Ala	Arg	His	Arg	Glu	Glu	Met	Ala	Leu	Arg
			565						570					575	
Gly	Lys	Asp	Ala	Ala	Glu	Ile	Val	Leu	Glu	Ala	Phe	Cys	Ala	His	Ala
			580					585					590		
Ser	Gln	Arg	Ile	Ser	Gln	Leu	Glu	Gln	Asp	Leu	Ala	Ser	Met	Arg	Glu
		595					600					605			
Phe	Arg	Gly	Leu	Leu	Lys	Asp	Ala	Gln	Thr	Gln	Leu	Val	Gly	Leu	His
	610					615					620				
Ala	Lys	Gln	Glu	Glu	Leu	Val	Gln	Gln	Thr	Val	Ser	Leu	Thr	Ser	Thr
625					630					635					640
Leu	Gln	Gln	Asp	Trp	Arg	Ser	Met	Gln	Leu	Asp	Tyr	Thr	Thr	Trp	Thr
			645						650					655	
Ala	Leu	Leu	Ser	Arg	Ser	Arg	Gln	Leu	Thr	Glu	Lys	Leu	Thr	Val	Lys
			660					665					670		
Ser	Gln	Gln	Ala	Leu	Gln	Glu	Arg	Asp	Val	Ala	Ile	Glu	Glu	Lys	Gln
		675					680					685			
Glu	Val	Ser	Arg	Val	Leu	Glu	Gln	Val	Ser	Ala	Gln	Leu	Glu	Glu	Cys
	690					695				700					
Lys	Gly	Gln	Thr	Glu	Gln	Leu	Glu	Leu	Glu	Asn	Ile	Arg	Leu	Ala	Thr
705					710					715					720
Asp	Leu	Arg	Ala	Gln	Leu	Gln	Ile	Leu	Ala	Asn	Met	Asp	Ser	Gln	Leu
			725						730					735	
Lys	Glu	Leu	Gln	Ser	Gln	His	Thr	His	Cys	Ala	Gln	Asp	Leu	Ala	Met
			740					745					750		
Lys	Asp	Glu	Leu	Leu	Cys	Gln	Leu	Thr	Gln	Ser	Asn	Glu	Glu	Gln	Ala

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755					760					765					
Ala	Gln	Cys	Val	Lys	Glu	Glu	Met	Ala	Leu	Lys	His	Met	Gln	Ala	Glu
770						775					780				
Leu	Gln	Gln	Gln	Gln	Ala	Val	Leu	Ala	Lys	Glu	Val	Arg	Asp	Leu	Lys
785					790					795					800
Glu	Thr	Leu	Glu	Phe	Ala	Asp	Gln	Glu	Asn	Gln	Val	Ala	His	Leu	Glu
				805					810					815	
Leu	Gly	Gln	Val	Glu	Cys	Gln	Leu	Lys	Thr	Thr	Leu	Glu	Val	Leu	Arg
			820					825					830		
Glu	Arg	Ser	Leu	Gln	Cys	Glu	Asn	Leu	Lys	Asp	Thr	Val	Glu	Asn	Leu
		835					840					845			
Thr	Ala	Lys	Leu	Ala	Ser	Thr	Ile	Ala	Asp	Asn	Gln	Glu	Gln	Asp	Leu
						855					860				
Glu	Lys	Thr	Arg	Gln	Tyr	Ser	Gln	Lys	Leu	Gly	Leu	Leu	Thr	Glu	Gln
865					870					875					880
Leu	Gln	Ser	Leu	Thr	Leu	Phe	Leu	Gln	Thr	Lys	Leu	Lys	Glu	Lys	Thr
				885					890					895	
Glu	Gln	Glu	Thr	Leu	Leu	Leu	Ser	Thr	Ala	Cys	Pro	Pro	Thr	Gln	Glu
			900					905					910		
His	Pro	Leu	Pro	Asn	Asp	Arg	Thr	Phe	Leu	Gly	Ser	Ile	Leu	Thr	Ala
		915					920					925			
Val	Ala	Asp	Glu	Glu	Pro	Glu	Ser	Thr	Pro	Val	Pro	Leu	Leu	Gly	Ser
		930				935					940				
Asp	Lys	Ser	Ala	Phe	Thr	Arg	Val	Ala	Ser	Met	Val	Ser	Leu	Gln	Pro
945					950					955					960
Ala	Glu	Thr	Pro	Gly	Met	Glu	Glu	Ser	Leu	Ala	Glu	Met	Ser	Ile	Met
				965					970					975	
Thr	Thr	Glu	Leu	Gln	Ser	Leu	Cys	Ser	Leu	Leu	Gln	Glu	Ser	Lys	Glu
			980					985					990		
Glu	Ala	Ile	Arg	Thr	Leu	Gln	Arg	Lys	Ile	Cys	Glu	Leu	Gln	Ala	Arg
		995					1000					1005			
Leu	Gln	Ala	Gln	Glu	Glu	Gln	His	Gln	Glu	Val	Gln	Lys	Ala	Lys	Glu
						1015					1020				
Ala	Asp	Ile	Glu	Lys	Leu	Asn	Gln	Ala	Leu	Cys	Leu	Arg	Tyr	Lys	Asn
1025					1030					1035					1040
Glu	Lys	Glu	Leu	Gln	Glu	Val	Ile	Gln	Gln	Asn	Glu	Lys	Ile	Leu	Glu
				1045					1050					1055	
Gln	Ile	Asp	Lys	Ser	Gly	Glu	Leu	Ile	Ser	Leu	Arg	Glu	Glu	Val	Thr
			1060					1065				1070			
His	Leu	Thr	Arg	Ser	Leu	Arg	Arg	Ala	Glu	Thr	Glu	Thr	Lys	Val	Leu
		1075					1080					1085			
Gln	Glu	Ala	Trp	Gln	Ala	Ser	Trp	Thr	Pro	Thr	Ala	Ser	Leu	Trp	Pro
		1090				1095					1100				
Pro	Ile	Gly	Ser	Arg	Arg	Lys	Cys	Gly	Ser	Leu	Arg	Arg	Trp	Thr	Asn
1105					1110					1115					1120

<210> 96

<211> 540

<212> PRT

<213> Homo sapiens

<400> 96

Met	Gly	Thr	Thr	Ala	Arg	Ala	Ala	Leu	Val	Leu	Thr	Tyr	Leu	Ala	Val
1				5					10					15	
Ala	Ser	Ala	Ala	Ser	Glu	Gly	Gly	Phe	Thr	Ala	Thr	Gly	Gln	Arg	Gln
			20					25					30		
Leu	Arg	Pro	Glu	His	Phe	Gln	Glu	Val	Gly	Tyr	Ala	Ala	Pro	Pro	Ser

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35	40	45
Pro Pro Leu Ser Arg Ser Leu Pro Met Asp His Pro Asp Ser Ser Gln		
50	55	60
His Gly Pro Pro Phe Glu Gly Gln Ser Gln Val Gln Pro Pro Pro Ser		
65	70	75
Gln Glu Ala Thr Pro Leu Gln Gln Glu Lys Leu Leu Pro Ala Gln Leu		80
	85	90
Pro Ala Glu Lys Glu Val Gly Pro Pro Leu Pro Gln Glu Ala Val Pro		95
	100	105
Leu Gln Lys Glu Leu Pro Ser Leu Gln His Pro Asn Glu Gln Lys Glu		110
	115	120
Gly Thr Pro Ala Pro Phe Gly Asp Gln Ser His Pro Glu Pro Glu Ser		125
	130	135
Trp Asn Ala Ala Gln His Cys Gln Gln Asp Arg Ser Gln Gly Gly Trp		140
145	150	155
Gly His Arg Leu Asp Gly Phe Pro Pro Gly Arg Pro Ser Pro Asp Asn		160
	165	170
Leu Asn Gln Ile Cys Leu Pro Asn Arg Gln His Val Val Tyr Gly Pro		175
	180	185
Trp Asn Leu Pro Gln Ser Ser Tyr Ser His Leu Thr Arg Gln Gly Glu		190
	195	200
Thr Leu Asn Phe Leu Glu Ile Gly Tyr Ser Arg Cys Cys His Cys Arg		205
	210	215
Ser His Thr Asn Arg Leu Glu Cys Ala Lys Leu Val Trp Glu Glu Ala		220
225	230	235
Met Ser Arg Phe Cys Glu Ala Glu Phe Ser Val Lys Thr Arg Pro His		240
	245	250
Trp Cys Cys Thr Arg Gln Gly Glu Ala Arg Phe Ser Cys Phe Gln Glu		255
	260	265
Glu Ala Pro Gln Pro His Tyr Gln Leu Arg Ala Cys Pro Ser His Gln		270
	275	280
Pro Asp Ile Ser Ser Gly Leu Glu Leu Pro Phe Pro Pro Gly Val Pro		285
	290	295
Thr Leu Asp Asn Ile Lys Asn Ile Cys His Leu Arg Arg Phe Arg Ser		300
305	310	315
Val Pro Arg Asn Leu Pro Ala Thr Asp Pro Leu Gln Arg Glu Leu Leu		320
	325	330
Ala Leu Ile Gln Leu Glu Arg Glu Phe Gln Arg Cys Cys Arg Gln Gly		335
	340	345
Asn Asn His Thr Cys Thr Trp Lys Ala Trp Glu Asp Thr Leu Asp Lys		350
	355	360
Tyr Cys Asp Arg Glu Tyr Ala Val Lys Thr His His His Leu Cys Cys		365
	370	375
Arg His Pro Pro Ser Pro Thr Arg Asp Glu Cys Phe Ala Arg Arg Ala		380
385	390	395
Pro Tyr Pro Asn Tyr Asp Arg Asp Ile Leu Thr Ile Asp Ile Ser Arg		400
	405	410
Val Thr Pro Asn Leu Met Gly His Leu Cys Gly Asn Gln Arg Val Leu		415
	420	425
Thr Lys His Lys His Ile Pro Gly Leu Ile His Asn Met Thr Ala Arg		430
	435	440
Cys Cys Asp Leu Pro Phe Pro Glu Gln Ala Cys Cys Ala Glu Glu Glu		445
	450	455
Lys Leu Thr Phe Ile Asn Asp Leu Cys Gly Pro Arg Arg Asn Ile Trp		460
465	470	475
Arg Asp Pro Ala Leu Cys Cys Tyr Leu Ser Pro Gly Asp Glu Gln Val		480
	485	490
		495

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Asn Cys Phe Asn Ile Asn Tyr Leu Arg Asn Val Ala Leu Val Ser Gly
 500 505 510
 Asp Thr Glu Asn Ala Lys Gly Gln Gly Glu Gln Gly Ser Thr Gly Gly
 515 520 525
 Thr Asn Ile Ser Ser Thr Ser Glu Pro Lys Glu Glu
 530 535 540

<210> 97

<211> 462

<212> PRT

<213> Homo sapiens

<400> 97

Met Gly Lys Glu Lys Thr His Ile Asn Ile Val Val Ile Gly His Val
 1 5 10 15
 Asp Ser Gly Lys Ser Thr Thr Thr Gly His Leu Ile Tyr Lys Cys Gly
 20 25 30
 Gly Ile Asp Lys Arg Thr Ile Glu Lys Phe Glu Lys Glu Ala Ala Glu
 35 40 45
 Met Gly Lys Gly Ser Phe Lys Tyr Ala Trp Val Leu Asp Lys Leu Lys
 50 55 60
 Ala Glu Arg Glu Arg Gly Ile Thr Ile Asp Ile Ser Leu Trp Lys Phe
 65 70 75 80
 Glu Thr Ser Lys Tyr Tyr Val Thr Ile Ile Asp Ala Pro Gly His Arg
 85 90 95
 Asp Phe Ile Lys Asn Met Ile Thr Gly Thr Ser Gln Ala Asp Cys Ala
 100 105 110
 Val Leu Ile Val Ala Ala Gly Val Gly Glu Phe Glu Ala Gly Ile Ser
 115 120 125
 Lys Asn Gly Gln Thr Arg Glu His Ala Leu Leu Ala Tyr Thr Leu Gly
 130 135 140
 Val Lys Gln Leu Ile Val Gly Val Asn Lys Met Asp Ser Thr Glu Pro
 145 150 155 160
 Pro Tyr Ser Gln Lys Arg Tyr Glu Glu Ile Val Lys Glu Val Ser Thr
 165 170 175
 Tyr Ile Lys Lys Ile Gly Tyr Asn Pro Asp Thr Val Ala Phe Val Pro
 180 185 190
 Ile Ser Gly Trp Asn Gly Asp Asn Met Leu Glu Pro Ser Ala Asn Met
 195 200 205
 Pro Trp Phe Lys Gly Trp Lys Val Thr Arg Lys Asp Gly Asn Ala Ser
 210 215 220
 Gly Thr Thr Leu Leu Glu Ala Val Asp Cys Ile Leu Pro Pro Thr Arg
 225 230 235 240
 Pro Thr Asp Lys Pro Leu Arg Leu Pro Leu Gln Asp Val Tyr Lys Ile
 245 250 255
 Gly Gly Ile Gly Thr Val Pro Val Gly Arg Val Glu Thr Gly Val Leu
 260 265 270
 Lys Pro Gly Met Val Val Thr Phe Ala Pro Val Asn Val Thr Thr Glu
 275 280 285
 Val Lys Ser Val Glu Met His His Glu Ala Leu Ser Glu Ala Leu Pro
 290 295 300
 Gly Asp Asn Val Gly Phe Asn Val Lys Asn Val Ser Val Lys Asp Val
 305 310 315 320
 Arg Arg Gly Asn Val Ala Gly Asp Ser Lys Asn Asp Pro Pro Met Glu
 325 330 335
 Ala Ala Gly Phe Thr Ala Gln Val Ile Ile Leu Asn His Pro Gly Gln
 340 345 350

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Ile Ser Ala Gly Tyr Ala Pro Val Leu Asp Cys His Thr Ala His Ile
 355 360 365
 Ala Cys Lys Phe Ala Glu Leu Lys Glu Lys Ile Asp Arg Arg Ser Gly
 370 375 380
 Lys Lys Leu Glu Asp Gly Pro Lys Phe Leu Lys Ser Gly Asp Ala Ala
 385 390 395 400
 Ile Val Asp Met Val Pro Gly Lys Pro Met Cys Val Glu Ser Phe Ser
 405 410 415
 Asp Tyr Pro Pro Leu Gly Arg Phe Ala Val Arg Asp Met Arg Gln Thr
 420 425 430
 Val Ala Val Gly Val Ile Lys Ala Val Asp Lys Lys Ala Ala Gly Ala
 435 440 445
 Gly Lys Val Thr Lys Ser Ala Gln Lys Ala Gln Lys Ala Lys
 450 455 460

<210> 98

<211> 2328

<212> PRT

<213> Homo sapiens

<400> 98

Lys Ser Lys Arg Gln Ala Gln Gln Met Val Gln Pro Gln Ser Pro Val
 1 5 10 15
 Ala Val Ser Gln Ser Lys Pro Gly Cys Tyr Asp Asn Gly Lys His Tyr
 20 25 30
 Gln Ile Asn Gln Gln Trp Glu Arg Thr Tyr Leu Gly Asn Val Leu Val
 35 40 45
 Cys Thr Cys Tyr Gly Gly Ser Arg Gly Phe Asn Cys Glu Ser Lys Pro
 50 55 60
 Glu Ala Glu Glu Thr Cys Phe Asp Lys Tyr Thr Gly Asn Thr Tyr Arg
 65 70 75 80
 Val Gly Asp Thr Tyr Glu Arg Pro Lys Asp Ser Met Ile Trp Asp Cys
 85 90 95
 Thr Cys Ile Gly Ala Gly Arg Gly Arg Ile Ser Cys Thr Ile Ala Asn
 100 105 110
 Arg Cys His Glu Gly Gly Gln Ser Tyr Lys Ile Gly Asp Thr Trp Arg
 115 120 125
 Arg Pro His Glu Thr Gly Gly Tyr Met Leu Glu Cys Val Cys Leu Gly
 130 135 140
 Asn Gly Lys Gly Glu Trp Thr Cys Lys Pro Ile Ala Glu Lys Cys Phe
 145 150 155 160
 Asp His Ala Ala Gly Thr Ser Tyr Val Val Gly Glu Thr Trp Glu Lys
 165 170 175
 Pro Tyr Gln Gly Trp Met Met Val Asp Cys Thr Cys Leu Gly Glu Gly
 180 185 190
 Ser Gly Arg Ile Thr Cys Thr Ser Arg Asn Arg Cys Asn Asp Gln Asp
 195 200 205
 Thr Arg Thr Ser Tyr Arg Ile Gly Asp Thr Trp Ser Lys Lys Asp Asn
 210 215 220
 Arg Gly Asn Leu Leu Gln Cys Ile Cys Thr Gly Asn Gly Arg Gly Glu
 225 230 235 240
 Trp Lys Cys Glu Arg His Thr Ser Val Gln Thr Thr Ser Ser Gly Ser
 245 250 255
 Gly Pro Phe Thr Asp Val Arg Ala Ala Val Tyr Gln Pro Gln Pro His
 260 265 270
 Pro Gln Pro Pro Pro Tyr Gly His Cys Val Thr Asp Ser Gly Val Val
 275 280 285

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Tyr	Ser	Val	Gly	Met	Gln	Trp	Leu	Lys	Thr	Gln	Gly	Asn	Lys	Gln	Met
290						295					300				
Leu	Cys	Thr	Cys	Leu	Gly	Asn	Gly	Val	Ser	Cys	Gln	Glu	Thr	Ala	Val
305					310					315					320
Thr	Gln	Thr	Tyr	Gly	Gly	Asn	Leu	Asn	Gly	Glu	Pro	Cys	Val	Leu	Pro
				325					330						335
Phe	Thr	Tyr	Asn	Gly	Arg	Thr	Phe	Tyr	Ser	Cys	Thr	Thr	Glu	Gly	Arg
			340					345					350		
Gln	Asp	Gly	His	Leu	Trp	Cys	Ser	Thr	Thr	Ser	Asn	Tyr	Glu	Gln	Asp
		355					360					365			
Gln	Lys	Tyr	Ser	Phe	Cys	Thr	Asp	His	Thr	Val	Leu	Val	Gln	Thr	Gln
		370					375				380				
Gly	Gly	Asn	Ser	Asn	Gly	Ala	Leu	Cys	His	Phe	Pro	Phe	Leu	Tyr	Asn
385					390					395					400
Asn	His	Asn	Tyr	Thr	Asp	Cys	Thr	Ser	Glu	Gly	Arg	Arg	Asp	Asn	Met
				405					410					415	
Lys	Trp	Cys	Gly	Thr	Thr	Gln	Asn	Tyr	Asp	Ala	Asp	Gln	Lys	Phe	Gly
			420					425					430		
Phe	Cys	Pro	Met	Ala	Ala	His	Glu	Glu	Ile	Cys	Thr	Thr	Asn	Glu	Gly
		435					440					445			
Val	Met	Tyr	Arg	Ile	Gly	Asp	Gln	Trp	Asp	Lys	Gln	His	Asp	Met	Gly
		450				455					460				
His	Met	Met	Arg	Cys	Thr	Cys	Val	Gly	Asn	Gly	Arg	Gly	Glu	Trp	Thr
465					470					475					480
Cys	Ile	Ala	Tyr	Ser	Gln	Leu	Arg	Asp	Gln	Cys	Ile	Val	Asp	Asp	Ile
				485					490					495	
Thr	Tyr	Asn	Val	Asn	Asp	Thr	Phe	His	Lys	Arg	His	Glu	Glu	Gly	His
			500					505					510		
Met	Leu	Asn	Cys	Thr	Cys	Phe	Gly	Gln	Gly	Arg	Gly	Arg	Trp	Lys	Cys
		515					520					525			
Asp	Pro	Val	Asp	Gln	Cys	Gln	Asp	Ser	Glu	Thr	Gly	Thr	Phe	Tyr	Gln
		530				535					540				
Ile	Gly	Asp	Ser	Trp	Glu	Lys	Tyr	Val	His	Gly	Val	Arg	Tyr	Gln	Cys
545					550					555					560
Tyr	Cys	Tyr	Gly	Arg	Gly	Ile	Gly	Glu	Trp	His	Cys	Gln	Pro	Leu	Gln
				565					570					575	
Thr	Tyr	Pro	Ser	Ser	Ser	Gly	Pro	Val	Glu	Val	Phe	Ile	Thr	Glu	Thr
			580					585					590		
Pro	Ser	Gln	Pro	Asn	Ser	His	Pro	Ile	Gln	Trp	Asn	Ala	Pro	Gln	Pro
		595					600					605			
Ser	His	Ile	Ser	Lys	Tyr	Ile	Leu	Arg	Trp	Arg	Pro	Lys	Asn	Ser	Val
		610				615					620				
Gly	Arg	Trp	Lys	Glu	Ala	Thr	Ile	Pro	Gly	His	Leu	Asn	Ser	Tyr	Thr
625					630					635					640
Ile	Lys	Gly	Leu	Lys	Pro	Gly	Val	Val	Tyr	Glu	Gly	Gln	Leu	Ile	Ser
				645					650					655	
Ile	Gln	Gln	Tyr	Gly	His	Gln	Glu	Val	Thr	Arg	Phe	Asp	Phe	Thr	Thr
			660					665					670		
Thr	Ser	Thr	Ser	Thr	Pro	Val	Thr	Ser	Asn	Thr	Val	Thr	Gly	Glu	Thr
		675					680					685			
Thr	Pro	Phe	Ser	Pro	Leu	Val	Ala	Thr	Ser	Glu	Ser	Val	Thr	Glu	Ile
		690				695						700			
Thr	Ala	Ser	Ser	Phe	Val	Val	Ser	Trp	Val	Ser	Ala	Ser	Asp	Thr	Val
705					710					715					720
Ser	Gly	Phe	Arg	Val	Glu	Tyr	Glu	Leu	Ser	Glu	Glu	Gly	Asp	Glu	Pro
				725					730					735	
Gln	Tyr	Leu	Asp	Leu	Pro	Ser	Thr	Ala	Thr	Ser	Val	Asn	Ile	Pro	Asp

			740					745					750			
Leu	Leu	Pro	Gly	Arg	Lys	Tyr	Ile	Val	Asn	Val	Tyr	Gln	Ile	Ser	Glu	
		755						760					765			
Asp	Gly	Glu	Gln	Ser	Leu	Ile	Leu	Ser	Thr	Ser	Gln	Thr	Thr	Ala	Pro	
	770					775					780					
Asp	Ala	Pro	Pro	Asp	Pro	Thr	Val	Asp	Gln	Val	Asp	Asp	Thr	Ser	Ile	
785					790					795					800	
Val	Val	Arg	Trp	Ser	Arg	Pro	Gln	Ala	Pro	Ile	Thr	Gly	Tyr	Arg	Ile	
				805					810					815		
Val	Tyr	Ser	Pro	Ser	Val	Glu	Gly	Ser	Ser	Thr	Glu	Leu	Asn	Leu	Pro	
			820					825					830			
Glu	Thr	Ala	Asn	Ser	Val	Thr	Leu	Ser	Asp	Leu	Gln	Pro	Gly	Val	Gln	
		835					840					845				
Tyr	Asn	Ile	Thr	Ile	Tyr	Ala	Val	Glu	Glu	Asn	Gln	Glu	Ser	Thr	Pro	
	850					855					860					
Val	Val	Ile	Gln	Gln	Glu	Thr	Thr	Gly	Thr	Pro	Arg	Ser	Asp	Thr	Val	
865					870					875					880	
Pro	Ser	Pro	Arg	Asp	Leu	Gln	Phe	Val	Glu	Val	Thr	Asp	Val	Lys	Val	
				885					890					895		
Thr	Ile	Met	Trp	Thr	Pro	Pro	Glu	Ser	Ala	Val	Thr	Gly	Tyr	Arg	Val	
			900					905					910			
Asp	Val	Ile	Pro	Val	Asn	Leu	Pro	Gly	Glu	His	Gly	Gln	Arg	Leu	Pro	
		915					920					925				
Ile	Ser	Arg	Asn	Thr	Phe	Ala	Glu	Val	Thr	Gly	Leu	Ser	Pro	Gly	Val	
	930					935					940					
Thr	Tyr	Tyr	Phe	Lys	Val	Phe	Ala	Val	Ser	His	Gly	Arg	Glu	Ser	Lys	
945				950						955					960	
Pro	Leu	Thr	Ala	Gln	Gln	Thr	Thr	Lys	Leu	Asp	Ala	Pro	Thr	Asn	Leu	
				965					970					975		
Gln	Phe	Val	Asn	Glu	Thr	Asp	Ser	Thr	Val	Leu	Val	Arg	Trp	Thr	Pro	
			980					985					990			
Pro	Arg	Ala	Gln	Ile	Thr	Gly	Tyr	Arg	Leu	Thr	Val	Gly	Leu	Thr	Arg	
		995					1000					1005				
Arg	Gly	Gln	Pro	Arg	Gln	Tyr	Asn	Val	Gly	Pro	Ser	Val	Ser	Lys	Tyr	
	1010					1015					1020					
Pro	Leu	Arg	Asn	Leu	Gln	Pro	Ala	Ser	Glu	Tyr	Thr	Val	Ser	Leu	Val	
1025				1030						1035					1040	
Ala	Ile	Lys	Gly	Asn	Gln	Glu	Ser	Pro	Lys	Ala	Thr	Gly	Val	Phe	Thr	
				1045					1050					1055		
Thr	Leu	Gln	Pro	Gly	Ser	Ser	Ile	Pro	Pro	Tyr	Asn	Thr	Glu	Val	Thr	
			1060					1065					1070			
Glu	Thr	Thr	Ile	Val	Ile	Thr	Trp	Thr	Pro	Ala	Pro	Arg	Ile	Gly	Phe	
		1075					1080									

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Asp Gln Ser Ser Cys Thr Phe Asp Asn Leu Ser Pro Gly Leu Glu Tyr
 1205 1210 1215
 Asn Val Ser Val Tyr Thr Val Lys Asp Asp Lys Glu Ser Val Pro Ile
 1220 1225 1230
 Ser Asp Thr Ile Ile Pro Ala Val Pro Pro Pro Thr Asp Leu Arg Phe
 1235 1240 1245
 Thr Asn Ile Gly Pro Asp Thr Met Arg Val Thr Trp Ala Pro Pro Pro
 1250 1255 1260
 Ser Ile Asp Leu Thr Asn Phe Leu Val Arg Tyr Ser Pro Val Lys Asn
 1265 1270 1275 1280
 Glu Glu Asp Val Ala Glu Leu Ser Ile Ser Pro Ser Asp Asn Ala Val
 1285 1290 1295
 Val Leu Thr Asn Leu Leu Pro Gly Thr Glu Tyr Val Val Ser Val Ser
 1300 1305 1310
 Ser Val Tyr Glu Gln His Glu Ser Thr Pro Leu Arg Gly Arg Gln Lys
 1315 1320 1325
 Thr Gly Leu Asp Ser Pro Thr Gly Ile Asp Phe Ser Asp Ile Thr Ala
 1330 1335 1340
 Asn Ser Phe Thr Val His Trp Ile Ala Pro Arg Ala Thr Ile Thr Gly
 1345 1350 1355 1360
 Tyr Arg Ile Arg His His Pro Glu His Phe Ser Gly Arg Pro Arg Glu
 1365 1370 1375
 Asp Arg Val Pro His Ser Arg Asn Ser Ile Thr Leu Thr Asn Leu Thr
 1380 1385 1390
 Pro Gly Thr Glu Tyr Val Val Ser Ile Val Ala Leu Asn Gly Arg Glu
 1395 1400 1405
 Glu Ser Pro Leu Leu Ile Gly Gln Gln Ser Thr Val Ser Asp Val Pro
 1410 1415 1420
 Arg Asp Leu Glu Val Val Ala Ala Thr Pro Thr Ser Leu Leu Ile Ser
 1425 1430 1435 1440
 Trp Asp Ala Pro Ala Val Thr Val Arg Tyr Tyr Arg Ile Thr Tyr Gly
 1445 1450 1455
 Glu Thr Gly Gly Asn Ser Pro Val Gln Glu Phe Thr Val Pro Gly Ser
 1460 1465 1470
 Lys Ser Thr Ala Thr Ile Ser Gly Leu Lys Pro Gly Val Asp Tyr Thr
 1475 1480 1485
 Ile Thr Val Tyr Ala Val Thr Gly Arg Gly Asp Ser Pro Ala Ser Ser
 1490 1495 1500
 Lys Pro Ile Ser Ile Asn Tyr Arg Thr Glu Ile Asp Lys Pro Ser Gln
 1505 1510 1515 1520
 Met Gln Val Thr Asp Val Gln Asp Asn Ser Ile Ser Val Lys Trp Leu
 1525 1530 1535
 Pro Ser Ser Ser Pro Val Thr Gly Tyr Arg Val Thr Thr Thr Pro Lys
 1540 1545 1550
 Asn Gly Pro Gly Pro Thr Lys Thr Lys Thr Ala Gly Pro Asp Gln Thr
 1555 1560 1565
 Glu Met Thr Ile Glu Gly Leu Gln Pro Thr Val Glu Tyr Val Val Ser
 1570 1575 1580
 Val Tyr Ala Gln Asn Pro Ser Gly Glu Ser Gln Pro Leu Val Gln Thr
 1585 1590 1595 1600
 Ala Val Thr Asn Ile Asp Arg Pro Lys Gly Leu Ala Phe Thr Asp Val
 1605 1610 1615
 Asp Val Asp Ser Ile Lys Ile Ala Trp Glu Ser Pro Gln Gly Gln Val
 1620 1625 1630
 Ser Arg Tyr Arg Val Thr Tyr Ser Ser Pro Glu Asp Gly Ile His Glu
 1635 1640 1645
 Leu Phe Pro Ala Pro Asp Gly Glu Glu Asp Thr Ala Glu Leu Gln Gly

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1650	1655	1660
Leu Arg Pro Gly Ser Glu Tyr Thr Val Ser Val Val Ala Leu His Asp		
1665	1670	1675
Asp Met Glu Ser Gln Pro Leu Ile Gly Thr Gln Ser Thr Ala Ile Pro		1680
	1685	1690
Ala Pro Thr Asp Leu Lys Phe Thr Gln Val Thr Pro Thr Ser Leu Ser		1695
	1700	1705
Ala Gln Trp Thr Pro Pro Asn Val Gln Leu Thr Gly Tyr Arg Val Arg		1710
	1715	1720
Val Thr Pro Lys Glu Lys Thr Gly Pro Met Lys Glu Ile Asn Leu Ala		1725
	1730	1735
Pro Asp Ser Ser Ser Val Val Val Ser Gly Leu Met Val Ala Thr Lys		1740
1745	1750	1755
Tyr Glu Val Ser Val Tyr Ala Leu Lys Asp Thr Leu Thr Ser Arg Pro		1760
	1765	1770
Ala Gln Gly Val Val Thr Thr Leu Glu Asn Val Ser Pro Pro Arg Arg		1775
	1780	1785
Ala Arg Val Thr Asp Ala Thr Glu Thr Thr Ile Thr Ile Ser Trp Arg		1790
	1795	1800
Thr Lys Thr Glu Thr Ile Thr Gly Phe Gln Val Asp Ala Val Pro Ala		1805
	1810	1815
Asn Gly Gln Thr Pro Ile Gln Arg Thr Ile Lys Pro Asp Val Arg Ser		1820
1825	1830	1835
Tyr Thr Ile Thr Gly Leu Gln Pro Gly Thr Asp Tyr Lys Ile Tyr Leu		1840
	1845	1850
Tyr Thr Leu Asn Asp Asn Ala Arg Ser Ser Pro Val Val Ile Asp Ala		1855
	1860	1865
Ser Thr Ala Ile Asp Ala Pro Ser Asn Leu Arg Phe Leu Ala Thr Thr		1870
	1875	1880
Pro Asn Ser Leu Leu Val Ser Trp Gln Pro Pro Arg Ala Arg Ile Thr		1885
	1890	1895
Gly Tyr Ile Ile Lys Tyr Glu Lys Pro Gly Ser Pro Pro Arg Glu Val		1900
1905	1910	1915
Val Pro Arg Pro Arg Pro Gly Val Thr Glu Ala Thr Ile Thr Gly Leu		1920
	1925	1930
Glu Pro Gly Thr Glu Tyr Thr Ile Tyr Val Ile Ala Leu Lys Asn Asn		1935
	1940	1945
Gln Lys Ser Glu Pro Leu Ile Gly Arg Lys Lys Thr Asp Glu Leu Pro		1950
	1955	1960
Gln Leu Val Thr Leu Pro His Pro Asn Leu His Gly Pro Glu Ile Leu		1965
	1970	1975
Asp Val Pro Ser Thr Val Gln Lys Thr Pro Phe Val Thr His Pro Gly		1980
1985	1990	1995
Tyr Asp Thr Gly Asn Gly Ile Gln Leu Pro Gly Thr Ser Gly Gln Gln		2000
	2005	2010
Pro Ser Val Gly Gln Gln Met Ile Phe Glu Glu His Gly Phe Arg Arg		2015
	2020	2025
Thr Thr Pro Pro Thr Thr Ala Thr Pro Ile Arg His Arg Pro Arg Pro		2030
	2035	2040
Tyr Pro Pro Asn Val Gly Gln Glu Ala Leu Ser Gln Thr Thr Ile Ser		2045
	2050	2055
Trp Ala Pro Phe Gln Asp Thr Ser Glu Tyr Ile Ile Ser Cys His Pro		2060
2065	2070	2075
Val Gly Thr Asp Glu Glu Pro Leu Gln Phe Arg Val Pro Gly Thr Ser		2080
	2085	2090
Thr Ser Ala Thr Leu Thr Gly Leu Thr Arg Gly Ala Thr Tyr Asn Ile		2095
	2100	2105
		2110

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Ile Val Glu Ala Leu Lys Asp Gln Gln Arg His Lys Val Arg Glu Glu
 2115 2120 2125
 Val Val Thr Val Gly Asn Ser Val Asn Glu Gly Leu Asn Gln Pro Thr
 2130 2135 2140
 Asp Asp Ser Cys Phe Asp Pro Tyr Thr Val Ser His Tyr Ala Val Gly
 2145 2150 2155 2160
 Asp Glu Trp Glu Arg Met Ser Glu Ser Gly Phe Lys Leu Leu Cys Gln
 2165 2170 2175
 Cys Leu Gly Phe Gly Ser Gly His Phe Arg Cys Asp Ser Ser Arg Trp
 2180 2185 2190
 Cys His Asp Asn Gly Val Asn Tyr Lys Ile Gly Glu Lys Trp Asp Arg
 2195 2200 2205
 Gln Gly Glu Asn Gly Gln Met Met Ser Cys Thr Cys Leu Gly Asn Gly
 2210 2215 2220
 Lys Gly Glu Phe Lys Cys Asp Pro His Glu Ala Thr Cys Tyr Asp Asp
 2225 2230 2235 2240
 Gly Lys Thr Tyr His Val Gly Glu Gln Trp Gln Lys Glu Tyr Leu Gly
 2245 2250 2255
 Ala Ile Cys Ser Cys Thr Cys Phe Gly Gly Gln Arg Gly Trp Arg Cys
 2260 2265 2270
 Asp Asn Cys Arg Arg Pro Gly Gly Glu Pro Ser Pro Glu Gly Thr Thr
 2275 2280 2285
 Gly Gln Ser Tyr Asn Gln Tyr Ser Gln Arg Tyr His Gln Arg Thr Asn
 2290 2295 2300
 Thr Asn Val Asn Cys Pro Ile Glu Cys Phe Met Pro Leu Asp Val Gln
 2305 2310 2315 2320
 Ala Asp Arg Glu Asp Ser Arg Glu
 2325

<210> 99

<211> 188

<212> PRT

<213> Homo sapiens

<400> 99

His Gln Thr His Lys Glu Gly Gly Ser Thr His Ala Ser Ala Asp Ala
 1 5 10 15
 Trp Glu Ile Ile Glu Leu Glu Thr Glu Ile Glu Lys Phe Lys Ala Glu
 20 25 30
 Asn Ala Ser Leu Ala Lys Leu Arg Ile Glu Arg Glu Ser Ala Leu Glu
 35 40 45
 Lys Leu Arg Lys Glu Ile Ala Asp Phe Glu Gln Gln Lys Ala Lys Glu
 50 55 60
 Leu Ala Arg Ile Glu Glu Phe Lys Lys Glu Glu Met Arg Lys Leu Gln
 65 70 75 80
 Lys Glu Arg Lys Val Phe Glu Lys Tyr Thr Thr Ala Ala Arg Thr Phe
 85 90 95
 Pro Asp Lys Lys Glu Arg Glu Glu Ile Gln Thr Leu Lys Gln Gln Ile
 100 105 110
 Ala Asp Leu Arg Glu Asp Leu Lys Arg Lys Glu Thr Lys Trp Ser Ser
 115 120 125
 Thr His Ser Arg Leu Arg Ser Gln Ile Gln Met Leu Val Arg Glu Asn
 130 135 140
 Thr Asp Leu Arg Glu Glu Ile Lys Val Met Glu Arg Phe Arg Leu Asp
 145 150 155 160
 Ala Trp Lys Arg Ala Glu Ala Ile Glu Ser Ser Leu Glu Val Glu Lys
 165 170 175

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Lys Asp Lys Leu Ala Asn Thr Ser Val Arg Phe Gln
 180 185

<210> 100
 <211> 284
 <212> PRT
 <213> Homo sapiens

<400> 100
 Met Glu Pro Gly Asn Tyr Ala Thr Leu Asp Gly Ala Lys Asp Ile Glu
 1 5 10 15
 Gly Leu Leu Gly Ala Gly Gly Gly Arg Asn Leu Val Ala His Ser Pro
 20 25 30
 Leu Thr Ser His Pro Ala Ala Pro Thr Leu Met Pro Ala Val Asn Tyr
 35 40 45
 Ala Pro Leu Asp Leu Pro Gly Ser Ala Glu Pro Pro Lys Gln Cys His
 50 55 60
 Pro Cys Pro Gly Val Pro Gln Gly Thr Ser Pro Ala Pro Val Pro Tyr
 65 70 75 80
 Gly Tyr Phe Gly Gly Gly Tyr Tyr Ser Cys Arg Val Ser Arg Ser Ser
 85 90 95
 Leu Lys Pro Cys Ala Gln Ala Ala Thr Leu Ala Ala Tyr Pro Ala Glu
 100 105 110
 Thr Pro Thr Ala Gly Glu Glu Tyr Pro Ser Arg Pro Thr Glu Phe Ala
 115 120 125
 Phe Tyr Pro Gly Tyr Pro Gly Thr Tyr His Ala Met Ala Ser Tyr Leu
 130 135 140
 Asp Val Ser Val Val Gln Thr Leu Gly Ala Pro Gly Glu Pro Arg His
 145 150 155 160
 Asp Ser Leu Leu Pro Val Asp Ser Tyr Gln Ser Trp Ala Leu Ala Gly
 165 170 175
 Gly Trp Asn Ser Gln Met Cys Cys Gln Gly Glu Gln Asn Pro Pro Gly
 180 185 190
 Pro Phe Trp Lys Ala Ala Phe Ala Asp Ser Ser Gly Gln His Pro Pro
 195 200 205
 Asp Ala Cys Ala Phe Arg Arg Gly Arg Lys Lys Arg Ile Pro Tyr Ser
 210 215 220
 Lys Gly Gln Leu Arg Glu Leu Glu Arg Glu Tyr Ala Ala Asn Lys Phe
 225 230 235 240
 Ile Thr Lys Asp Lys Arg Arg Lys Ile Ser Ala Ala Thr Ser Leu Ser
 245 250 255
 Glu Arg Gln Ile Thr Ile Trp Phe Gln Asn Arg Arg Val Lys Glu Lys
 260 265 270
 Lys Val Leu Ala Lys Val Lys Asn Ser Ala Thr Pro
 275 280

<210> 101
 <211> 676
 <212> PRT
 <213> Homo sapiens

<400> 101
 Met Asp Lys Tyr Asp Asp Leu Gly Leu Glu Ala Ser Lys Phe Ile Glu
 1 5 10 15
 Asp Leu Asn Met Tyr Glu Ala Ser Lys Asp Gly Leu Phe Arg Val Asp
 20 25 30
 Lys Gly Ala Gly Asn Asn Pro Glu Phe Glu Glu Thr Arg Arg Val Phe

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35	40	45
Ala Thr Lys Met Ala Lys	Ile His Leu Gln Gln Gln Gln Gln Leu	
50	55	60
Leu Gln Glu Glu Thr Leu	Pro Arg Gly Ser Arg Gly Pro Val Asn Gly	
65	70	75
Gly Gly Arg Leu Gly Pro	Gln Ala Arg Trp Glu Val Val Gly Ser Lys	80
85	90	95
Leu Thr Val Asp Gly Ala	Ala Lys Pro Pro Leu Ala Ala Ser Thr Gly	
100	105	110
Ala Pro Gly Ala Val Thr	Thr Leu Ala Ala Gly Gln Pro Pro Tyr Pro	
115	120	125
Pro Gln Glu Gln Arg Ser	Arg Pro Tyr Leu His Gly Thr Arg His Gly	
130	135	140
Ser Gln Asp Cys Gly Ser	Arg Glu Ser Leu Ala Thr Ser Glu Met Ser	160
145	150	155
Ala Phe His Gln Pro Gly	Pro Cys Glu Asp Pro Ser Cys Leu Thr His	
165	170	175
Gly Asp Tyr Tyr Asp Asn	Leu Ser Leu Ala Ser Pro Lys Trp Gly Asp	
180	185	190
Lys Pro Gly Val Ser Pro	Ser Ile Gly Leu Ser Val Gly Ser Gly Trp	
195	200	205
Pro Ser Ser Pro Gly Ser	Asp Pro Pro Leu Pro Lys Pro Cys Gly Asp	
210	215	220
His Pro Leu Asn His Arg	Gln Leu Ser Leu Ser Ser Ser Arg Ser Ser	
225	230	235
Glu Gly Ser Leu Gly Gly	Gln Asn Ser Gly Ile Gly Gly Arg Ser Ser	
245	250	255
Glu Lys Pro Thr Gly Leu	Trp Ser Thr Ala Ser Ser Gln Arg Val Ser	
260	265	270
Pro Gly Leu Pro Ser Pro	Asn Leu Glu Asn Gly Ala Pro Ala Val Gly	
275	280	285
Pro Val Gln Pro Arg Thr	Pro Ser Val Ser Ala Pro Leu Ala Leu Ser	
290	295	300
Cys Pro Arg Gln Gly Gly	Leu Pro Arg Ser Asn Ser Gly Leu Gly Gly	
305	310	315
Glu Val Ser Gly Val Met	Ser Lys Pro Asn Val Asp Pro Gln Pro Trp	
325	330	335
Phe Gln Asp Gly Pro Lys	Ser Tyr Leu Ser Ser Ser Ala Pro Ser Ser	
340	345	350
Ser Pro Ala Gly Leu Asp	Gly Ser Gln Gln Gly Ala Val Pro Gly Leu	
355	360	365
Gly Pro Lys Pro Gly Cys	Thr Asp Leu Gly Thr Gly Pro Lys Leu Ser	
370	375	380
Pro Thr Ser Leu Val His	Pro Val Met Ser Thr Leu Pro Glu Leu Ser	
385	390	395
Cys Lys Glu Gly Pro Leu	Gly Trp Ser Ser Asp Gly Ser Leu Gly Ser	
405	410	415
Val Leu Leu Asp Ser Pro	Ser Ser Pro Arg Val Arg Leu Pro Cys Gln	
420	425	430
Pro Leu Val Pro Gly Pro	Glu Leu Arg Pro Ser Ala Ala Glu Leu Lys	
435	440	445
Leu Glu Ala Leu Thr Gln	Arg Leu Glu Arg Glu Met Asp Ala His Pro	
450	455	460
Lys Ala Asp Tyr Phe Gly	Ala Cys Val Lys Cys Ser Lys Gly Val Phe	
465	470	475
Gly Ala Gly Gln Ala Cys	Gln Ala Met Gly Asn Leu Tyr His Asp Thr	
485	490	495

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Cys Phe Thr Cys Ala Ala Cys Ser Arg Lys Leu Arg Gly Lys Ala Phe
 500 505 510
 Tyr Phe Val Asn Gly Lys Val Phe Cys Glu Glu Asp Phe Leu Tyr Ser
 515 520 525
 Gly Phe Gln Gln Ser Ala Asp Arg Cys Phe Leu Cys Gly His Leu Ile
 530 535 540
 Met Asp Met Ile Leu Gln Ala Leu Gly Lys Ser Tyr His Pro Gly Cys
 545 550 555 560
 Phe Arg Cys Val Ile Cys Asn Glu Cys Leu Asp Gly Val Pro Phe Thr
 565 570 575
 Val Asp Ser Glu Asn Lys Ile Tyr Cys Val Arg Asp Tyr His Lys Val
 580 585 590
 Leu Ala Pro Lys Cys Ala Ala Cys Gly Leu Pro Ile Leu Pro Pro Glu
 595 600 605
 Gly Ser Asp Glu Thr Ile Arg Val Val Ser Met Asp Arg Asp Tyr His
 610 615 620
 Val Glu Cys Tyr His Cys Glu Asp Cys Gly Leu Glu Leu Asn Asp Glu
 625 630 635 640
 Asp Gly His Arg Cys Tyr Pro Leu Glu Asp His Leu Phe Cys His Ser
 645 650 655
 Cys His Val Lys Arg Leu Glu Lys Arg Pro Ser Ser Thr Ala Leu His
 660 665 670
 Gln His His Phe
 675

<210> 102

<211> 296

<212> PRT

<213> Homo sapiens

<400> 102

Ser Thr Gly Ser Glu Phe Pro Leu Cys Thr Lys Ala Ser Pro Cys Ser
 1 5 10 15
 Ala Ala Arg Ala Gly Gly Arg Ala Leu Gly Trp Arg Leu Gln Gln Gln
 20 25 30
 Arg Glu Thr Arg Gly Asn Pro Gly Asn Pro Gly Leu Gly Val Ala Ala
 35 40 45
 Thr Met Thr Gly Ser Asn Met Ser Asp Ala Leu Ala Asn Ala Val Cys
 50 55 60
 Gln Arg Cys Gln Ala Arg Phe Ser Pro Ala Glu Arg Ile Val Asn Ser
 65 70 75 80
 Asn Gly Glu Leu Tyr His Glu His Cys Phe Val Cys Ala Gln Cys Phe
 85 90 95
 Arg Pro Phe Pro Glu Gly Leu Phe Tyr Glu Phe Glu Gly Arg Lys Tyr
 100 105 110
 Cys Glu His Asp Phe Gln Met Leu Phe Ala Pro Cys Cys Gly Ser Cys
 115 120 125
 Gly Glu Phe Ile Ile Gly Arg Val Ile Lys Ala Met Asn Asn Asn Trp
 130 135 140
 His Pro Gly Cys Phe Arg Cys Glu Leu Cys Asp Val Glu Leu Ala Asp
 145 150 155 160
 Leu Gly Phe Val Lys Asn Ala Gly Arg His Leu Cys Arg Pro Cys His
 165 170 175
 Asn Arg Glu Lys Ala Lys Gly Leu Gly Lys Tyr Ile Cys Gln Arg Cys
 180 185 190
 His Leu Val Ile Asp Glu Gln Pro Leu Met Phe Arg Ser Asp Ala Tyr
 195 200 205

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His	Pro	Asp	His	Phe	Asn	Cys	Thr	His	Cys	Gly	Lys	Glu	Leu	Thr	Ala	
210						215					220					
Glu	Ala	Arg	Glu	Leu	Lys	Gly	Glu	Leu	Tyr	Cys	Leu	Pro	Cys	His	Asp	
225					230					235					240	
Lys	Met	Gly	Val	Pro	Ile	Cys	Gly	Ala	Cys	Arg	Arg	Pro	Ile	Glu	Gly	
				245					250					255		
Arg	Val	Val	Asn	Ala	Leu	Gly	Lys	Gln	Trp	His	Val	Glu	His	Phe	Val	
			260					265					270			
Cys	Ala	Lys	Cys	Glu	Lys	Pro	Phe	Leu	Gly	His	Arg	His	Tyr	Glu	Lys	
	275						280					285				
Lys	Gly	Leu	Ala	Tyr	Cys	Glu	Leu									
290						295										

<210> 103

<211> 500

<212> PRT

<213> Homo sapiens

<400> 103

Met	Gly	Ile	Gly	Leu	Ser	Ala	Gln	Gly	Val	Asn	Met	Asn	Arg	Leu	Pro	
1				5					10					15		
Gly	Trp	Asp	Lys	His	Ser	Tyr	Gly	Tyr	His	Gly	Asp	Asp	Gly	His	Ser	
			20					25					30			
Phe	Cys	Ser	Ser	Gly	Thr	Gly	Gln	Pro	Tyr	Gly	Pro	Thr	Phe	Thr	Thr	
		35					40					45				
Gly	Asp	Val	Ile	Gly	Cys	Cys	Val	Asn	Leu	Ile	Asn	Asn	Thr	Cys	Phe	
	50					55					60					
Tyr	Thr	Lys	Asn	Gly	His	Ser	Leu	Gly	Ile	Ala	Phe	Thr	Asp	Leu	Pro	
65					70				75						80	
Pro	Asn	Leu	Tyr	Pro	Thr	Val	Gly	Leu	Gln	Thr	Pro	Gly	Glu	Val	Val	
				85					90					95		
Asp	Ala	Asn	Phe	Gly	Gln	His	Pro	Phe	Val	Phe	Asp	Ile	Glu	Asp	Tyr	
			100					105					110			
Met	Arg	Glu	Trp	Arg	Thr	Lys	Ile	Gln	Ala	Gln	Ile	Asp	Arg	Phe	Pro	
		115					120					125				
Ile	Gly	Asp	Arg	Glu	Gly	Glu	Trp	Gln	Thr	Met	Ile	Gln	Lys	Met	Val	
	130					135					140					
Ser	Ser	Tyr	Leu	Val	His	His	Gly	Tyr	Cys	Ala	Thr	Ala	Glu	Ala	Phe	
145					150					155					160	
Ala	Arg	Ser	Thr	Asp	Gln	Thr	Val	Leu	Glu	Glu	Leu	Ala	Ser	Ile	Lys	
				165					170					175		
Asn	Arg	Gln	Arg	Ile	Gln	Lys	Leu	Val	Leu	Ala	Gly	Arg	Met	Gly	Glu	
			180					185					190			
Ala	Ile	Glu	Thr	Thr	Gln	Gln	Leu	Tyr	Pro	Ser	Leu	Leu	Glu	Arg	Asn	
		195					200					205				
Pro	Asn	Leu	Leu	Phe	Thr	Leu	Lys	Val	Arg	Gln	Phe	Ile	Glu	Met	Val	
	210					215					220					
Asn	Gly	Thr	Asp	Ser	Glu	Val	Arg	Cys	Leu	Gly	Gly	Arg	Ser	Pro	Lys	
225					230					235					240	
Ser	Gln	Asp	Ser	Tyr	Pro	Val	Ser	Pro	Arg	Pro	Phe	Ser	Ser	Pro	Ser	
				245					250					255		
Met	Ser	Pro	Ser	His	Gly	Met	Asn	Ile	His	Asn	Leu	Ala	Ser	Gly	Lys	
			260					265					270			
Gly	Ser	Thr	Ala	His	Phe	Ser	Gly	Phe	Glu	Ser	Cys	Ser	Asn	Gly	Val	
		275					280					285				
Ile	Ser	Asn	Lys	Ala	His	Gln	Ser	Tyr	Cys	His	Ser	Asn	Lys	His	Gln	
290						295					300					

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Ser Ser Asn Leu Asn Val Pro Glu Leu Asn Ser Ile Asn Met Ser Arg
 305 310 315 320
 Ser Gln Gln Val Asn Asn Phe Thr Ser Asn Asp Val Asp Met Glu Thr
 325 330 335
 Asp His Tyr Ser Asn Gly Val Gly Glu Thr Ser Ser Asn Gly Phe Leu
 340 345 350
 Asn Gly Ser Ser Lys His Asp His Glu Met Glu Asp Cys Asp Thr Glu
 355 360 365
 Met Glu Val Asp Ser Ser Gln Leu Arg Arg Gln Leu Cys Gly Gly Ser
 370 375 380
 Gln Ala Ala Ile Glu Arg Met Ile His Phe Gly Arg Glu Leu Gln Ala
 385 390 395 400
 Met Ser Glu Gln Leu Arg Arg Asp Cys Gly Lys Asn Thr Ala Asn Lys
 405 410 415
 Lys Met Leu Lys Asp Ala Phe Ser Leu Leu Ala Tyr Ser Asp Pro Trp
 420 425 430
 Asn Ser Pro Val Gly Asn Gln Leu Asp Pro Ile Gln Arg Glu Pro Val
 435 440 445
 Cys Ser Ala Leu Asn Ser Ala Ile Leu Glu Thr His Asn Leu Pro Lys
 450 455 460
 Gln Pro Pro Leu Ala Leu Ala Met Gly Gln Ala Thr Gln Cys Leu Gly
 465 470 475 480
 Leu Met Ala Arg Ser Gly Ile Gly Ser Cys Ala Phe Ala Thr Val Glu
 485 490 495
 Asp Tyr Leu His
 500

<210> 104

<211> 387

<212> PRT

<213> Homo sapiens

<400> 104

Met Ala Thr Ser Gly Val Leu Pro Gly Gly Gly Phe Val Ala Ser Ala
 1 5 10 15
 Ala Ala Val Ala Gly Pro Glu Met Gln Thr Gly Arg Asn Asn Phe Val
 20 25 30
 Ile Arg Arg Asn Pro Ala Asp Pro Gln Arg Ile Pro Ser Asn Pro Ser
 35 40 45
 His Arg Ile Gln Cys Ala Ala Gly Tyr Glu Gln Ser Glu His Asn Val
 50 55 60
 Cys Gln Asp Ile Asp Glu Cys Thr Ala Gly Thr His Asn Cys Arg Ala
 65 70 75 80
 Asp Gln Val Cys Ile Asn Leu Arg Gly Ser Phe Ala Cys Gln Cys Pro
 85 90 95
 Pro Gly Tyr Gln Lys Arg Gly Glu Gln Cys Val Asp Ile Asp Glu Cys
 100 105 110
 Thr Ile Pro Pro Tyr Cys His Gln Arg Cys Val Asn Thr Pro Gly Ser
 115 120 125
 Phe Tyr Cys Gln Cys Ser Pro Gly Phe Gln Leu Ala Ala Asn Asn Tyr
 130 135 140
 Thr Cys Val Asp Ile Asn Glu Cys Asp Ala Ser Asn Gln Cys Ala Gln
 145 150 155 160
 Gln Cys Tyr Asn Ile Leu Gly Ser Phe Ile Cys Gln Cys Asn Gln Gly
 165 170 175
 Tyr Glu Leu Ser Ser Asp Arg Leu Asn Cys Glu Asp Ile Asp Glu Cys
 180 185 190

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Arg Thr Ser Ser Tyr Leu Cys Gln Tyr Gln Cys Val Asn Glu Pro Gly
 195 200 205
 Lys Phe Ser Cys Met Cys Pro Gln Gly Tyr Gln Val Val Arg Ser Arg
 210 215 220
 Thr Cys Gln Asp Ile Asn Glu Cys Glu Thr Thr Asn Glu Cys Arg Glu
 225 230 235 240
 Asp Glu Met Cys Trp Asn Tyr His Gly Gly Phe Arg Cys Tyr Pro Arg
 245 250 255
 Asn Pro Cys Gln Asp Pro Tyr Ile Leu Thr Pro Glu Asn Arg Cys Val
 260 265 270
 Cys Pro Val Ser Asn Ala Met Cys Arg Glu Leu Pro Gln Ser Ile Val
 275 280 285
 Tyr Lys Tyr Met Ser Ile Arg Ser Asp Arg Ser Val Pro Ser Asp Ile
 290 295 300
 Phe Gln Ile Gln Ala Thr Thr Ile Tyr Ala Asn Thr Ile Asn Thr Phe
 305 310 315 320
 Arg Ile Lys Ser Gly Asn Glu Asn Gly Glu Phe Tyr Leu Arg Gln Thr
 325 330 335
 Ser Pro Val Ser Ala Met Leu Val Leu Val Lys Ser Leu Ser Gly Pro
 340 345 350
 Arg Glu His Ile Val Asp Leu Glu Met Leu Thr Val Ser Ser Ile Gly
 355 360 365
 Thr Phe Arg Thr Ser Ser Val Leu Arg Leu Thr Ile Ile Val Gly Pro
 370 375 380
 Phe Ser Phe
 385

<210> 105

<211> 531

<212> PRT

<213> Homo sapiens

<400> 105

Met Ser Lys Pro His Ser Glu Ala Gly Thr Ala Phe Ile Gln Thr Gln
 1 5 10 15
 Gln Leu His Ala Ala Met Ala Asp Thr Phe Leu Glu His Met Cys Arg
 20 25 30
 Leu Asp Ile Asp Ser Pro Pro Ile Thr Ala Arg Asn Thr Gly Ile Ile
 35 40 45
 Cys Thr Ile Gly Pro Ala Ser Arg Ser Val Glu Thr Leu Lys Glu Met
 50 55 60
 Ile Lys Ser Gly Met Asn Val Ala Arg Leu Asn Phe Ser His Gly Thr
 65 70 75 80
 His Glu Tyr His Ala Glu Thr Ile Lys Asn Val Arg Thr Ala Thr Glu
 85 90 95
 Ser Phe Ala Ser Asp Pro Tyr Leu Tyr Arg Pro Val Ala Val Ala Leu
 100 105 110
 Asp Thr Lys Gly Pro Glu Ile Arg Thr Gly Leu Ile Lys Gly Ser Gly
 115 120 125
 Thr Ala Glu Leu Glu Leu Lys Lys Gly Ala Thr Leu Lys Ile Thr Leu
 130 135 140
 Asp Asn Ala Tyr Met Glu Lys Cys Asp Glu Asn Ile Leu Trp Leu Asp
 145 150 155 160
 Tyr Lys Asn Ile Cys Lys Val Val Glu Val Gly Ser Lys Ile Tyr Val
 165 170 175
 Asp Asp Gly Leu Ile Ser Leu Gln Val Lys Gln Lys Gly Ala Asp Phe
 180 185 190

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Leu Val Thr Glu Val Glu Asn Gly Gly Ser Leu Gly Ser Lys Lys Gly
195 200 205
Val Asn Leu Pro Gly Ala Ala Val Asp Leu Pro Ala Val Ser Glu Lys
210 215 220
Asp Ile Gln Asp Leu Lys Phe Gly Val Glu Gln Asp Val Asp Met Val
225 230 235 240
Phe Ala Ser Phe Ile Arg Lys Ala Ser Asp Val His Glu Val Arg Lys
245 250 255
Val Leu Gly Glu Lys Gly Lys Asn Ile Lys Ile Ile Ser Lys Ile Glu
260 265 270
Asn His Glu Gly Val Arg Arg Phe Asp Glu Ile Leu Glu Ala Ser Asp
275 280 285
Gly Ile Met Val Ala Arg Gly Asp Leu Gly Ile Glu Ile Pro Ala Glu
290 295 300
Lys Val Phe Leu Ala Gln Lys Met Met Ile Gly Arg Cys Asn Arg Ala
305 310 315 320
Gly Lys Pro Val Ile Cys Ala Thr Gln Met Leu Glu Ser Met Ile Lys
325 330 335
Lys Pro Arg Pro Thr Arg Ala Glu Gly Ser Asp Val Ala Asn Ala Val
340 345 350
Leu Asp Gly Ala Asp Cys Ile Met Leu Ser Gly Glu Thr Ala Lys Gly
355 360 365
Asp Tyr Pro Leu Glu Ala Val Arg Met Gln His Leu Ile Ala Arg Glu
370 375 380
Ala Glu Ala Ala Ile Tyr His Leu Gln Leu Phe Glu Glu Leu Arg Arg
385 390 395 400
Leu Ala Pro Ile Thr Ser Asp Pro Thr Glu Ala Thr Ala Val Gly Ala
405 410 415
Val Glu Ala Ser Phe Lys Cys Cys Ser Gly Ala Ile Ile Val Leu Thr
420 425 430
Lys Ser Gly Arg Ser Ala His Gln Val Ala Arg Tyr Arg Pro Arg Ala
435 440 445
Pro Ile Ile Ala Val Thr Arg Asn Pro Gln Thr Ala Arg Gln Ala His
450 455 460
Leu Tyr Arg Gly Ile Phe Pro Val Leu Cys Lys Asp Pro Val Gln Glu
465 470 475 480
Ala Trp Ala Glu Asp Val Asp Leu Arg Val Asn Phe Ala Met Asn Val
485 490 495
Gly Lys Ala Arg Gly Phe Phe Lys Lys Gly Asp Val Val Ile Val Leu
500 505 510
Thr Gly Trp Arg Pro Gly Ser Gly Phe Thr Asn Thr Met Arg Val Val
515 520 525
Pro Val Pro
530

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<210> 106

<211> 480

<212> PRT

<213> Homo sapiens

<400> 106

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Met Ala Ala Arg Cys Ser Thr Arg Trp Leu Leu Val Val Val Gly Thr
1 5 10 15
Pro Arg Leu Pro Ala Ile Ser Gly Arg Gly Ala Arg Pro Pro Arg Glu
20 25 30
Gly Val Val Gly Ala Trp Leu Ser Arg Lys Leu Ser Val Pro Ala Phe
35 40 45

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Ala	Ser	Ser	Leu	Thr	Ser	Cys	Gly	Pro	Arg	Ala	Leu	Leu	Thr	Leu	Arg
50						55					60				
Pro	Gly	Val	Ser	Leu	Thr	Gly	Thr	Lys	His	Asn	Pro	Phe	Ile	Cys	Thr
65					70					75					80
Ala	Ser	Phe	His	Thr	Ser	Ala	Pro	Leu	Ala	Lys	Glu	Asp	Tyr	Tyr	Gln
			85						90					95	
Ile	Leu	Gly	Val	Pro	Arg	Asn	Ala	Ser	Gln	Lys	Glu	Ile	Lys	Lys	Ala
			100					105					110		
Tyr	Tyr	Gln	Leu	Ala	Lys	Lys	Tyr	His	Pro	Asp	Thr	Asn	Lys	Asp	Asp
		115					120					125			
Pro	Lys	Ala	Lys	Glu	Lys	Phe	Ser	Gln	Leu	Ala	Glu	Ala	Tyr	Glu	Val
	130					135					140				
Leu	Ser	Asp	Glu	Val	Lys	Arg	Lys	Gln	Tyr	Asp	Ala	Tyr	Gly	Ser	Ala
145					150					155					160
Gly	Phe	Asp	Pro	Gly	Ala	Ser	Gly	Ser	Gln	His	Ser	Tyr	Trp	Lys	Gly
			165						170					175	
Gly	Pro	Thr	Val	Asp	Pro	Glu	Glu	Leu	Phe	Arg	Lys	Ile	Phe	Gly	Glu
			180					185					190		
Phe	Ser	Ser	Ser	Ser	Phe	Gly	Asp	Phe	Gln	Thr	Val	Phe	Asp	Gln	Pro
		195					200					205			
Gln	Glu	Tyr	Phe	Met	Glu	Leu	Thr	Phe	Asn	Gln	Ala	Ala	Lys	Gly	Val
	210					215					220				
Asn	Lys	Glu	Phe	Thr	Val	Asn	Ile	Met	Asp	Thr	Cys	Glu	Arg	Cys	Asn
225					230					235					240
Gly	Lys	Gly	Asn	Glu	Pro	Gly	Thr	Lys	Val	Gln	His	Cys	His	Tyr	Cys
			245						250					255	
Gly	Gly	Ser	Gly	Met	Glu	Thr	Ile	Asn	Thr	Gly	Pro	Phe	Val	Met	Arg
			260					265					270		
Ser	Thr	Cys	Arg	Arg	Cys	Gly	Gly	Arg	Gly	Ser	Ile	Ile	Ile	Ser	Pro
		275					280					285			
Cys	Val	Val	Cys	Arg	Gly	Ala	Gly	Gln	Ala	Lys	Gln	Lys	Lys	Arg	Val
	290					295					300				
Met	Ile	Pro	Val	Pro	Ala	Gly	Val	Glu	Asp	Gly	Gln	Thr	Val	Arg	Met
305					310				315						320
Pro	Val	Gly	Lys	Arg	Glu	Ile	Phe	Ile	Thr	Phe	Arg	Val	Gln	Lys	Ser
			325						330					335	
Pro	Val	Phe	Arg	Arg	Asp	Gly	Ala	Asp	Ile	His	Ser	Asp	Leu	Phe	Ile
			340					345					350		
Ser	Ile	Ala	Gln	Ala	Leu	Leu	Gly	Gly	Thr	Ala	Arg	Ala	Gln	Gly	Leu
		355					360					365			
Tyr	Glu	Thr	Ile	Asn	Val	Thr	Ile	Pro	Pro	Gly	Thr	Gln	Thr	Asp	Gln
	370					375					380				
Lys	Ile	Arg	Met	Gly	Gly	Lys	Gly	Ile	Pro	Arg	Ile	Asn	Ser	Tyr	Gly
385					390					395					400
Tyr	Gly	Asp	His	Tyr	Ile	His	Ile	Lys	Ile	Arg	Val	Pro	Lys	Arg	Leu
			405						410					415	
Thr	Ser	Arg	Gln	Gln	Ser	Leu	Ile	Leu	Ser	Tyr	Ala	Glu	Asp	Glu	Thr
			420					425					430		
Asp	Val	Glu	Gly	Thr	Val	Asn	Gly	Val	Thr	Leu	Thr	Ser	Ser	Gly	Gly
	435						440					445			
Ser	Thr	Met	Asp	Ser	Ser	Ala	Gly	Ser	Lys	Ala	Arg	Arg	Glu	Ala	Gly
	450					455					460				
Glu	Asp	Glu	Glu	Gly	Phe	Leu	Ser	Lys	Leu	Lys	Lys	Met	Phe	Thr	Ser
465					470				475						480

<210> 107

<211> 572

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<212> PRT

<213> Homo sapiens

<400> 107

Met	Ala	Ala	Pro	Arg	Pro	Ser	Pro	Ala	Ile	Ser	Val	Ser	Val	Ser	Ala
1				5					10					15	
Pro	Ala	Phe	Tyr	Ala	Pro	Gln	Lys	Lys	Phe	Gly	Pro	Val	Val	Ala	Pro
			20					25					30		
Lys	Pro	Lys	Val	Asn	Pro	Phe	Arg	Pro	Gly	Asp	Ser	Glu	Pro	Pro	Pro
		35					40					45			
Ala	Pro	Gly	Ala	Gln	Arg	Ala	Gln	Met	Gly	Arg	Val	Gly	Glu	Ile	Pro
	50					55					60				
Pro	Pro	Pro	Pro	Glu	Asp	Phe	Pro	Leu	Pro	Pro	Pro	Pro	Leu	Ala	Gly
65					70					75					80
Asp	Gly	Asp	Asp	Ala	Glu	Gly	Ala	Leu	Gly	Gly	Ala	Phe	Pro	Pro	Pro
				85					90					95	
Pro	Pro	Pro	Ile	Glu	Glu	Ser	Phe	Pro	Pro	Ala	Pro	Leu	Glu	Glu	Glu
			100					105					110		
Ile	Phe	Pro	Ser	Pro	Pro	Pro	Pro	Pro	Glu	Glu	Glu	Gly	Gly	Pro	Glu
		115					120						125		
Ala	Pro	Ile	Pro	Pro	Pro	Pro	Gln	Pro	Arg	Glu	Lys	Val	Ser	Ser	Ile
	130						135					140			
Asp	Leu	Glu	Ile	Asp	Ser	Leu	Ser	Ser	Leu	Leu	Asp	Asp	Met	Thr	Lys
145					150					155					160
Asn	Asp	Pro	Phe	Lys	Ala	Arg	Val	Ser	Ser	Gly	Tyr	Val	Pro	Pro	Pro
				165					170					175	
Val	Ala	Thr	Pro	Phe	Ser	Ser	Lys	Ser	Ser	Thr	Lys	Pro	Ala	Ala	Gly
			180					185					190		
Gly	Thr	Ala	Pro	Leu	Pro	Pro	Trp	Lys	Ser	Pro	Ser	Ser	Ser	Gln	Pro
		195					200					205			
Leu	Pro	Gln	Val	Pro	Ala	Pro	Ala	Gln	Ser	Gln	Thr	Gln	Phe	His	Val
	210					215					220				
Gln	Pro	Gln	Pro	Gln	Pro	Lys	Pro	Gln	Val	Gln	Leu	His	Val	Gln	Ser
225					230					235					240
Gln	Thr	Gln	Pro	Val	Ser	Leu	Ala	Asn	Thr	Gln	Pro	Arg	Gly	Pro	Pro
				245					250					255	
Ala	Ser	Ser	Pro	Ala	Pro	Ala	Pro	Lys	Phe	Ser	Pro	Val	Thr	Pro	Lys
			260					265					270		
Phe	Thr	Pro	Val	Ala	Ser	Lys	Phe	Ser	Pro	Gly	Ala	Pro	Gly	Gly	Ser
		275					280					285			
Gly	Ser	Gln	Pro	Asn	Gln	Lys	Leu	Gly	His	Pro	Glu	Ala	Leu	Ser	Ala
	290					295					300				
Gly	Thr	Gly	Ser	Pro	Gln	Pro	Pro	Ser	Phe	Thr	Tyr	Ala	Gln	Gln	Arg
305					310					315					320
Glu	Lys	Pro	Arg	Val	Gln	Glu	Lys	Gln	His	Pro	Val	Pro	Pro	Pro	Ala
				325					330					335	
Gln	Asn	Gln	Asn	Gln	Val	Arg	Ser	Pro	Gly	Ala	Pro	Gly	Pro	Leu	Thr
			340					345					350		
Leu	Lys	Glu	Val	Glu	Glu	Leu	Glu	Gln	Leu	Thr	Gln	Gln	Leu	Met	Gln
		355					360					365			
Asp	Met	Glu	His	Pro	Gln	Arg	Gln	Asn	Val	Ala	Val	Asn	Glu	Leu	Cys
	370					375					380				
Gly	Arg	Cys	His	Gln	Pro	Leu	Ala	Arg	Ala	Gln	Pro	Ala	Val	Arg	Ala
385					390					395					400
Leu	Gly	Gln	Leu	Phe	His	Ile	Ala	Cys	Phe	Thr	Cys	His	Gln	Cys	Ala
				405					410					415	
Gln	Gln	Leu	Gln	Gly	Gln	Gln	Phe	Tyr	Ser	Leu	Glu	Gly	Ala	Pro	Tyr

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			420					425					430				
Cys	Glu	Gly	Cys	Tyr	Thr	Asp	Thr	Leu	Glu	Lys	Cys	Asn	Thr	Cys	Gly		
		435						440					445				
Glu	Pro	Ile	Thr	Asp	Arg	Met	Leu	Arg	Ala	Thr	Gly	Lys	Ala	Tyr	His		
	450					455					460						
Pro	His	Cys	Phe	Thr	Cys	Val	Val	Cys	Ala	Arg	Pro	Leu	Glu	Gly	Thr		
465					470					475					480		
Ser	Phe	Ile	Val	Asp	Gln	Ala	Asn	Arg	Pro	His	Cys	Val	Pro	Asp	Tyr		
				485					490					495			
His	Lys	Gln	Tyr	Ala	Pro	Arg	Cys	Ser	Val	Cys	Ser	Glu	Pro	Ile	Met		
			500					505					510				
Pro	Glu	Pro	Gly	Arg	Asp	Glu	Thr	Val	Arg	Val	Val	Ala	Leu	Asp	Lys		
		515					520					525					
Asn	Phe	His	Met	Lys	Cys	Tyr	Lys	Cys	Glu	Asp	Cys	Gly	Lys	Pro	Leu		
	530					535					540						
Ser	Ile	Glu	Ala	Asp	Asp	Asn	Gly	Cys	Phe	Pro	Leu	Asp	Gly	His	Val		
545					550					555					560		
Leu	Cys	Arg	Lys	Cys	His	Thr	Ala	Arg	Ala	Gln	Thr						
			565					570									

<210> 108

<211> 2861

<212> PRT

<213> Homo sapiens

<400> 108

Met	Lys	Ala	Met	Asp	Val	Leu	Pro	Ile	Leu	Lys	Glu	Lys	Val	Ala	Tyr		
1			5						10					15			
Leu	Ser	Gly	Gly	Arg	Asp	Lys	Arg	Gly	Gly	Pro	Ile	Leu	Thr	Phe	Pro		
		20						25					30				
Ala	Arg	Ser	Asn	His	Asp	Arg	Ile	Arg	Gln	Glu	Asp	Leu	Arg	Arg	Leu		
		35					40					45					
Ile	Ser	Tyr	Leu	Ala	Cys	Ile	Pro	Ser	Glu	Glu	Val	Cys	Lys	Arg	Gly		
	50					55					60						
Phe	Thr	Val	Ile	Val	Asp	Met	Arg	Gly	Ser	Lys	Trp	Asp	Ser	Ile	Lys		
65					70					75					80		
Pro	Leu	Leu	Lys	Ile	Leu	Gln	Glu	Ser	Phe	Pro	Cys	Cys	Ile	His	Val		
			85						90					95			
Ala	Leu	Ile	Ile	Lys	Pro	Asp	Asn	Phe	Trp	Gln	Lys	Gln	Arg	Thr	Asn		
			100					105					110				
Phe	Gly	Ser	Ser	Lys	Phe	Glu	Phe	Glu	Thr	Asn	Met	Val	Ser	Leu	Glu		
	115						120					125					
Gly	Leu	Thr	Lys	Val	Val	Asp	Pro	Ser	Gln	Leu	Thr	Pro	Glu	Phe	Asp		
	130					135					140						
Gly	Cys	Leu	Glu	Tyr	Asn	His	Glu	Glu	Trp	Ile	Glu	Ile	Arg	Val	Ala		
145					150					155					160		
Phe	Glu	Asp	Tyr	Ile	Ser	Asn	Ala	Thr	His	Met	Leu	Ser	Arg	Leu	Glu		
			165					170						175			
Glu	Leu	Gln	Asp	Ile	Leu	Ala	Lys	Lys	Glu	Leu	Pro	Gln	Asp	Leu	Glu		
		180						185					190				
Gly	Ala	Arg	Asn	Met	Ile	Glu	Glu	His	Ser	Gln	Leu	Lys	Lys	Lys	Val		
	195						200					205					
Ile	Lys	Ala	Pro	Ile	Glu	Asp	Leu	Asp	Leu	Glu	Gly	Gln	Lys	Leu	Leu		
	210					215					220						
Gln	Arg	Ile	Gln	Ser	Ser	Glu	Ser	Phe	Pro	Lys	Lys	Asn	Ser	Gly	Ser		
225					230					235					240		
Gly	Asn	Ala	Asp	Leu	Gln	Asn	Leu	Leu	Pro	Lys	Val	Ser	Thr	Met	Leu		

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				245					250					255			
Asp	Arg	Leu	His	Ser	Thr	Arg	Gln	His	Leu	His	Gln	Met	Trp	His	Val		
			260					265					270				
Arg	Lys	Leu	Lys	Leu	Asp	Gln	Cys	Phe	Gln	Leu	Arg	Leu	Phe	Glu	Gln		
		275					280					285					
Asp	Ala	Glu	Lys	Met	Phe	Asp	Trp	Ile	Thr	His	Asn	Lys	Gly	Leu	Phe		
	290					295					300						
Leu	Asn	Ser	Tyr	Thr	Glu	Ile	Gly	Thr	Ser	His	Pro	His	Ala	Met	Glu		
305					310					315					320		
Leu	Gln	Thr	Gln	His	Asn	His	Phe	Ala	Met	Asn	Cys	Met	Asn	Val	Tyr		
				325					330					335			
Val	Asn	Ile	Asn	Arg	Ile	Met	Ser	Val	Ala	Asn	Arg	Leu	Val	Glu	Ser		
			340					345					350				
Gly	His	Tyr	Ala	Ser	Gln	Gln	Ile	Arg	Gln	Ile	Ala	Ser	Gln	Leu	Glu		
		355					360					365					
Gln	Glu	Trp	Lys	Ala	Phe	Ala	Ala	Ala	Leu	Asp	Glu	Arg	Ser	Thr	Leu		
	370					375					380						
Leu	Asp	Met	Ser	Ser	Ile	Phe	His	Gln	Lys	Ala	Glu	Lys	Tyr	Met	Ser		
385					390					395					400		
Asn	Val	Asp	Ser	Trp	Cys	Lys	Ala	Cys	Gly	Glu	Val	Asp	Leu	Pro	Ser		
				405					410					415			
Glu	Leu	Gln	Asp	Leu	Glu	Asp	Ala	Ile	His	His	His	Gln	Gly	Ile	Tyr		
			420					425					430				
Glu	His	Ile	Thr	Leu	Ala	Tyr	Ser	Glu	Val	Ser	Gln	Asp	Gly	Lys	Ser		
		435					440					445					
Leu	Leu	Asp	Lys	Leu	Gln	Arg	Pro	Leu	Thr	Pro	Gly	Ser	Ser	Asp	Ser		
	450					455					460						
Leu	Thr	Ala	Ser	Ala	Asn	Tyr	Ser	Lys	Ala	Val	His	His	Val	Leu	Asp		
465					470					475					480		
Val	Ile	His	Glu	Val	Leu	His	His	Gln	Arg	His	Val	Arg	Thr	Ile	Trp		
				485					490					495			
Gln	His	Arg	Lys	Val	Arg	Leu	His	Gln	Arg	Leu	Gln	Leu	Cys	Val	Phe		
			500					505					510				
Gln	Gln	Glu	Val	Gln	Gln	Val	Leu	Asp	Trp	Ile	Glu	Asn	His	Gly	Glu		
		515					520					525					
Ala	Phe	Leu	Ser	Lys	His	Thr	Gly	Val	Gly	Lys	Ser	Leu	His	Arg	Ala		
	530					535					540						
Arg	Ala	Leu	Gln	Lys	Arg	His	Glu	Asp	Phe	Glu	Glu	Val	Ala	Gln	Asn		
545					550					555					560		
Thr	Tyr	Thr	Asn	Ala	Asp	Lys	Leu	Leu	Glu	Ala	Ala	Glu	Gln	Leu	Ala		
				565					570					575			
Gln	Thr	Gly	Glu	Cys	Asp	Pro	Glu	Glu	Ile	Tyr	Gln	Ala	Ala	His	Gln		
		580						585					590				
Leu	Glu	Asp	Arg	Ile	Gln	Asp	Phe	Val	Arg	Arg	Val	Glu	Gln	Arg	Lys		
		595				600						605					
Ile	Leu	Leu	Asp	Met	Ser	Val	Ser	Phe	His	Thr	His	Val	Lys	Glu	Leu		
	610					615					620						
Trp	Thr	Trp	Leu	Glu	Glu	Leu	Gln	Lys	Glu	Leu	Leu	Asp	Asp	Val	Tyr		
625				630						635					640		
Ala	Glu	Ser	Val	Glu	Ala	Val	Gln	Asp	Leu	Ile	Lys	Arg	Phe	Gly	Gln		
				645					650					655			
Gln	Gln	Gln	Thr	Thr	Leu	Gln	Val	Thr	Val	Asn	Val	Ile	Lys	Glu	Gly		
			660					665					670				
Glu	Asp	Leu	Ile	Gln	Gln	Leu	Arg	Asp	Ser	Ala	Ile	Ser	Ser	Asn	Lys		
		675					680					685					
Thr	Pro	His	Asn	Ser	Ser	Ile	Asn	His	Ile	Glu	Thr	Val	Leu	Gln	Gln		
	690					695					700						

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Leu	Asp	Glu	Ala	Gln	Ser	Gln	Met	Glu	Glu	Leu	Phe	Gln	Glu	Arg	Lys
705					710					715					720
Ile	Lys	Leu	Glu	Leu	Phe	Leu	His	Val	Arg	Ile	Phe	Glu	Arg	Asp	Ala
				725					730					735	
Ile	Asp	Ile	Ile	Ser	Asp	Leu	Glu	Ser	Trp	Asn	Asp	Glu	Leu	Ser	Gln
			740					745					750		
Gln	Met	Asn	Asp	Phe	Asp	Thr	Glu	Asp	Leu	Thr	Ile	Ala	Glu	Gln	Arg
		755					760					765			
Leu	Gln	His	His	Ala	Asp	Lys	Ala	Leu	Thr	Met	Asn	Asn	Leu	Thr	Phe
	770					775					780				
Asp	Val	Ile	His	Gln	Gly	Gln	Asp	Leu	Leu	Gln	Tyr	Val	Asn	Glu	Val
785					790					795					800
Gln	Ala	Ser	Gly	Val	Glu	Leu	Leu	Cys	Asp	Arg	Asp	Val	Asp	Met	Ala
				805					810					815	
Thr	Arg	Val	Gln	Asp	Leu	Leu	Glu	Phe	Leu	His	Glu	Lys	Gln	Gln	Glu
			820					825						830	
Leu	Asp	Leu	Ala	Ala	Glu	Gln	His	Arg	Lys	His	Leu	Glu	Gln	Cys	Val
		835					840					845			
Gln	Leu	Arg	His	Leu	Gln	Ala	Glu	Val	Lys	Gln	Val	Leu	Gly	Trp	Ile
	850					855					860				
Arg	Asn	Gly	Glu	Ser	Met	Leu	Asn	Ala	Gly	Leu	Ile	Thr	Ala	Ser	Ser
865					870					875					880
Leu	Gln	Glu	Ala	Glu	Gln	Leu	Gln	Arg	Glu	His	Glu	Gln	Phe	Gln	His
				885					890					895	
Ala	Ile	Glu	Lys	Thr	His	Gln	Ser	Ala	Leu	Gln	Val	Gln	Gln	Lys	Ala
			900					905					910		
Glu	Ala	Met	Leu	Gln	Ala	Asn	His	Tyr	Asp	Met	Asp	Met	Ile	Arg	Asp
		915					920					925			
Cys	Ala	Glu	Lys	Val	Ala	Ser	His	Trp	Gln	Gln	Leu	Met	Leu	Lys	Met
	930					935					940				
Glu	Asp	Arg	Leu	Lys	Leu	Val	Asn	Ala	Ser	Val	Ala	Phe	Tyr	Lys	Thr
945					950					955					960
Ser	Glu	Gln	Val	Cys	Ser	Val	Leu	Glu	Ser	Leu	Glu	Gln	Glu	Tyr	Lys
				965					970					975	
Arg	Glu	Glu	Asp	Trp	Cys	Gly	Gly	Ala	Asp	Lys	Leu	Gly	Pro	Asn	Ser
			980					985					990		
Glu	Thr	Asp	His	Val	Thr	Pro	Met	Ile	Ser	Lys	His	Leu	Glu	Gln	Lys
		995					1000					1005			
Glu	Ala	Phe	Leu	Lys	Ala	Cys	Thr	Leu	Ala	Arg	Arg	Asn	Ala	Asp	Val
	1010					1015					1020				
Phe	Leu	Lys	Tyr	Leu	His	Arg	Asn	Ser	Val	Asn	Met	Pro	Gly	Met	Val
1025					1030					1035					1040
Thr	His	Ile	Lys	Ala	Pro	Glu	Gln	Gln	Val	Lys	Asn	Ile	Leu	Asn	Glu
				1045					1050					1055	
Leu	Phe	Gln	Arg	Glu	Asn	Arg	Val	Leu	His	Tyr	Trp	Thr	Met	Arg	Lys
			1060					1065					1070		
Arg	Arg	Leu	Asp	Gln	Cys	Gln	Gln	Tyr	Val	Val	Phe	Glu	Arg	Ser	Ala
		1075					1080					1085			
Lys	Gln	Ala	Leu	Glu	Trp	Ile	His	Asp	Asn	Gly	Glu	Phe	Tyr	Leu	Ser
	1090					1095					1100				
Thr	His	Thr	Ser	Thr	Gly	Ser	Ser	Ile	Gln	His	Thr	Gln	Glu	Leu	Leu
1105					1110					1115					1120
Lys	Glu	His	Glu	Glu	Phe	Gln	Ile	Thr	Ala	Lys	Gln	Thr	Lys	Glu	Arg
				1125					1130					1135	
Val	Lys	Leu	Leu	Ile	Gln	Leu	Ala	Asp	Gly	Phe	Cys	Glu	Lys	Gly	His
			1140					1145					1150		
Ala	His	Ala	Ala	Glu	Ile	Lys	Lys	Cys	Val	Thr	Ala	Val	Asp	Lys	Arg

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1155	1160	1165
Tyr Arg Asp Phe Ser Leu	Arg Met Glu Lys Tyr	Arg Thr Ser Leu Glu
1170	1175	1180
Lys Ala Leu Gly Ile Ser	Ser Asp Ser Asn Lys	Ser Ser Lys Ser Leu
1185	1190	1195
Gln Leu Asp Ile Ile Pro	Ala Ser Ile Pro Gly	Ser Glu Val Lys Leu
1205	1210	1215
Arg Asp Ala Ala His Glu	Leu Asn Glu Glu Lys	Arg Lys Ser Ala Arg
1220	1225	1230
Arg Lys Glu Phe Ile Met	Ala Glu Leu Ile Gln	Thr Glu Lys Ala Tyr
1235	1240	1245
Val Arg Asp Leu Arg Glu	Cys Met Asp Thr Tyr	Leu Trp Glu Met Thr
1250	1255	1260
Ser Gly Val Glu Glu Ile	Pro Pro Gly Ile Val	Asn Lys Glu Leu Ile
1265	1270	1275
Ile Phe Gly Asn Met Gln	Glu Ile Tyr Glu Phe	His Asn Asn Ile Phe
1285	1290	1295
Leu Lys Glu Leu Glu Lys	Tyr Glu Gln Leu Pro	Glu Asp Val Gly His
1300	1305	1310
Cys Phe Val Thr Trp Ala	Asp Lys Phe Gln Met	Tyr Val Thr Tyr Cys
1315	1320	1325
Lys Asn Lys Pro Asp Ser	Thr Gln Leu Ile Leu	Glu His Ala Gly Ser
1330	1335	1340
Tyr Phe Asp Glu Ile Gln	Gln Arg His Gly Leu	Ala Asn Ser Ile Ser
1345	1350	1355
Ser Tyr Leu Ile Lys Pro	Val Gln Arg Ile Thr	Lys Tyr Gln Leu Leu
1365	1370	1375
Leu Lys Glu Leu Leu Thr	Cys Cys Glu Glu Gly	Lys Gly Glu Ile Lys
1380	1385	1390
Asp Gly Leu Glu Val Met	Leu Ser Val Pro Lys	Arg Ala Asn Asp Ala
1395	1400	1405
Met His Leu Ser Met Leu	Glu Gly Phe Asp Glu	Asn Ile Glu Ser Gln
1410	1415	1420
Gly Glu Leu Ile Leu Gln	Glu Ser Phe Gln Val	Trp Asp Pro Lys Thr
1425	1430	1435
Leu Ile Arg Lys Gly Arg	Glu Arg His Leu Phe	Leu Phe Glu Met Ser
1445	1450	1455
Leu Val Phe Ser Lys Glu	Val Lys Asp Ser Ser	Gly Arg Ser Lys Tyr
1460	1465	1470
Leu Tyr Lys Ser Lys Leu	Phe Thr Ser Glu Leu	Gly Val Thr Glu His
1475	1480	1485
Val Glu Gly Asp Pro Cys	Lys Phe Ala Leu Trp	Val Gly Arg Thr Pro
1490	1495	1500
Thr Ser Asp Asn Lys Ile	Val Leu Lys Ala Ser	Ser Ile Glu Asn Lys
1505	1510	1515
Gln Asp Trp Ile Lys His	Ile Arg Glu Val Ile	Gln Glu Arg Thr Ile
1525	1530	1535
His Leu Lys Gly Ala Leu	Lys Glu Pro Ile His	Ile Pro Lys Thr Ala
1540	1545	1550
Pro Ala Thr Arg Gln Lys	Gly Arg Arg Asp Gly	Glu Asp Leu Asp Ser
1555	1560	1565
Gln Gly Asp Gly Ser Ser	Gln Pro Asp Thr Ile	Ser Ile Ala Ser Arg
1570	1575	1580
Thr Ser Gln Asn Thr Leu	Asp Ser Asp Lys Leu	Ser Gly Gly Cys Glu
1585	1590	1595
Leu Thr Val Val Ile His	Asp Phe Thr Ala Cys	Asn Ser Asn Glu Leu
1605	1610	1615

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Thr Ile Arg Arg Gly Gln Thr Val Glu Val Leu Glu Arg Pro His Asp
 1620 1625 1630
 Lys Pro Asp Trp Cys Leu Val Arg Thr Thr Asp Arg Ser Pro Ala Ala
 1635 1640 1645
 Glu Gly Leu Val Pro Cys Gly Ser Leu Cys Ile Ala His Ser Arg Ser
 1650 1655 1660
 Ser Met Glu Met Glu Gly Ile Phe Asn His Lys Asp Ser Leu Ser Val
 1665 1670 1675 1680
 Ser Ser Asn Asp Ala Ser Pro Pro Ala Ser Val Ala Ser Leu Gln Pro
 1685 1690 1695
 His Met Ile Gly Ala Gln Ser Ser Pro Gly Pro Lys Arg Pro Gly Asn
 1700 1705 1710
 Thr Leu Arg Lys Trp Leu Thr Ser Pro Val Arg Arg Leu Ser Ser Gly
 1715 1720 1725
 Lys Ala Asp Gly His Val Lys Lys Leu Ala His Lys His Lys Lys Ser
 1730 1735 1740
 Arg Glu Val Arg Lys Ser Ala Asp Ala Gly Ser Gln Lys Asp Ser Asp
 1745 1750 1755 1760
 Asp Ser Ala Ala Thr Pro Gln Asp Glu Thr Val Glu Glu Arg Gly Arg
 1765 1770 1775
 Asn Glu Gly Leu Ser Ser Gly Thr Leu Ser Lys Ser Ser Ser Ser Gly
 1780 1785 1790
 Met Gln Ser Cys Gly Glu Glu Glu Gly Glu Glu Gly Ala Asp Ala Val
 1795 1800 1805
 Pro Leu Pro Pro Pro Met Ala Ile Gln Gln His Ser Leu Leu Gln Pro
 1810 1815 1820
 Asp Ser Gln Asp Asp Lys Ala Ser Ser Arg Leu Leu Val Arg Pro Thr
 1825 1830 1835 1840
 Ser Ser Glu Thr Pro Ser Ala Ala Glu Leu Val Ser Ala Ile Glu Glu
 1845 1850 1855
 Leu Val Lys Ser Lys Met Ala Leu Glu Asp Arg Pro Ser Ser Leu Leu
 1860 1865 1870
 Val Asp Gln Gly Asp Ser Ser Ser Pro Ser Phe Asn Pro Ser Asp Asn
 1875 1880 1885
 Ser Leu Leu Ser Ser Ser Ser Pro Ile Asp Glu Met Glu Glu Arg Lys
 1890 1895 1900
 Ser Ser Ser Leu Lys Arg Arg His Tyr Val Leu Gln Glu Leu Val Glu
 1905 1910 1915 1920
 Thr Glu Arg Asp Tyr Val Arg Asp Leu Gly Tyr Val Val Glu Gly Tyr
 1925 1930 1935
 Met Ala Leu Met Lys Glu Asp Gly Val Pro Asp Asp Met Lys Gly Lys
 1940 1945 1950
 Asp Lys Ile Val Phe Gly Asn Ile His Gln Ile Tyr Asp Trp His Arg
 1955 1960 1965
 Asp Phe Phe Leu Gly Glu Leu Glu Lys Cys Leu Glu Asp Pro Glu Lys
 1970 1975 1980
 Leu Gly Ser Leu Phe Val Lys His Glu Arg Arg Leu His Met Tyr Ile
 1985 1990 1995 2000
 Ala Tyr Cys Gln Asn Lys Pro Lys Ser Glu His Ile Val Ser Glu Tyr
 2005 2010 2015
 Ile Asp Thr Phe Phe Glu Asp Leu Lys Gln Arg Leu Gly His Arg Leu
 2020 2025 2030
 Gln Leu Thr Asp Leu Leu Ile Lys Pro Val Gln Arg Ile Met Lys Tyr
 2035 2040 2045
 Gln Leu Leu Leu Lys Asp Phe Leu Lys Tyr Ser Lys Lys Ala Ser Leu
 2050 2055 2060
 Asp Thr Ser Glu Leu Glu Arg Ala Val Glu Val Met Cys Ile Val Pro

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2065		2070		2075		2080
Arg Arg Cys Asn Asp Met Met Asn Val Gly Arg Leu Gln Gly Phe Asp						
	2085		2090		2095	
Gly Lys Ile Val Ala Gln Gly Lys Leu Leu Leu Gln Asp Thr Phe Leu						
	2100		2105		2110	
Val Thr Asp Gln Asp Ala Gly Leu Leu Pro Arg Cys Arg Glu Arg Arg						
	2115		2120		2125	
Ile Phe Leu Phe Glu Gln Ile Val Ile Phe Ser Glu Pro Leu Asp Lys						
	2130		2135		2140	
Lys Lys Gly Phe Ser Met Pro Gly Phe Leu Phe Lys Asn Ser Ile Lys						
2145	2150		2155		2160	
Val Ser Cys Leu Cys Leu Glu Glu Asn Val Glu Asn Asp Pro Cys Lys						
	2165		2170		2175	
Phe Ala Leu Thr Ser Arg Thr Gly Asp Val Val Glu Thr Phe Ile Leu						
	2180		2185		2190	
His Ser Ser Ser Pro Ser Val Arg Gln Thr Trp Ile His Glu Ile Asn						
	2195		2200		2205	
Gln Ile Leu Glu Asn Gln Arg Asn Phe Leu Asn Ala Leu Thr Ser Pro						
	2210		2215		2220	
Ile Glu Tyr Gln Arg Asn His Ser Gly Gly Gly Gly Gly Gly Ser						
2225	2230		2235		2240	
Gly Ala Ala Ala Gly Val Gly Ala Ala Ala Ala Ala Gly Pro Pro Val						
	2245		2250		2255	
Ala Ala Ala Ala Thr Val Ala Ala Pro Ala Ala Ala Ala Ala Pro Pro						
	2260		2265		2270	
Ala Arg Ala Gly Ala Gly Pro Pro Gly Ser Pro Ser Leu Ser Asp Thr						
	2275		2280		2285	
Thr Pro Pro Cys Trp Ser Pro Leu Gln Pro Arg Ala Arg Gln Arg Gln						
	2290		2295		2300	
Thr Arg Cys Gln Ser Glu Ser Ser Ser Ser Ser Asn Ile Ser Thr Met						
2305	2310		2315		2320	
Leu Val Thr His Asp Tyr Thr Ala Val Lys Glu Asp Glu Ile Asn Val						
	2325		2330		2335	
Tyr Gln Gly Glu Val Val Gln Ile Leu Ala Ser Asn Gln Gln Asn Met						
	2340		2345		2350	
Phe Leu Val Phe Arg Ala Ala Thr Asp Gln Cys Pro Ala Ala Glu Gly						
	2355		2360		2365	
Trp Ile Pro Gly Phe Val Leu Gly His Thr Ser Ala Val Ile Val Glu						
	2370		2375		2380	
Asn Pro Asp Gly Thr Leu Lys Lys Ser Thr Ser Trp His Thr Ala Leu						
2385	2390		2395		2400	
Arg Leu Arg Lys Lys Ser Glu Lys Lys Asp Lys Asp Gly Lys Arg Glu						
	2405		2410		2415	
Gly Lys Leu Glu Asn Gly Tyr Arg Lys Ser Arg Glu Gly Leu Ser Asn						
	2420		2425		2430	
Lys Val Ser Val Lys Leu Leu Asn Pro Asn Tyr Ile Tyr Asp Val Pro						
	2435		2440		2445	
Pro Glu Phe Val Ile Pro Leu Ser Glu Val Thr Cys Glu Thr Gly Glu						
	2450		2455		2460	
Thr Val Val Leu Arg Cys Arg Val Cys Gly Arg Pro Lys Ala Ser Ile						
2465	2470		2475		2480	
Thr Trp Lys Gly Pro Glu His Asn Thr Leu Asn Asn Asp Gly His Tyr						
	2485		2490		2495	
Ser Ile Ser Tyr Ser Asp Leu Gly Glu Ala Thr Leu Lys Ile Val Gly						
	2500		2505		2510	
Val Thr Thr Glu Asp Asp Gly Ile Tyr Thr Cys Ile Ala Val Asn Asp						
	2515		2520		2525	

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Met Gly Ser Ala Ser Ser Ser Ala Ser Leu Arg Val Leu Gly Pro Gly
 2530 2535 2540
 Met Asp Gly Ile Met Val Thr Trp Lys Asp Asn Phe Asp Ser Phe Tyr
 2545 2550 2555 2560
 Ser Glu Val Ala Glu Leu Gly Arg Gly Arg Phe Ser Val Val Lys Lys
 2565 2570 2575
 Cys Asp Gln Lys Gly Thr Lys Arg Ala Val Ala Thr Lys Phe Val Asn
 2580 2585 2590
 Lys Lys Leu Met Lys Arg Asp Gln Val Thr His Glu Leu Gly Ile Leu
 2595 2600 2605
 Gln Ser Leu Gln His Pro Leu Leu Val Gly Leu Leu Asp Thr Phe Glu
 2610 2615 2620
 Thr Pro Thr Ser Tyr Ile Leu Val Leu Glu Met Ala Asp Gln Gly Arg
 2625 2630 2635 2640
 Leu Leu Asp Cys Val Val Arg Trp Gly Ser Leu Thr Glu Gly Lys Ile
 2645 2650 2655
 Arg Ala His Leu Gly Glu Val Leu Glu Ala Val Arg Tyr Leu His Asn
 2660 2665 2670
 Cys Arg Ile Ala His Leu Asp Leu Lys Pro Glu Asn Ile Leu Val Asp
 2675 2680 2685
 Glu Ser Leu Ala Lys Pro Thr Ile Lys Leu Ala Asp Phe Gly Asp Ala
 2690 2695 2700
 Val Gln Leu Asn Thr Thr Tyr Tyr Ile His Gln Leu Leu Gly Asn Pro
 2705 2710 2715 2720
 Glu Phe Ala Ala Pro Glu Ile Ile Leu Gly Asn Pro Val Ser Leu Thr
 2725 2730 2735
 Ser Asp Thr Trp Ser Val Gly Val Leu Thr Tyr Val Leu Leu Ser Gly
 2740 2745 2750
 Val Ser Pro Phe Leu Asp Asp Ser Val Glu Glu Thr Cys Leu Asn Ile
 2755 2760 2765
 Cys Arg Leu Asp Phe Ser Phe Pro Asp Asp Tyr Phe Lys Gly Val Ser
 2770 2775 2780
 Gln Lys Ala Lys Glu Phe Val Cys Phe Leu Leu Gln Glu Asp Pro Ala
 2785 2790 2795 2800
 Lys Arg Pro Ser Ala Ala Leu Ala Leu Gln Glu Gln Trp Leu Gln Ala
 2805 2810 2815
 Gly Asn Gly Arg Ser Thr Gly Val Leu Asp Thr Ser Arg Leu Thr Ser
 2820 2825 2830
 Phe Ile Glu Arg Arg Lys His Gln Asn Asp Val Arg Pro Ile Arg Ser
 2835 2840 2845
 Ile Lys Asn Phe Leu Gln Ser Arg Leu Leu Pro Arg Val
 2850 2855 2860

<210> 109

<211> 271

<212> PRT

<213> Homo sapiens

<400> 109

Met Val Leu Ile Lys Glu Phe Arg Val Val Leu Pro Cys Ser Val Gln
 1 5 10 15
 Glu Tyr Gln Val Gly Gln Leu Tyr Ser Val Ala Glu Ala Ser Lys Asn
 20 25 30
 Glu Thr Gly Gly Gly Glu Gly Ile Glu Val Leu Lys Asn Glu Pro Tyr
 35 40 45
 Glu Lys Asp Gly Glu Lys Gly Gln Tyr Thr His Lys Ile Tyr His Leu
 50 55 60

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Lys Ser Lys Val Pro Ala Phe Val Arg Met Ile Ala Pro Glu Gly Ser
 65 70 75 80
 Leu Val Phe His Glu Lys Ala Trp Asn Ala Tyr Pro Tyr Cys Arg Thr
 85 90 95
 Ile Val Thr Asn Glu Tyr Met Lys Asp Asp Phe Phe Ile Lys Ile Glu
 100 105 110
 Thr Trp His Lys Pro Asp Leu Gly Thr Leu Glu Asn Val His Gly Leu
 115 120 125
 Asp Pro Asn Thr Trp Lys Thr Val Glu Ile Val His Ile Asp Ile Ala
 130 135 140
 Asp Arg Ser Gln Val Glu Pro Ala Asp Tyr Lys Ala Asp Glu Asp Pro
 145 150 155 160
 Ala Leu Phe Gln Ser Val Lys Thr Lys Arg Gly Pro Leu Gly Pro Asn
 165 170 175
 Trp Lys Lys Glu Leu Ala Asn Ser Pro Asp Cys Pro Gln Met Cys Ala
 180 185 190
 Tyr Lys Leu Val Thr Ile Lys Phe Lys Trp Trp Gly Leu Gln Ser Lys
 195 200 205
 Val Glu Asn Phe Ile Gln Lys Gln Glu Lys Arg Ile Phe Thr Asn Phe
 210 215 220
 His Arg Gln Leu Phe Cys Trp Ile Asp Lys Trp Ile Asp Leu Thr Met
 225 230 235 240
 Glu Asp Ile Arg Arg Met Glu Asp Glu Thr Gln Lys Glu Leu Glu Thr
 245 250 255
 Met Arg Lys Arg Gly Ser Val Arg Gly Thr Ser Ala Ala Asp Val
 260 265 270

<210> 110

<211> 233

<212> PRT

<213> Homo sapiens

<400> 110

Asn Ser Val Leu Asn Ser Asn Ala Ile Lys Asn Leu Pro Pro Pro Leu
 1 5 10 15
 Gly Gly Ala Ala Gly His Pro Gly Ser Ala Val Ser Ala Ala Pro Gly
 20 25 30
 Ile Leu Tyr Pro Gly Gly Asn Lys Tyr Gln Thr Ile Asp Asn Tyr Gln
 35 40 45
 Pro Tyr Pro Cys Ala Glu Asp Glu Glu Cys Gly Thr Asp Glu Tyr Cys
 50 55 60
 Ala Ser Pro Thr Arg Gly Gly Asp Ala Gly Val Gln Ile Cys Leu Ala
 65 70 75 80
 Cys Arg Lys Arg Arg Lys Arg Cys Met Arg His Ala Met Cys Cys Pro
 85 90 95
 Gly Asn Tyr Cys Lys Asn Gly Ile Cys Val Ser Ser Asp Gln Asn His
 100 105 110
 Phe Arg Gly Glu Ile Glu Glu Thr Ile Thr Glu Ser Phe Gly Asn Asp
 115 120 125
 His Ser Thr Leu Asp Gly Tyr Ser Arg Arg Thr Thr Leu Ser Ser Lys
 130 135 140
 Met Tyr His Thr Lys Gly Gln Glu Gly Ser Val Cys Leu Arg Ser Ser
 145 150 155 160
 Asp Cys Ala Ser Gly Leu Cys Cys Ala Arg His Phe Trp Ser Lys Ile
 165 170 175
 Cys Lys Pro Val Leu Lys Glu Gly Gln Val Cys Thr Lys His Arg Arg
 180 185 190

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Lys Gly Ser His Gly Leu Glu Ile Phe Gln Arg Cys Tyr Cys Gly Glu
 195 200 205
 Gly Leu Ser Cys Arg Ile Gln Lys Asp His His Gln Ala Ser Asn Ser
 210 215 220
 Ser Arg Leu His Thr Cys Gln Arg His
 225 230

<210> 111
 <211> 212
 <212> PRT
 <213> Homo sapiens

<400> 111
 Asn Ser Val Leu Asn Ser Asn Ala Ile Lys Asn Leu Pro Pro Pro Leu
 1 5 10 15
 Gly Gly Ala Ala Gly His Pro Gly Ser Ala Val Ser Ala Ala Pro Gly
 20 25 30
 Ile Leu Tyr Pro Gly Gly Asn Lys Tyr Gln Thr Ile Asp Asn Tyr Gln
 35 40 45
 Pro Tyr Pro Cys Ala Glu Asp Glu Glu Cys Gly Thr Asp Glu Tyr Cys
 50 55 60
 Ala Ser Pro Thr Arg Gly Gly Asp Ala Gly Val Gln Ile Cys Leu Ala
 65 70 75 80
 Cys Arg Lys Arg Arg Lys Arg Cys Met Arg His Ala Met Cys Cys Pro
 85 90 95
 Gly Asn Tyr Cys Lys Asn Gly Ile Cys Val Ser Ser Asp Gln Asn His
 100 105 110
 Phe Arg Gly Glu Ile Glu Glu Thr Ile Thr Glu Ser Phe Gly Asn Asp
 115 120 125
 His Ser Thr Leu Asp Gly Tyr Ser Arg Arg Thr Thr Leu Ser Ser Lys
 130 135 140
 Met Tyr His Thr Lys Gly Gln Glu Gly Ser Val Cys Leu Arg Ser Ser
 145 150 155 160
 Asp Cys Ala Ser Gly Leu Cys Cys Ala Arg His Phe Trp Ser Lys Ile
 165 170 175
 Cys Lys Pro Val Leu Lys Glu Gly Gln Val Cys Thr Lys His Arg Arg
 180 185 190
 Lys Gly Ser His Gly Leu Glu Ile Phe Gln Arg Cys Tyr Cys Gly Glu
 195 200 205
 Gly Leu Ser Cys
 210

<210> 112
 <211> 149
 <212> PRT
 <213> Homo sapiens

<400> 112
 Asn Ser Val Leu Asn Ser Asn Ala Ile Lys Asn Leu Pro Pro Pro Leu
 1 5 10 15
 Gly Gly Ala Ala Gly His Pro Gly Ser Ala Val Ser Ala Ala Pro Gly
 20 25 30
 Ile Leu Tyr Pro Gly Gly Asn Lys Tyr Gln Thr Ile Asp Asn Tyr Gln
 35 40 45
 Pro Tyr Pro Cys Ala Glu Asp Glu Glu Cys Gly Thr Asp Glu Tyr Cys

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50		55		60
Ala Ser Pro Thr Arg Gly Gly Asp Ala Gly Val Gln Ile Cys Leu Ala				
65		70		75
Cys Arg Lys Arg Arg Lys Arg Cys Met Arg His Ala Met Cys Cys Pro				
	85		90	
Gly Asn Tyr Cys Lys Asn Gly Ile Cys Val Ser Ser Asp Gln Asn His				
	100		105	
Phe Arg Gly Glu Ile Glu Glu Thr Ile Thr Glu Ser Phe Gly Asn Asp				
	115		120	
His Ser Thr Leu Asp Gly Tyr Ser Arg Arg Thr Thr Leu Ser Ser Lys				
	130		135	
Met Tyr His Thr Lys			140	
145				

<210> 113
 <211> 170
 <212> PRT
 <213> Homo sapiens

<400> 113

Cys Ala Ser Pro Thr Arg Gly Gly Asp Ala Gly Val Gln Ile Cys Leu				
1	5		10	15
Ala Cys Arg Lys Arg Arg Lys Arg Cys Met Arg His Ala Met Cys Cys				
	20		25	30
Pro Gly Asn Tyr Cys Lys Asn Gly Ile Cys Val Ser Ser Asp Gln Asn				
	35		40	45
His Phe Arg Gly Glu Ile Glu Glu Thr Ile Thr Glu Ser Phe Gly Asn				
	50		55	60
Asp His Ser Thr Leu Asp Gly Tyr Ser Arg Arg Thr Thr Leu Ser Ser				
65	70		75	80
Lys Met Tyr His Thr Lys Gly Gln Glu Gly Ser Val Cys Leu Arg Ser				
	85		90	95
Ser Asp Cys Ala Ser Gly Leu Cys Cys Ala Arg His Phe Trp Ser Lys				
	100		105	110
Ile Cys Lys Pro Val Leu Lys Glu Gly Gln Val Cys Thr Lys His Arg				
	115		120	125
Arg Lys Gly Ser His Gly Leu Glu Ile Phe Gln Arg Cys Tyr Cys Gly				
	130		135	140
Glu Gly Leu Ser Cys Arg Ile Gln Lys Asp His His Gln Ala Ser Asn				
145	150		155	160
Ser Ser Arg Leu His Thr Cys Gln Arg His				
	165		170	

<210> 114
 <211> 128
 <212> PRT
 <213> Homo sapiens

<400> 114

Val Ser Ser Asp Gln Asn His Phe Arg Gly Glu Ile Glu Glu Thr Ile				
1	5		10	15
Thr Glu Ser Phe Gly Asn Asp His Ser Thr Leu Asp Gly Tyr Ser Arg				
	20		25	30
Arg Thr Thr Leu Ser Ser Lys Met Tyr His Thr Lys Gly Gln Glu Gly				
	35		40	45

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Ser Val Cys Leu Arg Ser Ser Asp Cys Ala Ser Gly Leu Cys Cys Ala
  50          55          60
Arg His Phe Trp Ser Lys Ile Cys Lys Pro Val Leu Lys Glu Gly Gln
  65          70          75          80
Val Cys Thr Lys His Arg Arg Lys Gly Ser His Gly Leu Glu Ile Phe
          85          90          95
Gln Arg Cys Tyr Cys Gly Glu Gly Leu Ser Cys Arg Ile Gln Lys Asp
          100          105          110
His His Gln Ala Ser Asn Ser Ser Arg Leu His Thr Cys Gln Arg His
          115          120          125

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<210> 115
 <211> 84
 <212> PRT
 <213> Homo sapiens

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<400> 115
Gly Gln Glu Gly Ser Val Cys Leu Arg Ser Ser Asp Cys Ala Ser Gly
  1          5          10          15
Leu Cys Cys Ala Arg His Phe Trp Ser Lys Ile Cys Lys Pro Val Leu
          20          25          30
Lys Glu Gly Gln Val Cys Thr Lys His Arg Arg Lys Gly Ser His Gly
          35          40          45
Leu Glu Ile Phe Gln Arg Cys Tyr Cys Gly Glu Gly Leu Ser Cys Arg
          50          55          60
Ile Gln Lys Asp His His Gln Ala Ser Asn Ser Ser Arg Leu His Thr
  65          70          75          80
Cys Gln Arg His

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<210> 116
 <211> 149
 <212> PRT
 <213> Homo sapiens
 <400> 116

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Cys Ala Ser Pro Thr Arg Gly Gly Asp Ala Gly Val Gln Ile Cys Leu
  1          5          10          15
Ala Cys Arg Lys Arg Arg Lys Arg Cys Met Arg His Ala Met Cys Cys
          20          25          30
Pro Gly Asn Tyr Cys Lys Asn Gly Ile Cys Val Ser Ser Asp Gln Asn
          35          40          45
His Phe Arg Gly Glu Ile Glu Glu Thr Ile Thr Glu Ser Phe Gly Asn
          50          55          60
Asp His Ser Thr Leu Asp Gly Tyr Ser Arg Arg Thr Thr Leu Ser Ser
  65          70          75          80
Lys Met Tyr His Thr Lys Gly Gln Glu Gly Ser Val Cys Leu Arg Ser
          85          90          95
Ser Asp Cys Ala Ser Gly Leu Cys Cys Ala Arg His Phe Trp Ser Lys
          100          105          110
Ile Cys Lys Pro Val Leu Lys Glu Gly Gln Val Cys Thr Lys His Arg
          115          120          125
Arg Lys Gly Ser His Gly Leu Glu Ile Phe Gln Arg Cys Tyr Cys Gly
          130          135          140
Glu Gly Leu Ser Cys
  145

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<210> 117
 <211> 107
 <212> PRT
 <213> Homo sapiens

<400> 117
 Val Ser Ser Asp Gln Asn His Phe Arg Gly Glu Ile Glu Glu Thr Ile
 1 5 10 15
 Thr Glu Ser Phe Gly Asn Asp His Ser Thr Leu Asp Gly Tyr Ser Arg
 20 25 30
 Arg Thr Thr Leu Ser Ser Lys Met Tyr His Thr Lys Gly Gln Glu Gly
 35 40 45
 Ser Val Cys Leu Arg Ser Ser Asp Cys Ala Ser Gly Leu Cys Cys Ala
 50 55 60
 Arg His Phe Trp Ser Lys Ile Cys Lys Pro Val Leu Lys Glu Gly Gln
 65 70 75 80
 Val Cys Thr Lys His Arg Arg Lys Gly Ser His Gly Leu Glu Ile Phe
 85 90 95
 Gln Arg Cys Tyr Cys Gly Glu Gly Leu Ser Cys
 100 105

<210> 118
 <211> 15
 <212> PRT
 <213> Homo sapiens

<400> 118
 Gly Asn Lys Tyr Gln Thr Ile Asp Asn Tyr Gln Pro Tyr Pro Cys
 1 5 10 15

<210> 119
 <211> 22
 <212> PRT
 <213> Homo sapiens

<400> 119
 Leu Asp Gly Tyr Ser Arg Arg Thr Thr Leu Ser Ser Lys Met Tyr His
 1 5 10 15
 Thr Lys Gly Gln Glu Gly
 20

<210> 120
 <211> 21
 <212> PRT
 <213> Homo sapiens

<400> 120
 Arg Ile Gln Lys Asp His His Gln Ala Ser Asn Ser Ser Arg Leu His
 1 5 10 15
 Thr Cys Gln Arg His
 20

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<210> 121
<211> 15
<212> PRT
<213> Homo sapiens

<400> 121
Arg Gly Glu Ile Glu Glu Thr Ile Thr Glu Ser Phe Gly Asn Asp
1 5 10 15

<210> 122
<211> 19
<212> PRT
<213> Homo sapiens

<400> 122
Glu Ile Phe Gln Arg Cys Tyr Cys Gly Glu Gly Leu Ser Cys Arg Ile
1 5 10 15
Gln Lys Asp

<210> 123
<211> 13
<212> PRT
<213> Artificial Sequence

<220>
<223> LRP5/HBM amino acid sequence

<400> 123
Met Tyr Trp Thr Asp Trp Val Glu Thr Pro Arg Ile Glu
1 5 10

<210> 124
<211> 13
<212> PRT
<213> Artificial Sequence

<220>
<223> LRP5/HBM amino acid sequence

<400> 124
Met Tyr Trp Thr Asp Trp Gly Glu Thr Pro Arg Ile Glu
1 5 10

<210> 125
<211> 13
<212> PRT
<213> Artificial Sequence

<220>
<223> LRP5/HBM amino acid sequence

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<400> 125

Lys Arg Thr Gly Gly Lys Arg Lys Glu Ile Leu Ser Ala
 1 5 10

<210> 126

<211> 17

<212> PRT

<213> Artificial Sequence

<220>

<223> LRP5/HBM amino acid sequence

<400> 126

Glu Arg Val Glu Lys Thr Thr Gly Asp Lys Arg Thr Arg Ile Gln Gly
 1 5 10 15
 Arg

<210> 127

<211> 16

<212> PRT

<213> Artificial Sequence

<220>

<223> LRP5/HBM amino acid sequence

<400> 127

Lys Gln Gln Cys Asp Ser Phe Pro Asp Cys Ile Asp Gly Ser Asp Glu
 1 5 10 15

<210> 128

<211> 266

<212> PRT

<213> Homo sapiens

<400> 128

Met Met Ala Leu Gly Ala Ala Gly Ala Thr Arg Val Phe Val Ala Met
 1 5 10 15
 Val Ala Ala Ala Leu Gly Gly His Pro Leu Leu Gly Val Ser Ala Thr
 20 25 30
 Leu Asn Ser Val Leu Asn Ser Asn Ala Ile Lys Asn Leu Pro Pro Pro
 35 40 45
 Leu Gly Gly Ala Ala Gly His Pro Gly Ser Ala Val Ser Ala Ala Pro
 50 55 60
 Gly Ile Leu Tyr Pro Gly Gly Asn Lys Tyr Gln Thr Ile Asp Asn Tyr
 65 70 75 80
 Gln Pro Tyr Pro Cys Ala Glu Asp Glu Glu Cys Gly Thr Asp Glu Tyr
 85 90 95
 Cys Ala Ser Pro Thr Arg Gly Gly Asp Ala Gly Val Gln Ile Cys Leu
 100 105 110
 Ala Cys Arg Lys Arg Arg Lys Arg Cys Met Arg His Ala Met Cys Cys
 115 120 125
 Pro Gly Asn Tyr Cys Lys Asn Gly Ile Cys Val Ser Ser Asp Gln Asn
 130 135 140

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His	Phe	Arg	Gly	Glu	Ile	Glu	Glu	Thr	Ile	Thr	Glu	Ser	Phe	Gly	Asn
145					150					155					160
Asp	His	Ser	Thr	Leu	Asp	Gly	Tyr	Ser	Arg	Arg	Thr	Thr	Leu	Ser	Ser
				165					170					175	
Lys	Met	Tyr	His	Thr	Lys	Gly	Gln	Glu	Gly	Ser	Val	Cys	Leu	Arg	Ser
			180					185					190		
Ser	Asp	Cys	Ala	Ser	Gly	Leu	Cys	Cys	Ala	Arg	His	Phe	Trp	Ser	Lys
	195					200						205			
Ile	Cys	Lys	Pro	Val	Leu	Lys	Glu	Gly	Gln	Val	Cys	Thr	Lys	His	Arg
	210					215					220				
Arg	Lys	Gly	Ser	His	Gly	Leu	Glu	Ile	Phe	Gln	Arg	Cys	Tyr	Cys	Gly
225					230					235					240
Glu	Gly	Leu	Ser	Cys	Arg	Ile	Gln	Lys	Asp	His	His	Gln	Ala	Ser	Asn
				245					250					255	
Ser	Ser	Arg	Leu	His	Thr	Cys	Gln	Arg	His						
			260					265							

<210> 129

<211> 39

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 129

tggtcagcgg cctggaggat gtggccgcag tggacttcc

39

<210> 130

<211> 39

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 130

ggaagtccac tgcggccaca tcctccaggc cgctgacca

39

<210> 131

<211> 40

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 131

aagctgtact ggacggactc agtgaccaac cgcacgagg

40

<210> 132

<211> 40

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

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<400> 132
cctcgatgcg gttggtcact gagtccgtcc agtacagctt 40

<210> 133
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> complementary synthetic oligonucleotide

<400> 133
atgtactgga cagactggaa ggagacgccc cggattgagc g 41

<210> 134
<211> 41
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<220>
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<400> 134
cgctcaatcc ggggcgtctc ctccagtct gtccagtaca t 41

<210> 135
<211> 41
<212> DNA
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<220>
<223> complementary synthetic oligonucleotide

<400> 135
atgtactgga cagactgggt tgagacgccc cggattgagc g 41

<210> 136
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
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<400> 136
cgctcaatcc ggggcgtctc aaaccagtct gtccagtaca t 41

<210> 137
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> complementary synthetic oligonucleotide

<400> 137

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atgtactgga cagactggat tgagacgccc cggattgagc g 41

<210> 138

<211> 41

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 138

cgctcaatcc ggggcgtctc aatccagtct gtccagtaca t 41

<210> 139

<211> 41

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 139

atgtactgga cagactggca ggagacgccc cggattgagc g 41

<210> 140

<211> 41

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 140

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<210> 141

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 141

cggacattta ctggcccaat gtactgacca tcgacctgga gg 42

<210> 142

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 142

cctccaggtc gatggtcagt acattgggcc agtaaattgtc cg 42

<210> 143

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<211> 40
<212> DNA
<213> Artificial Sequence

<220>
<223> complementary synthetic oligonucleotide

<400> 143
agctctactg ggctgacgtc aagctcagct tcatccaccg 40

<210> 144
<211> 40
<212> DNA
<213> Artificial Sequence

<220>
<223> complementary synthetic oligonucleotide

<400> 144
cgggtggatga agctgagctt gacgtcagcc cagtagagct 40

<210> 145
<211> 41
<212> DNA
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<220>
<223> complementary synthetic oligonucleotide

<400> 145
gagtgccctc tactcaccgc tggacatcca ggtgctgagc c 41

<210> 146
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> complementary synthetic oligonucleotide

<400> 146
ggctcagcac ctggatgtcc acgggtgagt agagggcact c 41

<210> 147
<211> 45
<212> DNA
<213> Artificial Sequence

<220>
<223> complementary synthetic oligonucleotide

<400> 147
catgtactgg acagactggg tagagaaccc taaaatcgag tgtgc 45

<210> 148
<211> 45
<212> DNA
<213> Artificial Sequence

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<220>

<223> complementary synthetic oligonucleotide

<400> 148

gcacactcga ttttaggggtt ctctacccag tctgtccagt acatg

45

<210> 149

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 149

catctactgg accgagtggg tcggcaagcc gaggatcgtg cg

42

<210> 150

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 150

cgcacgatcc tcggcttgcc gaccactcg gtccagtaga tg

42

<210> 151

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 151

gtacttcacc aacatgggtgg accgggcagc caagatcgaa cg

42

<210> 152

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 152

cgttcgatct tggctgcccg gtccaccatg ttggtgaagt ac

42

<210> 153

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

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<223> complementary synthetic oligonucleotide

<400> 153

gtactggaca gactgggtag aagtgccaaa gatagaacgt gc

42

<210> 154

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

<223> complementary synthetic oligonucleotide

<400> 154

gcacgttcta tctttggcac ttctacccag tctgtccagt ac

42

<210> 155

<211> 31

<212> DNA

<213> Artificial Sequence

<220>

<223> primer

<400> 155

ttttttgtcg accaattcca acgctatcaa g

31

<210> 156

<211> 31

<212> DNA

<213> Artificial Sequence

<220>

<223> primer

<400> 156

ttttttgtcg acctgcgcta gtcccacccg c

31

<210> 157

<211> 32

<212> DNA

<213> Artificial Sequence

<220>

<223> primer

<400> 157

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32

<210> 158

<211> 33

<212> DNA

<213> Artificial Sequence

<220>

<223> primer

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<400> 158
ttttttgtcg accggacaag aaggttctgt ttg 33

<210> 159
<211> 37
<212> DNA
<213> Artificial Sequence

<220>
<223> primer

<400> 159
ttttttgcfg ccgcttattt ggtgtgatac atttttg 37

<210> 160
<211> 35
<212> DNA
<213> Artificial Sequence

<220>
<223> primer

<400> 160
ttttttgcfg ccgcttagca agacagacct tctcc 35

<210> 161
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<212> DNA
<213> Artificial Sequence

<220>
<223> primer

<400> 161
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<210> 162
<211> 34
<212> DNA
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<220>
<223> primer

<400> 162
cagtgaattc accatgcaaa acaccacttt gttc 34

<210> 163
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> primer

<400> 163
cagttgcggc cgctcatctc cggtggcctc tg 32

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<210> 164
 <211> 39
 <212> DNA
 <213> Homo sapiens

<220>
 <223> primer

<400> 164
 caatagtcga cgaattcacc atggctctgg gcgcagcgg 39

<210> 165
 <211> 42
 <212> DNA
 <213> Homo sapiens

<220>
 <223> primer

<400> 165
 gtattgcggc cgctctagat tagtgtctct gacaagtgtg aa 42

<210> 166
 <211> 32
 <212> DNA
 <213> Homo sapiens

<220>
 <223> primer

<400> 166
 ctaacggatc caccatggcc gcgttgatgc gg 32

<210> 167
 <211> 33
 <212> DNA
 <213> Homo sapiens

<220>
 <223> primer

<400> 167
 gattcgaatt ctcaaatttt ctgacacaca tgg 33

<210> 168
 <211> 690
 <212> DNA
 <213> Homo sapiens

<400> 168
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 tctgcagtca gcgccgcgcc gggaatcctg taccggggcg ggaataagta ccagaccatt 120
 gacaactacc agccgtaccc gtgcgcagag gacgaggagt gcggcactga tgagtactgc 180
 gctagtccca cccgcggagg ggacgcgggc gtgcaaattc gtctcgccctg caggaagcgc 240
 cgaaaacgct gcatgcgtca cgctatgtgc tgccccggga attactgcaa aaatggaata 300
 tgtgtgtctt ctgatcaaaa tcatttccga ggagaaattg aggaaacat cactgaaagc 360
 tttggtaatg atcatagcac cttggatggg tattccagaa gaaccacctt gtcttcaaaa 420

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atgtatcaca	ccaaaggaca	agaaggttct	gtttgtctcc	ggatcatcaga	ctgtgcctca	480
ggattgtgtt	gtgctagaca	cttctgggtcc	aagatctgta	aacctgtcct	gaaagaaggt	540
caagtgtgta	ccaagcatag	gagaaaaggc	tctcatggac	tagaaatatt	ccagcgttgt	600
tactgtggag	aaggtctgtc	ttgccggata	cagaaagatc	accatcaagc	cagtaattct	660
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<210> 169

<211> 1226

<212> DNA

<213> Homo sapiens

<400> 169

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cagccctttg	ttttgacctc	tctgagccaa	ggccaaaacc	cagacaggca	gccccacgac	180
ctcagcatcg	acatctacag	ccggacactg	ttctggacgt	gcgaggccac	caataccatc	240
aacgtccaca	ggctgagcgg	ggaagccatg	gggggtggtg	tgcgtgggga	ccgcgacaag	300
cccagggcca	tcgtcgtcaa	cgcgagcgca	gggtacctgt	acttcaccaa	catgcaggac	360
cgggcagcca	agatcgaacg	cgcagccctg	gacggcaccg	agcgcgaggt	cctcttcacc	420
accggcctca	tccgccctgt	ggccctgggtg	gtagacaaca	cactgggcaa	gctgttcttg	480
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ctggaggagt	tctcagccca	cccatgtgcc	cgtgacaatg	gtggctgctc	ccacatctgt	780
attgccaagg	gtgatgggac	accacgggtg	tcatgcccag	tccacctcgt	gctcctgcag	840
aacctgctga	cctgtggaga	gccgcccacc	tgtctcccgg	accagtttgc	atgtgccaca	900
ggggagatcg	actgtatccc	cggggcctgg	cgctgtgacg	gctttcccga	gtgcgatgac	960
cagagcgacg	aggagggctg	ccccgtgtgc	tccgccgcc	agttcccctg	cgcgcggggt	1020
cagtgtggtg	gacctgcgcc	tgcgctgcga	cggcgaggca	gactgtcagg	accgctcaga	1080
cgaggcggac	tgtgacgcca	tctgcctgcc	caaccagttc	cggtgtgcga	gcggccagtg	1140
tgtcctcatc	aaacagccag	tgcgactcct	tccccgactg	tatcgacggc	tccgacgagc	1200
tcatgtgtga	aatcaccaag	ccgccc				1226

<210> 170

<211> 934

<212> DNA

<213> Homo sapiens

<400> 170

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gcagccaaga	tcgaacgcgc	agccctggac	ggcaccgagc	gcgaggctct	cttcaccacc	120
ggcctcatcc	gccctgtggc	cctggtggta	gacaacacac	tgggcaagct	gttctgggtg	180
gacgcggacc	tgaagcgcac	tgagagctgt	gacctgtcag	gggccaaccg	cctgaccctg	240
gaggacgcca	acatcgtgca	gcctctgggc	ctgaccatcc	ttggcaagca	tctctactgg	300
atcgaccgcc	agcagcagat	gatcgagcgt	gtggagaaga	ccaccgggga	caagcggact	360
cgcattccagg	gccgtgtcgc	ccacctcact	ggcatccatg	cagtggagga	agtcagcctg	420
gaggagtctt	cagcccaccc	atgtgcccgt	gacaatggtg	gctgctccca	catctgtatt	480
gccaaggggtg	atgggacacc	acggtgctca	tgcccagttc	acctcgtgct	cctgcagaac	540
ctgctgacct	gtggagagcc	gcccacctgc	tccccggacc	agtttgcatg	tgccacaggg	600
gagatcgact	gtatccccgg	ggcctggcgc	tgtgacggct	ttcccagagt	cgatgaccag	660
agcgacgagg	agggctgccc	cgtgtgcctc	cgccgcccag	ttcccctgcg	cgcggggtca	720
gtgtgtggac	ctgcgcctgc	gctgcgacgg	cgaggcagac	tgtcaggacc	gctcagacga	780
ggcggactgt	gacgccatct	ggcctgcccc	accagttccg	gtgtgcgagc	ggccagtgtg	840
tcctcatcaa	acagcagtg	gactccttcc	ccgactgtat	cgacggctcc	gacgagctca	900
tgtgtgaaat	caccaagccg	ccctaagcgg	ccgc			934

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<210> 171
<211> 16
<212> PRT
<213> Homo sapiens

<400> 171
Ser Val Gly Cys Leu Leu Cys Ala Gly Leu Gly Val Trp Ser Leu Ser
1 5 10 15

<210> 172
<211> 19
<212> PRT
<213> Homo sapiens

<400> 172
Trp Cys Cys Cys Gly Leu Phe Arg Gly Val Cys Val Trp Ser Cys Gly
1 5 10 15
Ala Asp Asp

<210> 173
<211> 16
<212> PRT
<213> Homo sapiens

<400> 173
Gly Trp Arg Arg Cys Asp Trp Cys Gly Cys Val Ser Trp Cys Trp Val
1 5 10 15

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<213> Homo sapiens

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<400> 177

Val	Leu	Ser	Val	Cys	Glu	Val	Cys	Gly	Gly	Val	Phe	Val	Arg	Arg	Cys
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<213> Homo sapiens

<400> 178

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<400> 179

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<400> 181

Cys	Val	Cys	Ala	Ser	Phe	Cys	Cys	Cys	Val	Cys	Gly	Leu	Arg	Leu	Leu
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<210> 184
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<400> 184
Cys Cys Arg Cys Cys His Cys Trp Asp Cys Glu Trp His Met Cys Val
1 5 10 15

<210> 185
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<400> 185
Phe Cys Ala Ser Cys Cys Trp Cys Gly Cys Asp Cys Phe Gly Trp Val
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Cys Asp Tyr Cys Trp Ser Cys Gly Val Trp Cys Pro Ser Ser Trp Leu
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1

5

10

15

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<212> PRT

<213> Homo sapiens

<400> 189

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<213> Homo sapiens

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<212> DNA

<213> Homo sapiens

<400> 196

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gaaatattcc agcgttggtt ctgtggagaa ggtctgtctt gccggataca gaaagatcac 420
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          20          25          30
Asp Asp Ser Phe Asp Thr Asp Val Leu Lys Ala Asp Gly Ala Ile Leu
          35          40          45
Val Asp Phe Trp Ala Glu Trp Cys Gly Pro Asn Ser Val Val Leu Cys
          50          55          60
Ser Arg Cys Gly Arg Leu Trp Arg Trp Ser Cys Gly Thr Ser Gly Pro
65          70          75          80
Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp Glu Tyr Gln
          85          90          95
Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn Pro Gly Thr
          100          105          110
Ala Pro Lys Tyr Gly Ile Arg Gly Ile Pro Thr Leu Leu Leu Phe Lys
          115          120          125
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          130          135          140
Leu Lys Glu Phe Leu Asp Ala Asn Leu Ala
145          150

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Gly Ser Thr Gly Asp Gly Ser Met Ser Asp Lys Ile Ile His Leu Thr
          20          25          30
Asp Asp Ser Phe Asp Thr Asp Val Leu Lys Ala Asp Gly Ala Ile Leu
          35          40          45
Val Asp Phe Trp Ala Glu Trp Cys Gly Pro Asn Ser Gly Trp Arg Trp
          50          55          60
Cys Gly Arg Cys Gly Ala Leu Trp Trp Arg Arg Val Thr Ser Gly Pro
65          70          75          80
Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp Glu Tyr Gln
          85          90          95
Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn Pro Gly Thr
          100          105          110
Ala Pro Lys Tyr Gly Ile Arg Gly Ile Pro Thr Leu Leu Leu Phe Lys
          115          120          125
Asn Gly Glu Val Ala Ala Thr Lys Val Gly Ala Leu Ser Lys Gly Gln
          130          135          140
Leu Lys Glu Phe Leu Asp Ala Asn Leu Ala
145          150

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<210> 206

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<212> PRT

<213> Homo sapiens

<400> 206

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Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
 1          5          10          15

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Gly Ser Thr Gly Asp Gly Ser Met Ser Asp Lys Ile Ile His Leu Thr
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 Asp Asp Ser Phe Asp Thr Asp Val Leu Lys Ala Asp Gly Ala Ile Leu
 35 40 45
 Val Asp Phe Trp Ala Glu Trp Cys Gly Pro Asn Ser Glu Val Arg Gln
 50 55 60
 Val Thr Cys Ile Arg Cys Arg Arg Gly Phe Leu Leu Thr Ser Gly Pro
 65 70 75 80
 Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp Glu Tyr Gln
 85 90 95
 Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn Pro Gly Thr
 100 105 110
 Ala Pro Lys Tyr Gly Ile Arg Gly Ile Pro Thr Leu Leu Leu Phe Lys
 115 120 125
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 130 135 140
 Leu Lys Glu Phe Leu Asp Ala Asn Leu Ala
 145 150

<210> 207
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 <212> PRT
 <213> Homo sapiens

<400> 207

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 1 5 10 15
 Gly Ser Thr Gly Asp Gly Ser Met Ser Asp Lys Ile Ile His Leu Thr
 20 25 30
 Asp Asp Ser Phe Asp Thr Asp Val Leu Lys Ala Asp Gly Ala Ile Leu
 35 40 45
 Val Asp Phe Trp Ala Glu Trp Cys Gly Pro Asn Ser Gly Gly Gly Gly
 50 55 60
 Met Ile Trp Glu Ala Trp Ser Cys Tyr Ala Cys Gly Thr Ser Gly Pro
 65 70 75 80
 Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp Glu Tyr Gln
 85 90 95
 Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn Pro Gly Thr
 100 105 110
 Ala Pro Lys Tyr Gly Ile Arg Gly Ile Pro Thr Leu Leu Leu Phe Lys
 115 120 125
 Asn Gly Glu Val Ala Ala Thr Lys Val Gly Ala Leu Ser Lys Gly Gln
 130 135 140
 Leu Lys Glu Phe Leu Asp Ala Asn Leu Ala
 145 150

<210> 208
 <211> 154
 <212> PRT
 <213> Homo sapiens

<400> 208

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
 1 5 10 15
 Gly Ser Thr Gly Asp Gly Ser Met Ser Asp Lys Ile Ile His Leu Thr

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20	25	30
Asp Asp Ser Phe Asp Thr Asp Val Leu Lys Ala Asp Gly Ala Ile Leu		
35	40	45
Val Asp Phe Trp Ala Glu Trp Cys Gly Pro Asn Ser Leu Trp Ile Gly		
50	55	60
Pro Gly Asp Gln Gly Leu Phe Arg Arg Phe Val Phe Thr Ser Gly Pro		
65	70	75
Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp Glu Tyr Gln		
85	90	95
Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn Pro Gly Thr		
100	105	110
Ala Pro Lys Tyr Gly Ile Arg Gly Ile Pro Thr Leu Leu Leu Phe Lys		
115	120	125
Asn Gly Glu Val Ala Ala Thr Lys Val Gly Ala Leu Ser Lys Gly Gln		
130	135	140
Leu Lys Glu Phe Leu Asp Ala Asn Leu Ala		
145	150	

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<211> 151

<212> PRT

<213> Homo sapiens

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20	25
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35	40
Thr Leu Asp Gly Tyr Ser Arg Arg Thr Thr Leu Ser Ser Lys Met Tyr	
50	55
His Thr Lys Gly Gln Glu Gly Ser Val Cys Leu Arg Ser Ser Asp Cys	
65	70
Ala Ser Gly Leu Cys Cys Ala Arg His Phe Trp Ser Lys Ile Cys Lys	
85	90
Pro Val Leu Lys Glu Gly Gln Val Cys Thr Lys His Arg Arg Lys Gly	
100	105
Ser His Gly Leu Glu Ile Phe Gln Arg Cys Tyr Cys Gly Glu Gly Leu	
115	120
Ser Cys Arg Ile Gln Lys Asp His His Gln Ala Ser Asn Ser Ser Arg	
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Leu His Thr Cys Gln Arg His	
145	150

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<213> Homo sapiens

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20	25
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30	

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65					70					75					80
Ile	Thr	Glu	Ser	Phe	Gly	Asn	Asp	His	Ser	Thr	Leu	Asp	Gly	Tyr	Ser
				85					90					95	
Arg	Arg	Thr	Thr	Leu	Ser	Ser	Lys	Met	Tyr	His	Thr	Lys	Gly	Gln	Glu
			100					105					110		
Gly	Ser	Val	Cys	Leu	Arg	Ser	Ser	Asp	Cys	Ala	Ser	Gly	Leu	Cys	Cys
		115					120					125			
Ala	Arg	His	Phe	Trp	Ser	Lys	Ile	Cys	Lys	Pro	Val	Leu	Lys	Glu	Gly
	130					135					140				
Gln	Val	Cys	Thr	Lys	His	Arg	Arg	Lys	Gly	Ser	His	Gly	Leu	Glu	Ile
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Phe	Gln	Arg	Cys	Tyr	Cys	Gly	Glu	Gly	Leu	Ser	Cys				
				165					170						

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<400> 212

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 65 70 75 80
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 85 90 95
 Gly Ile Pro Thr Leu Leu Leu Phe Lys Asn Gly Glu Val Ala Ala Thr
 100 105 110
 Lys Val Gly Ala Leu Ser Lys Gly Gln Leu Lys Glu Phe Leu Asp Ala
 115 120 125
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<210> 213
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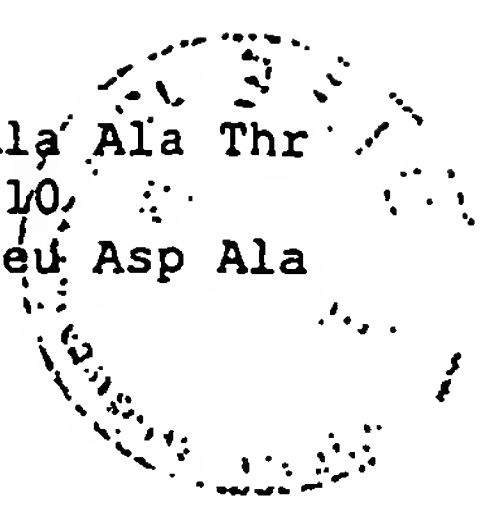
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 35 40 45
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 50 55 60
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 65 70 75 80
 Ala Lys Leu Asn Ile Asp Gln Asn Pro Gly Thr Ala Pro Lys Tyr Gly
 85 90 95
 Ile Arg Gly Ile Pro Thr Leu Leu Leu Phe Lys Asn Gly Glu Val Ala
 100 105 110
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 115 120 125
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 130

<210> 214
 <211> 131
 <212> PRT
 <213> Homo sapiens

<400> 214
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 35 40 45
 Arg Arg Phe Val Phe Thr Ser Gly Pro Cys Lys Met Ile Ala Pro Ile
 50 55 60
 Leu Asp Glu Ile Ala Asp Glu Tyr Gln Gly Lys Leu Thr Val Ala Lys
 65 70 75 80
 Leu Asn Ile Asp Gln Asn Pro Gly Thr Ala Pro Lys Tyr Gly Ile Arg
 85 90 95

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Gly Ile Pro Thr Leu Leu Leu Phe Lys Asn Gly Glu Val Ala Ala Thr
100 105 110
Lys Val Gly Ala Leu Ser Lys Gly Gln Leu Lys Glu Phe Leu Asp Ala
115 120 125
Asn Leu Ala
130



(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
21 November 2002 (21.11.2002)

PCT

(10) International Publication Number
WO 02/092015 A3

(51) International Patent Classification⁷: **A61K 39/395**,
39/00, 39/38

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(21) International Application Number: **PCT/US02/15982**

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(22) International Filing Date: **17 May 2002 (17.05.2002)**

(25) Filing Language: **English**

(26) Publication Language: **English**

(30) Priority Data:
60/291,311 17 May 2001 (17.05.2001) US
60/353,058 1 February 2002 (01.02.2002) US
60/361,293 4 March 2002 (04.03.2002) US

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU,
AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU,
CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH,
GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC,
LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW,
MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG,
SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ,
VN, YU, ZA, ZM, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM,
KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW),
Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM),
European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR,
GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent
(BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR,
NE, SN, TD, TG).

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Published:

— *with international search report*

(88) Date of publication of the international search report:
23 October 2003

*For two-letter codes and other abbreviations, refer to the "Guid-
ance Notes on Codes and Abbreviations" appearing at the begin-
ning of each regular issue of the PCT Gazette.*

(54) Title: **REAGENTS AND METHODS FOR MODULATING DKK-MEDIATED INTERACTIONS**

(57) Abstract: The present invention provides reagents, compounds, compositions, and methods relating to novel interactions of the extracellular domain of LRP5, HBM (a variant of LRP5), and/or LRP6 with Dkk, including Dkk-1. The various nucleic acids, polypeptides, antibodies, assay methods, diagnostic methods, and methods of treatment of the present invention are related to and impact on Dkk, LRP5, LRP6, HBM, and Wnt signaling. Dkk, LRP5, LRP6, HBM, and Wnt are implicated in bone and lipid cellular signaling. Thus, the present invention provides reagents and methods for modulating lipid levels and/or bone mass and is useful in the treatment and diagnosis of abnormal lipid levels and bone mass disorders, such as osteoporosis.

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INTERNATIONAL SEARCH REPORT

International application No.

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A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : A61K 39/395, 00, 38

US CL : 424/130.1, 184.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/130.1, 184.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
Please See Continuation Sheet

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	GONG, Y et al. LDL RECEPTOR-RELATED PROTEIN 5 (LRP-5) AFFECTS BONE ACCRUAL AND EYE DEVELOPMENT, CELL, 2001 VOL.107, PAGES 513-523	1-11 and 13-23
	ZORN A. WNT SIGNALLING: ANTAGONISTIC DICKKOPFS, CURRENT BIOLOGY, 2001, VOL.11, No.15, PAGES R592-R595	1-11 and 13-23
A	WO 9846743 A1 (THE WELL-COME TRUST LIMITED), 22 OCTOBER 1998 (22.10.98) see entire document.	1-11 and 13-23

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

18 March 2003 (18.03.2003)

Date of mailing of the international search report

08 AUG 2003

Name and mailing address of the ISA/US

Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231

Facsimile No. (703)305-3230

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Telephone No. 703/308-0196

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Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claim Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claim Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claim Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:
Please See Continuation Sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-11 and 13-23

Remark on Protest ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING

This application contains the following inventions or groups of inventions which are not so linked as to form a single general inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

- I. Claims 1-11 and 13-23, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which inhibits Dkk binding to LRP5.
- II. Claims 1-11 and 13-20, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which inhibits Dkk binding to LRP6.
- III. Claims 1-11 and 13-20, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which inhibits Dkk binding to HBM.
- IV. Claims 1-10, 12-14 and 16-23 drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which enhances Dkk binding to LRP5.
- V. Claims 1-10, 12-14 and 16-20, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which enhances Dkk binding to LRP6.
- VI. Claims 1-10, 12-14 and 16-20, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which enhances Dkk binding to HBM.
- VII. Claims 2 - 4, 7-11, 13-20, 21-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.
- VIII. Claims 2 - 4, 7-11, 13-20, 24-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.
- IX. Claims 2 - 4, 7-11, 13-20, 24-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.
- X. Claims 2 - 4, 7-10, 12-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP5, wherein said composition enhances Dkk binding to LRP5.
- XI. Claims 2 - 4, 7-10, 12-20, 24-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.
- XII. Claims 2 - 4, 7-10, 12-20, 24-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with HBM, wherein said composition enhances Dkk binding to HBM.
- XIII. Claims 2 - 4, 7-11, 13-23, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.
- XIV. Claims 2 - 4, 7-11, 13-23, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.
- XV. Claims 2 - 4, 7-11, 13-23, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.

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XVI. Claims 2 - 4, 7-10, 12-23, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition enhances Dkk binding to LRP5.

XIV. Claims 2 - 4, 7-10, 12-20, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.

XV. Claims 2 - 4, 7-10, 12-20, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition enhances Dkk binding to HBM.

XVI. Claims 2 - 4, 7-11, 13-23, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.

XVII. XVI. Claims 2 - 4, 7-11, 13-23, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.

XVIII. Claims 2 - 4, 7-11, 13-23, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.

XIX. Claims 2 - 4, 7-10, 12-23, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition enhances Dkk binding to LRP5.

XX. Claims 2 - 4, 7-10, 12-20, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.

XXI. Claims 2 - 4, 7-10, 12-20, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition enhances Dkk binding to HBM.

XXII. Claims 2, 35 and 36 drawn to a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of Dkk.

XXIII. Claims 2, 35 and 36 drawn to a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of LRP5.

XXIV. Claims 2, 35 and 36 drawn to a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of LRP6.

XXV. Claims 2, 35 and 36 drawn to a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of HBM.

XXVI. Claims 2, 37-43, 44-47, drawn to a method of screening for a compound which modulates the interaction of DKK with LRP5.

XXVII. Claims 2, 37-43, 47, drawn to a method of screening for a compound which modulates the interaction of DKK with LRP6.

XXVIII. Claims 2, 37-43, 47, drawn to a method of screening for a compound which modulates the interaction of DKK with HBM.

XXIX. Claims 2, 41-43, 48-49, drawn to a method of screening for a compound which modulates the interaction of Dkk with a Dkk interacting proteins.

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- XXX. Claims 50-63 drawn to a composition comprising a LRP5 and a pharmaceutical acceptable carrier thereof.
- XXXI. Claims 50- 59, 63 drawn to a composition comprising a LRP6 and a pharmaceutical acceptable carrier thereof.
- XXXII. Claims 50- 59, 63 drawn to a composition comprising a HBM and a pharmaceutical acceptable carrier thereof.
- XXXIII. Claims 2 and 64 drawn to a method for identifying compound which modulate Dkk and LRP5 interaction.
- XXXIV. Claims 2 and 64 drawn to a method for identifying compound which modulate Dkk and LRP6 interaction.
- XXXV. Claims 2 and 64 drawn to a method for identifying compound which modulate Dkk and HBM interaction.
- XXXVI. Claims 2 and 65 drawn to a method of identifying binding partners for a Dkk protein.
- XXXVII. Claims 66-68 drawn to a nucleic acid and a vector encoding a Dkk interacting protein.
- XXXVIII. Claims 69-90, drawn to a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a LRP5.
- XXXIX. Claims 69-87, drawn to a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a LRP6.
- XL. Claims 69-87, drawn to a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a HBM.
- XLI. Claims 91-92 drawn to a transgenic animal.
- XLII. Claims 2 and 93 drawn to a method for identifying potential compound which modulate Dkk activity.
- XLIII - LXII. Claim 94, drawn to one specific peptide aptamer of one specific SEQ ID NOs : 171-88; 189-192.
- LXIII- LXXIX. Claims 95-97, drawn to an antibody which specifically recognizes and binds to specific peptides of SEQ ID NOs : 110-127.
- LXXX . Claims 2, 98-100 , drawn to a method of identifying Dkk interacting protein which modulate the interaction of Dkk with Wnt signaling pathway.
- LXXXI. Claims 2, 25 and 101-104, drawn to a method for identifying Dkk interacting proteins.
- LXXXII. Claims 2, 105-106, drawn to a method for identifying compounds which modulate Dkk and LRP5 interaction.
- LXXXIII. Claims 2, 105-106, drawn to a method for identifying compounds which modulate Dkk and LRP6 interaction.
- LXXXIV. Claims 2, 105-106, drawn to a method for identifying compounds which modulate Dkk and HBM interaction.
- LXXXV. Claims 2, 25, 107-110, drawn to a method for identifying compound which modulate the interaction of Dkk with Wnt signaling pathway.
- LXXXVI. Claims 2, 111, drawn to a method of testing compounds that modulate Dkk-mediated activity in a mammal.
- LXXXVII. Claims 2, 112, 113, drawn to method of screening for compound or composition which modulate the interaction of Dkk and Dkk interacting protein.
- LXXXVIII-CIX. Claim 114 drawn to antibody which recognizes and binds to one specific SEQ ID NOs: 171-192.

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The inventions listed as Groups 1-109 do not relate to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons:

The invention listed as groups 1-109 do not related to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, hey lack the same or corresponding special technical features for the following reasons:

The special technical features of Group I is considered a method of regulating LRP5 activity in a subject, comprising administering a composition which inhibits Dkk binding to LRP5.

The special technical features of Group II is considered a method of regulating LRP5 activity in a subject, comprising administering a composition which inhibits Dkk binding to LRP6.

The special technical features of Group III is considered a method of regulating LRP5 activity in a subject, comprising administering a composition which inhibits Dkk binding to HBM.

The special technical features of Group IV is considered a method of regulating LRP5 activity in a subject, comprising administering a composition which enhances Dkk binding to LRP5.

The special technical features of Group V is considered a method of regulating LRP5 activity in a subject, comprising administering a composition which enhances Dkk binding to LRP6.

The special technical features of Group VI is considered a method of regulating LRP5 activity in a subject, comprising administering a composition which enhances Dkk binding to HBM.

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The special technical features of Group VII is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.

The special technical features of Group VIII is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.

The special technical features of Group IX is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.

The special technical features of Group X is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with RP5, wherein said composition enhances Dkk binding to LRP5.

The special technical features of Group XI is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.

The special technical features of Group XII is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with HBM, wherein said composition enhances Dkk binding to HBM.

The special technical features of Group XIII is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.

The special technical features of Group XIV is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.

The special technical features of Group XV is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.

The special technical features of Group XVI is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition enhances Dkk binding to LRP5.

The special technical features of Group XIV is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.

The special technical features of Group XV is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition enhances Dkk binding to HBM.

The special technical features of Group XVI is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.

The special technical features of Group XVII is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.

The special technical features of Group XVIII is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.

The special technical features of Group XIX is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition enhances Dkk binding to LRP5.

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The special technical features of Group XX is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.

The special technical features of Group XXI is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition enhances Dkk binding to HBM.

The special technical features of Group XXII is considered a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of Dkk.

The special technical features of Group XXIII is considered a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of LRP5.

The special technical features of Group XXIV is considered a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of LRP6.

The special technical features of Group XXV is considered a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of HBM.

The special technical features of Group XXVI is considered a method of screening for a compound which modulates the interaction of DKK with LRP5.

The special technical features of Group XXVII is considered a method of screening for a compound which modulates the interaction of DKK with LRP6.

The special technical features of Group XXVIII is considered a method of screening for a compound which modulates the interaction of DKK with HBM.

The special technical features of Group XXIX is considered a method of screening for a compound which modulates the interaction of Dkk with a Dkk interacting proteins.

The special technical features of Group XXX is considered a composition comprising a LRP5 and a pharmaceutical acceptable carrier thereof.

The special technical features of Group XXXI is considered a composition comprising a LRP6 and a pharmaceutical acceptable carrier thereof.

The special technical features of Group XXXII is considered a composition comprising a HBM and a pharmaceutical acceptable carrier thereof.

The special technical features of Group XXXIII is considered a method for identifying compound which modulate Dkk and LRP5 interaction.

The special technical features of Group XXXIV is considered a method for identifying compound which modulate Dkk and LRP6 interaction.

The special technical features of Group XXXV is considered a method for identifying compound which modulate Dkk and HBM interaction.

The special technical features of Group XXXVI is considered a method of identifying binding partners for a Dkk protein.

The special technical features of Group XXXVII is considered a nucleic acid and a vector encoding a Dkk interacting protein.

The special technical features of Group XXXVIII is considered a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a LRP5.

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The special technical features of Group XXXIX is considered a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a LRP6.

The special technical features of Group XL is considered a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a HBM.

The special technical features of Group XLI is considered a transgenic animal.

The special technical features of Group XLII is considered a method for identifying potential compound which modulate Dkk activity.

The special technical features of Group XLIII - LXII is considered one specific peptide aptamer of one specific SEQ ID NOs : 171-88; 189-192.

The special technical features of Group LXIII- LXXIX is considered an antibody which specifically recognizes and binds to specific peptides of SEQ ID NOs : 110-127.

The special technical features of Group LXXX is considered a method of identifying Dkk interacting protein which modulate the interaction of Dkk with Wnt signaling pathway.

The special technical features of Group LXXXI is considered a method for identifying Dkk interacting proteins.

The special technical features of Group LXXXII is considered a method for identifying compounds which modulate Dkk and LRP5 interaction.

The special technical features of Group LXXXIII is considered a method for identifying compounds which modulate Dkk and LRP6 interaction.

The special technical features of Group LXXXIV is considered a method for identifying compounds which modulate Dkk and HBM interaction.

The special technical features of Group LXXXV is considered a method for identifying compound which modulate the interaction of Dkk with Wnt signaling pathway.

The special technical features of Group LXXXVI is considered a method of testing compounds that modulate Dkk-mediated activity in a mammal.

The special technical features of Group LXXXVII is considered a method of screening for compound or composition which modulate the interaction of Dkk and Dkk interacting protein.

The special technical features of Group LXXXVIII-CIX. is considered an antibody which recognizes and binds to one specific SEQ ID NOs: 171-192.

Accordingly, Groups I-CIX are not so linked by the same or corresponding special technical feature within meaning of PCT Rule 13.2 so as to form a single general inventive concept.

Continuation of B. FIELDS SEARCHED Item 3:

INTERNATIONAL SEARCH REPORT

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Biosis, CAPLUS, SciSearch, Medline, EMBASE, WEST, USPATFULL, PCTFULL
search terms; Allen K; Anisowicz,A; Bhat,B; Damagnez,V, Robinson,J;Yaworsky,P; DKK, Dkk1, LRP5, SEQ ID NO:28, protein
OST262; osteoporosis.